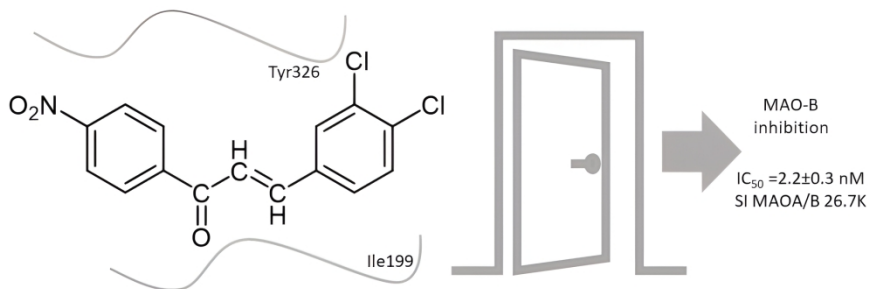


This document is confidential and is proprietary to the American Chemical Society and its authors. Do not copy or disclose without written permission. If you have received this item in error, notify the sender and delete all copies.

Exploring the 1-(4-nitrophenyl) -3-arylprop-2-en-1-one scaffold for the selective inhibition of monoamine oxidase B

Journal:	<i>ACS Medicinal Chemistry Letters</i>
Manuscript ID	ml-2024-002463.R2
Manuscript Type:	Letter
Date Submitted by the Author:	n/a
Complete List of Authors:	<p>Meleddu, Rita; University of Cagliari Department of Life Sciences and Environment, Life and Environmental Science FAIS, ANTONELLA; University of Cagliari, Department of Life and Environmental Sciences Era, Benedetta; University of Cagliari, Sciences Applied to Biosystems Floris, Sonia; University of Cagliari Department of Life Sciences and Environment, Life and Environmental Science Distinto, Simona; Università degli Studi di Cagliari, Dipartimento di Scienze della Vita e dell'Ambiente Lupia, Antonio; University of Cagliari, Cottiglia, Filippo; Università degli Studi di Cagliari, Life and Environmental Sciences Onali, Alessia; University of Cagliari Department of Life Sciences and Environment, Life and Environmental Science Sanna, Erica; University of Cagliari Department of Life Sciences and Environment, Life and Environmental Science Secci, Daniela; University of Ljubljana Faculty of Pharmacy Atzeni, Giulia; University of Cagliari Department of Life Sciences and Environment, Life and Environmental Science Demuru, Laura; University of Cagliari Department of Life Sciences and Environment, Life and Environmental Science Caboni, Pierluigi; Università degli Studi di Cagliari, Life and Environmental Sciences Valenti, Donatella; University of Cagliari Department of Life Sciences and Environment, Life and Environmental Science Maccioni, Elias; University of Cagliari Department of Life Sciences and Environment, Life and Environmental Science</p>

SCHOLARONE™
Manuscripts



Graphical Abstract

216x121mm (300 x 300 DPI)

Exploring the 1-(4-nitrophenyl)-3-arylprop-2-en-1-one scaffold for the selective inhibition of monoamine oxidase B

Rita Meleddu†‡, Antonella Fais†‡*, Benedetta Era†, Sonia Floris†, Simona Distinto†, Antonio Lupia†¶, Filippo Cottiglia†, Alessia Onali†, Erica Sanna†, Daniela Secci†#, Giulia Atzeni†, Laura Demuru†, Pierluigi Caboni†, Donatella Valenti†, and Elias Maccioni†*.

†Department of Life and Environmental Sciences, University of Cagliari, University Campus, S.P. 8 km 0.700, 09042 Monserrato, Italy

¶Net4Science Srl, University "Magna Græcia", Campus Salvatore Venuta, Viale Europa, 88100, Catanzaro, Italy

ABSTRACT A small library of 1-(4-nitrophenyl)-3-arylprop-2-en-1-one derivatives was synthesized to identify new human monoamine oxidase B selective inhibitors. Their inhibitory activity toward MAO-A and MAO-B isoforms was evaluated to determine potency and selectivity. All newly synthesized compounds were nanomolar inhibitors of the B isoform with IC_{50} concentrations ranging from 120 to 2.2 nM. Conversely, their activity towards the A isozyme was only observed at micromolar concentrations. Our results bear out the hypothesis that the 1,3-diarylpropenone scaffold could represent a valuable starting point for designing efficient and selective MAO B inhibitors.

KEYWORDS MAO-B selective inhibitors; 1,3-diarylpropenone; oxidative stress; neurodegeneration.

