



UNICA

UNIVERSITÀ
DEGLI STUDI
DI CAGLIARI



Università di Cagliari

UNICA IRIS Institutional Research Information System

This is the Author's [*accepted*] manuscript version of the following contribution: (author name, title, publisher, ecc)

Giglio, Sabrina

Evidence for a Pathogenic Role of CSMD1 in Childhood Apraxia of Speech

NEUROPEDIATRICS

The publisher's version is available at: cod.

DOI: [10.1055/s-0043-1771033](https://doi.org/10.1055/s-0043-1771033)

When citing, please refer to the published version.

Accepted Manuscript (post-print con embargo)

This full text was downloaded from UNICA IRIS <https://iris.unica.it/>

Evidence for a Pathogenic Role of *CSMD1* in Childhood Apraxia of Speech

Daniela Formicola^{Q21,#} Irina Podda² Marilena Pantaleo³ Elena Andreucci³  Diego Lopergolo¹
Sabrina Giglio⁴ Filippo Maria Santorelli^{1,5} Anna Maria Chilosi⁵

Q2

¹ Department of Neurobiology and Molecular Medicine, IRCCS Fondazione Stella Maris, Pisa, Italy

² Parole al Centro Studio di Logopedia, Genoa, Italy

³ Medical Genetics Unit, Meyer Children's University Hospital, Florence, Italy

⁴ Department of Medical Sciences and Public Health, Medical Genetics Unit, University of Cagliari, Cagliari, Italy

⁵ Department of Developmental Neuroscience, IRCCS "Stella Maris Foundation" Scientific Institute, Pisa, Italy

Address for correspondence Anna Chilosi/Filippo M. Santorelli,

^{Q3}Molecular Medicine, IRCCS Stella Maris, Via dei Giacinti 2- 56128 Calambrone, Pisa, Italy

(e-mail: anna.chilosi@fsm.unipi.it; /filippo3364@gmail.com).

Q3

Neuropediatrics 2023;00:1–5.

Abstract

Keywords

- ▶ Childhood apraxia of speech
- ▶ language impairment
- ▶ *CSMD1*
- ▶ array-CGH
- ▶ WES

Childhood apraxia of speech (CAS) is a pediatric motor speech disorder. The genetic etiology of this complex neurological condition is not yet well understood, although some genes have been linked to it.

We describe the case of a boy with a severe and persistent motor speech disorder, consistent with CAS, and a coexisting language impairment.

Whole exome sequencing in our case revealed a *de novo* and splicing mutation in the *CSMD1* gene.

Introduction

Childhood apraxia of speech (CAS) is a subtype of motor speech disorder defined by the American Speech-Language-Hearing Association¹ as “a neurological childhood (pediatric) speech sound disorder, in which the precision and consistency of movements underlying speech are impaired in the absence of neuromuscular deficits (e.g., abnormal reflexes, abnormal tone)”. According to the ASHA consensus, three speech features are characteristic of CAS: “Inconsistent errors on consonants and vowels in repeated productions of syllables or words”, (b) “Lengthened and disrupted coarticulatory transitions between sounds and syllables”, and (c) “Inappropriate prosody, especially in the realization of lexical or phrasal stress”. Other speech features variably found in CAS are: increased difficulty with longer words and phrases, slow speech and diadochokinetic rate, intrusive sounds,

voicing errors, difficulty achieving initial articulatory configurations, scanned speech, and lexical stress assignment errors.² CAS may co-occur with persistent language and learning disorders,³ resulting in effortful, poorly intelligible speech which negatively impacts the child's social communication in daily activities, peer interactions, and literacy. With regard to the etiology of the condition, increasing evidence suggests that CAS has a genetic basis.

We report the case of a boy with a severe and persistent motor speech disorder, consistent with CAS, and a co-occurring language impairment, in whom a novel *de novo*, probably pathogenic variant in the *CSMD1* gene was detected by WES.

Methods

Having received written parental informed consent, we performed genome-wide array-CGH in the proband and Whole Exome Sequencing analysis on the family trio. The study received local ethics committee approval.

Present address: UOC Medical Genetics, S. Camillo Hospital, Rome, Italy.

received
November 7, 2022
accepted after revision
June 14, 2023

© 2023, Thieme. All rights reserved.
Georg Thieme Verlag KG,
Rüdigerstraße 14,
70469 Stuttgart, Germany

DOI <https://doi.org/10.1055/s-0043-1771033>.
ISSN 0174-304X.

Array-CGH was performed as previously described to identify potential genomic rearrangements. The hg19 human genome build GRCh37 was used as the consensus reference sequence. Data analyses were performed using Agilent CytoGenomics V.2.5.8.1. (Agilent, Santa Clara, CA).

Copy number variants (CNVs) were classified as “causative” or “non-causative” according to the American College of Medical Genetics and Genomics (ACMG) guidelines. Segregation analyses in parental DNA were performed by quantitative real-time polymerase chain reaction (qPCR).

We performed WES using NimbleGen SeqCap EZ Exome v3 capture kits (Roche, Italy) of family trio by assuming de novo, autosomal recessive, and X-linked mode of inheritance and by standard filtering methodologies outlined elsewhere.⁴

Variants were prioritized according to the following criteria:

- (1) not present in gnomAD or present with a mean allelic frequency <0.05%;
- (2) predicted as damaging with Combined Annotation-Dependent Depletion score ≥ 20 ;
- (3) (for splicing variants) predicted to affect splicing by AdaBoost and Random Forest software (we selected variants with a score ≥ 0.6);
- (4) classification as pathogenic, likely pathogenic, or of uncertain significance according to ACMG guidelines.

Results

This patient has been followed by our institute (IRCSS Fondazione Stella Maris) for his language impairment from the age of 4 years, when he was administered a thorough

speech and language examination³ according to the standard of care in our hospital.

The boy was born at term by C-section due to breech presentation after an uneventful pregnancy in which his mother was treated with levothyroxine because of hypothyroidism. His weight at birth was 3690 g.; his neonatal and perinatal history was unremarkable. His family history was unremarkable for speech, language and cognitive disorders. Motor development was normal, with independent walking achieved at 12 months, whilst oral language acquisition was delayed from the early vocal stages: canonical babbling was scarce and showed little variation. Up to 30 months of age, the child produced only single vocalic sounds and some isolated syllables. His social and communicative skills were normal: he used several nonverbal strategies to communicate (signs, mimicking, eye contact). When first examined at the age of 4 years and 5 months, neurological examination detected only mild limb hypotonia. Routine blood tests were also unremarkable. We observed normal nonverbal cognitive skills (WPPSI-III PIQ = 107). The child had no orofacial abnormalities and his hearing was within the normal range. Brain MRI revealed a small area of signal hyperintensity on T1-weighted images, involving the deep white matter adjacent to the superior external angle of the left ventricle. This punctiform non-specific alteration can be interpreted either as a prominent vascular space or as a small gliotic area. No other structural brain abnormalities were detected.

At 5 years of age his speech was markedly impaired and showed all the three ASHA features and several of the ten speech characteristics listed in Strand's 10-point checklist² (► **Table 1**). His phonetic inventory included 12/21 consonant

Table 1 CAS speech features shown by the patient

		Patient 1 S.Q.
ASHA: 3 speech features	1) Inconsistent errors on consonants and vowels in repeated productions of syllables or words.	+
	2) Lengthened and disrupted co-articulatory transitions between sounds and syllables.	+
	3) Inappropriate prosody, especially in the realization of lexical or phrasal stress.	+
Strand's 10-point checklist		
	1. Vowel or consonant errors including distorted substitutions.	+
	2. Intrusive schwa.	–
	3. Voicing errors.	–
	4. Lexical stress errors or equal stress.	–
	5. Syllable segregation.	+
	6. Slow rate.	–
	7. Difficulty achieving initial articulatory configurations and transitions into vowels.	+
	8. Slow DDK rate.	+
	9. Groping (articulatory searching prior to phonating).	+
	10. Increased difficulty with longer or more phonetically complex words.	+

ASHA: American Speech-Language-Hearing Association; CAS: childhood apraxia of speech. + indicates the presence of a speech feature.

sounds. His speech production was characterized by a 57% rate of inaccurate repetitions of two-syllable nonwords over successive attempts at the same target, and 21% of the errors were inconsistent. The diadochokinetic rate of three-syllable nonword sequences was markedly inaccurate, inconsistent, and slow for the child's age (11 repetitions over 20 seconds). In repeated production of real two- and three-syllable words, 55% of productions were inaccurate and 19% of the errors were inconsistent. Inaccuracy and inconsistency increased in longer utterances and the child's verbal production became poorly intelligible, scanned, and slow. On the Verbal Motor Production Assessment for Children⁵ his verbal motor skills were globally impaired.