Monoamine oxidases (MAOs) are ubiquitous enzymes that catalyze the oxidative deamination of endogenous and exogenous amines^{1, 2}. Dopamine (DA), norepinephrine (NE), serotonin (5HT), and phenylethylamine are common substrates. Hence, MAOs play an essential role in the metabolism of monoamine neurotransmitters and have been extensively investigated for their role in several neurological disorders³⁻¹⁰. The mammalian family of MAO comprises two isozymes, MAO-A and MAO-B. These two isoforms differ in substrates and inhibitors selectivity, being 5HT and NE preferential substrates of MAO-A, while MAO-B preferentially deaminates PEA and benzylamine. Moreover, although MAO-A and MAO-B exhibit a high sequence similarity, they present substantial differences in the shape and volume of the catalytic site, allowing for the design of selective inhibitors^{11, 12}. MAO-A presents a single cavity with a volume of about $\approx 550 \text{ \AA}^3$, while MAO-B has three functional domains named an "entrance cavity", "substrate pocket", and "aromatic cage" (Y398 and Y435). Altogether, these form the active site with a volume of approximately $\approx 700 \text{ \AA}^3$ ¹³⁻¹⁵. Not surprisingly, according to this structural information and to selectively target one or the other isoform, several selective inhibitors have been identified and investigated for their therapeutic applications for the treatment of depression and neurological diseases^{5-7, 16, 17}. Noteworthy, the oxidative deamination catalyzed by MAOs produces hydrogen peroxide and other reactive species that are involved in neurodegeneration, such as in Alzheimer's disease (AD), Parkinson's disease (PD) and other neurodegenerative diseases (NDDs)¹⁸. Therefore, MAO inhibition could exert an indirect *in vivo* antioxidant activity at neuronal level¹⁹⁻²¹. In addition, overexpression of MAO-B and DA metabolism, as well as the formation of α -synuclein (α -Syn) aggregates, are observed in PD²². It has also been reported that α -Syn selectively binds MAO-B and increases its catalytic activity²³. More recently, the key role of MAO-B in PD was further evidenced in animal models by Nakamura and co-workers²⁴. Indeed, they observed that either inhibition or knock-down of MAO-B can reduce α -Syn intracellular accumulation. In addition, they observed that selegiline delayed the striatal formation of α -Syn

aggregates, and mitigated loss of nigrostriatal dopaminergic neurons²⁴. According to these findings, MAO-B selective inhibitors might not only increase the level of DA in nigrostriatal dopaminergic neurons but also exert a neuroprotective effect by reducing α -Syn aggregation and oxidative stress. Therefore, MAO-B inhibitors might be useful for ameliorating motor and non-motor symptoms, reducing "OFF" time, and preventing or delaying disease progression²⁵. Our research group has already focused on the design of selective MAO-B inhibitors, and we have identified potent and selective agents through scaffold optimization from 3,5-diaryl dihydropyrazoles to 3,5-diaryl dihydroisoxazoles²⁶⁻³⁸. Based on our previous findings (Figure 1) and with the aim to further explore the structural requirements for the selective inhibition of MAO-B, we have synthesized a small library of 1-(4-nitrophenyl)-3-arylprop-2-en-1-one derivatives **3 a-g** and evaluated their activity and selectivity toward the two isozymes human MAO-A and MAO-B (*h*MAO-A and *h*MAO-B).

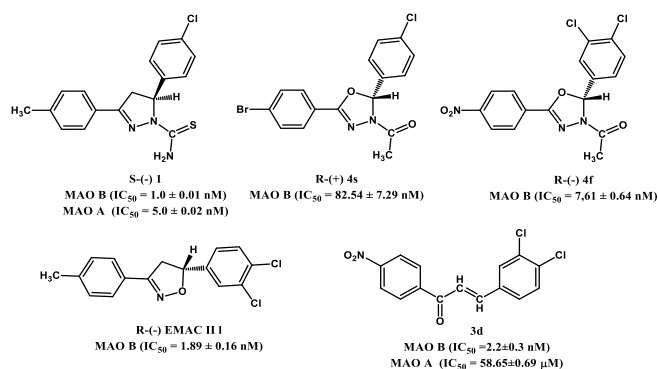
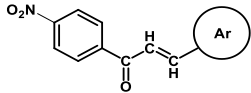


Figure 1. Comparison of previously reported MAO B inhibitors (S-(-) 1, R-(+) 4s, R-(-) 4f, and R-(-) EMAC II 1), named as in the original paper, and compound 3d.^{26, 27, 31, 38}

3 a-g derivatives could be considered as structurally simplified bioisosteres of previously investigated 3-acetyl-2,5-diaryl-2,3-dihydro-1,3,4-oxadiazoles^{27, 31} and dihydro-isoxazole nucleus²⁶. Our previous findings evidenced that the construction of an appropriately substituted five terms heterocycle, bearing two aryl substituents, could efficiently and selectively inhibit MAO-B isozyme. In this respect, and to further explore scaffolds and ligand geometries we have replaced the heterocyclic cores with a propenone moiety in which the two aryl substituents bind the position 1 and 3. As in the previously studied derivatives, the presence of a 4-nitrophenyl moiety remained a constant structural feature throughout the new derivatives series, to allow an easier comparison. Substituted phenyl moieties differing for steric hindrance and electronic distribution were introduced in position 3 of the propenone linker. Nevertheless, as a future development, replacements of the potentially toxic nitrophenyl group with bioisosteric moieties such as phenyl-carboxylic acid, benzo-oxadiazole, and dihydro-benzofuranone will be explored.