The child's receptive lexicon was within the normal limits, as were his receptive grammar skills.⁶ His expressive lexicon was normal, whilst his expressive grammar was impaired. Analysis of semi-spontaneous language samples showed that his expressive language was below the expected level for his age and corresponded to Grid of Analysis of Spontaneous Speech (GASS) level 4,^{7–8} characterized by poor control of grammar rules in complex phrases, with frequent omission of free morphemes and of subordinate clause functions. GASS level 4 is usually achieved by typically developing children by 32 months of age (a sample of his speech is presented online as **►Supplementary Material 1**).

Array-CGH analysis did not show any potentially relevant CNVs. Conversely, WES analysis identified the rare *de novo* variant c.9280 + 1G > A in *CSMD1* (**►Figure 1**).

This variant was not present in the gnomAD database, nor was it found in in-house WES data of >3500 individuals with unrelated genetic diseases. The variant c.9280 + 1G > A is located in canonical donor splicing site and different algorithms predict the effect of sequence changes on RNA splicing (SpliceAI Donor Loss = 1.0, Ada_score = 0.999915711 and

rf_score = 0.916); it is expected that exon 59 skipping does not disrupt reading frame. The role of exon 59 on *CSMD1* protein function is unknown. Currently, pathogenic missense variants are not known in exon 59 and a single missense variant in Clinvar database was reported in Sushi 24 domain (amino acids 3035–3094 on protein Q96PZ7). However, exon 59 is present in biologically-relevant transcript and exon 59 skipping removes about <10% of *CSMD1* protein. This mutation, meeting the PVS1-PM2 -PS2 criteria of the ACMG, was considered “likely pathogenic”.

Discussion

These findings add a potentially new gene to the list of etiological factors in CAS and language impairment.

We found a likely pathogenic *de novo* splice-site variant in *CSMD1*. *CSMD1* is classified as an autism spectrum disorder candidate gene in the SFARI (Simons Foundation Autism Research Initiative) database. The gene is expressed at intermediate levels in the brain, including the cerebellum, substantia nigra, and hippocampus, as well as in the fetal brain. It lies at 8p23.2 and *CSMD1* genetic variants have been shown to be associated with autism.^{9,10}

CSMD1 (OMIM ID 608397; 8p23.2) and its paralog *CSMD2* (OMIM ID 608398; 1p35.1) are members of the CUB and Sushi multiple domain protein family, and are described as regulators of complement activation and inflammation in the developing central nervous system.¹¹ Complement activation is gaining much attention because it is essential for synaptic plasticity and has recently been implicated in several brain-related conditions such as schizophrenia,¹¹ a condition characterized by poor and disorganized speech, and impairment of cognitive domains, including attention, memory, learning, and executive functions.

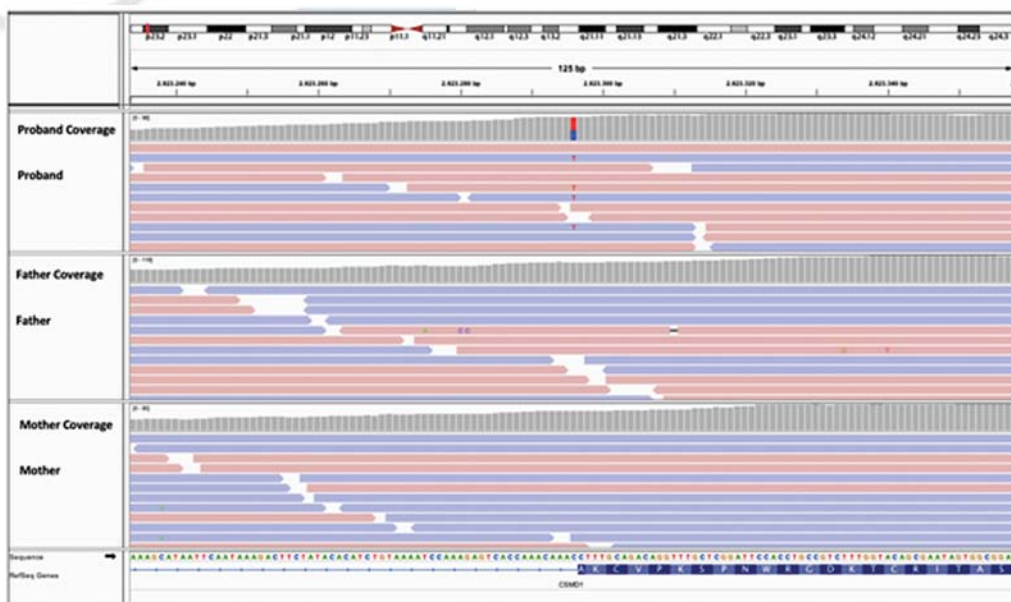


Fig. 1 IGV screenshot of WES alignments for the proband (top track), father, (middle track), and mother (bottom track). Each track comprises two parts: a histogram of the read depth and the reads as aligned to the reference sequence. Reads are colored according to the aligned strand (red = forward strand; blue = reverse strand).

A study on *CSMD1* knockout mice suggested that depletion of *CSMD1* expression is linked to abnormal emotional/affective responses, hyperactivity, and increased anxiety-related response.¹² Koiliari and colleagues highlighted the involvement of the *CSMD1* gene in cognitive disorders, reporting associations of *CSMD1* variants with poorer neuro-cognitive functioning, such as impaired general cognitive abilities, strategy formation, planning, set shifting, and episodic verbal memory.^{13,14}

Additionally, in a recent review by Akyuz and Bell¹⁵ the role of *CSMD1* is described in neurodevelopmental disorders and in some pathological conditions, such as Parkinson disease and schizophrenia. *CSMD1* variants are associated with immediate episodic memory and cognitive functions¹⁴ and *CSMD1* expression is high in the amygdala, thus affecting social behavior.¹⁶ Moreover, the fact that during early development, neuronal connections between the amygdala and the language processing centers are strengthened in the brain, highlights the role of the amygdala in language learning processes and also the probably potential involvement of *CSMD1* in speech impairment.

Our child meets the clinical criteria for a diagnosis of CAS. We performed in-depth language phenotyping, which helped us to prioritize variants in WES. A similar approach might allow future larger studies to define additional monogenic forms of CAS and/or language impairment. Interestingly, scrutiny of the pertinent literature revealed a description of a genomic duplication at the 8p23.1 locus encompassing the *CSMD1* gene in a patient who showed speech delay and learning difficulties.¹⁷ Even more interestingly, upon revision of our own database of 2715 children with neurodevelopmental disorders and speech impairment, we found an additional case with normal WES and a similar genomic rearrangement encompassing *CSMD1* and with some features resembling CAS (data not shown). Moreover, the DECIPHER database, too, includes patients (ID_394741, ID_287658, ID_254189, ID_379314) with speech delay, language impairment, and intellectual disability, who have 8p23.2 duplications involving *CSMD1*. Thus, forms related to this gene may be more common than originally believed.

In summary, our findings show that *CSMD1* may play a role in language development, and suggest it may be involved in the mechanism underlying CAS phenotype. These findings add to existing knowledge on the diagnostic genetic markers of CAS, and contribute to efforts to ensure that pediatric patients with speech disorders receive the most appropriate treatment options.

Author Contributions

AC, FMS, DF and IP drafted the manuscript. DF performed wet-lab (sample processing steps) and WES data analysis. MP performed array-CGH. AC and IP collected and analyzed clinical data. DL performed genetic counseling. AC,

FMS, DF, SG and EA critically reviewed the manuscript. AC and DF designed the study. FMS reviewed the study design. All authors have read and approved the final version of the manuscript.

Funding

This work was supported in part by funding from the Italian Ministry of Health (RF2016-02361560 to A.C.; RC 5 × 1000 to FMS and AC).

Conflict of Interest^{Q4}

None declared.

Acknowledgments

We would like to acknowledge the children and families who participated in this study. We would like to thank Dr. Paola Cipriani for her critical suggestions on the manuscript. We would also like to acknowledge Dr. Elia Dirupo at AOU Meyer for his bioinformatic support.

References^{Q5}

- 1 ASHA. American Speech-Language-Hearing Association - ASHA, Childhood apraxia of speech [Technical Report]. 2007; Retrieved from <https://www.asha.org/practice-portal/clinical-topics/childhood-apraxia-of-speech/>
- 2 Shriberg LD, Potter NL, Strand EA. Prevalence and phenotype of childhood apraxia of speech in youth with galactosemia. *J Speech Lang Hear Res* 2011;54(02):487–519
- 3 Chilosi AM, Podda I, Ricca I, et al. Differences and commonalities in children with childhood apraxia of speech and comorbid neurodevelopmental disorders: a multidimensional perspective. *J Pers Med* 2022;12(02):313
- 4 Provenzano A, La Barbera A, Scagnet M, et al. Chiari 1 malformation and exome sequencing in 51 trios: the emerging role of rare missense variants in chromatin-remodeling genes. *Hum Genet* 2021;140(04):625–647
- 5 Hayden DA, Square P. Verbal Motor Production Assessment for Children (VMPAC). San Antonio, TX: The Psychological Corporation; 1999
- 6 Chilosi AM, Cipriani P. TCGB, Test di Comprensione Grammaticale per Bambini: D. Cerro Ed. Pisa; 2005
- 7 Chilosi AM, Pfanner L, Pecini C, et al. Which linguistic measures distinguish transient from persistent language problems in Late Talkers from 2 to 4 years? A study on Italian speaking children. *Res Dev Disabil* 2019;89:59–68
- 8 Cipriani P, Chilosi AM, Bottari P, Pfanner L. L'acquisizione della morfosintassi in italiano: fasi e processi. Padova: Unipress; 1993
- 9 Sanders SJ, Murtha MT, Gupta AR, et al. De novo mutations revealed by whole-exome sequencing are strongly associated with autism. *Nature* 2012;485(7397):237–241
- 10 Guo H, Wang T, Wu H, et al. Inherited and multiple de novo mutations in autism/developmental delay risk genes suggest a multifactorial model. *Mol Autism* 2018;9:64
- 11 Kraus DM, Elliott GS, Chute H, et al. *CSMD1* is a novel multiple domain complement-regulatory protein highly expressed in the central nervous system and epithelial tissues. *J Immunol* 2006; 176(07):4419–4430

- 12 Steen VM, Nepal C, Erslund KM, et al. Neuropsychological deficits in mice depleted of the schizophrenia susceptibility gene *CSMD1*. *PLoS One* 2013;8(11):e79501
- 13 Koiliari E, Roussos P, Pasparakis E, et al. The *CSMD1* genome-wide associated schizophrenia risk variant rs10503253 affects general cognitive ability and executive function in healthy males. *Schizophr Res* 2014;154(1-3):42–47
- 14 Athanasiu L, Giddaluru S, Fernandes C, et al. A genetic association study of *CSMD1* and *CSMD2* with cognitive function. *Brain Behav Immun* 2017;61:209–216
- 15 Ermis Akyuz E, Bell SM. The Diverse Role of CUB and Sushi Multiple Domains 1 (*CSMD1*) in Human Diseases. *Genes (Basel)* 2022;13(12):2332
- 16 Bickart K, Napolioni V, et al. Genetic variation in *CSMD1* affects amygdala connectivity and prosocial behavior. *bioRxiv* 2020
- 17 Glancy M, Barnicoat A, Vijeratnam R, et al. Transmitted duplication of 8p23.1–8p23.2 associated with speech delay, autism and learning difficulties. *Eur J Hum Genet* 2009;17(01):37–43



THIEME

Author Query Form (NEP/1020223346SC)

Special Instructions: Author please write responses to queries directly on proofs and then return back.

- Q1: AU: Please check whether the suggested running head is acceptable. ok
- Q2: AU: Please confirm that given names (red), middle names (black) and surnames (green) have been identified correctly. Author names in bibliographic citations and online will appear as: Formicola D, Podda I, Pantaleo M, Andreucci E, Lopergolo D, Giglio S, Santorelli FM, Chilosi AM. Evidence for a Pathogenic Role of CSMD1 in Childhood Apraxia of Speech. Please confirm if this is correct. all correct but no middle name for author Chilosi
- Q3: AU: Please provide degree of corresponding author. Both are MD
- Q4: AU: Please provide the signed ICMJE COI forms available at <http://www.icmje.org/conflicts-of-interest/> for all the authors.
- Q5: AU/PM: References 16 and 14 were identical, therefore 16 was deleted and renumbered the list and in-text citations accordingly. ok correct ok see attached file



THIEME