Thus, compounds **3 a-g** were synthesized, characterized, and submitted to biochemical evaluation to assess their activity and selectivity towards the MAO-A and MAO-B isozymes. All derivatives are MAO-B isozyme nanomolar inhibitors with IC₅₀ values ranging from 120 to 2.2 nM. Contrarywise, their activity toward the MAO-A isoform is only observed at micromolar concentrations with IC₅₀ values varying from 89 to 2.7 μM (Table 1).

Table 1. Inhibition data towards hMAO-A and -B of compounds 3 a-g



Comp.	Ar	hMAO-A (IC ₅₀ μM)	hMAO-B (IC ₅₀ nM)	S.I.*
3a	4-Cl-Ph	52±0.6 ^a	10±0.02 ^a	5,200
3b	2-CH ₃ -Ph	89±0.9 ^b	120±7.9 ^b	740
3c	4-F-Ph	7.8±0.12 ^c	48±3.8 ^c	160
3d	3,4-Cl-Ph	59±0.7 ^d	2.2±0.3 ^a	27,000
3e	4-OCH ₃ -Ph	27±4.7 ^e	93±5.5 ^d	290
3f	2-Naph	13±2.6 ^{c,f}	21±3.4 ^{a,c}	620
3g	4-CH ₃ -Ph	2.7±1.9 ^c	13±2.5 ^{a,f}	210
	Selegiline	47±1.1 ^a	30±1.8 ^c	1,600

* Selectivity index (S.I.) = IC₅₀(hMAO-A)/IC₅₀(hMAO-B). Each value is the mean ± SD of three independent measurements (n = 3). Different letters within the same column indicate statistically significant differences between compounds (p < 0.0001); one-way ANOVA followed by Tukey's Multiple Comparisons Test).

According to these results, we can affirm that all derivatives are selective MAO-B inhibitors with selectivity indexes (S.I. [IC₅₀ (hMAO-A)] / [IC₅₀ (hMAO-B)] ranging from 160, in the case of compound **3c**, to 27,000 for compound **3d**. The latter compound is the most potent and selective among all the tested derivatives indicating that the presence of a 3,4-dichlorophenyl moiety is highly beneficial for the potency similarly to what was observed for the heterocyclic core based analogues²⁷. Conversely, the introduction of a 4-methoxyphenyl group led to a 42 folds reduction of the

activity compared with the most potent derivative **3d**. Thus, the introduction of a strong electron donating group is not well tolerated in position 4 of the phenyl ring. By introducing a single chlorine atom in the same position, as for compound **3a**, only a moderate reduction of the potency (less than 5 folds) compared with **3d** was observed, indicating that this substitution is well tolerated. Moreover, the isosteric substitution of the chlorine atom with a methyl group in position 4 of the phenyl ring (**3g**) is also well tolerated, leading to an IC₅₀ value statistically equal to that of the 4-chlorophenyl substituted derivative. According to this information, we can assume that the electronic density in the phenyl ring is most likely not the main actor for the activity and selectivity. On the other hand, the introduction of a non-polar substituent of an appropriate volume (Cl or CH₃), regardless of its electronic effect, in position 4 appeared as more determinant for the activity. Most probably, these substituents are optimal to interact with specific lipophilic residues of the enzyme. Moreover, we observed that the introduction of a proximal second non-polar substituent, as in the case of the 3,4-dichloro substituted compound **3d**, was able to maximize the ligand efficacy. This hypothesis is further corroborated by the fact that when a 4-fluorophenyl (**3c**) moiety is introduced a reduction of 22 folds of the potency was observed, with respect to **3d**. Probably, the fluorine atom is not sufficiently bulky to correctly interact with the enzyme lipophilic region, resulting in a decrease of ligand potency. The introduction of a methyl moiety in the position 2 of the phenyl ring (**3b**) leads to a reduction of the potency, with respect to the most potent compound of about 55 folds, probably due to its intramolecular steric effect that might limit the conformational freedom of the ligand. Surprisingly, the introduction of the bulky 2-naphthyl only led to a moderate reduction of the activity (less than 10 folds). All together this information indicates that both size and polarity of the substituents on the 3-aryl moiety can play a role in determining the activity of the compound. Indeed, the orientation of the aromatic moiety appeared to be relevant in determining the activity of these compounds, indicating that this portion must retain the freedom to correctly orientate within the binding pocket. However, considering the potential toxicity associated with the nitro group, QikProp software was employed to perform in silico ADMET³⁹ predictions to evaluate the drug-likeness of compound **3d**. The predicted values are provided in the Supplementary Information (SI, Table S3 and S4). All values fall within acceptable ranges, except for the electron affinity, which is borderline. Overall, the compound exhibits a favorable predicted drug-like profile. The predicted heart toxicity warrants further investigation but given the compound's IC₅₀ in the nanomolar range, this is not likely to create a significant issue. The compound also shows limited predicted ability to cross the blood-brain barrier (BBB). This should be considered during the optimization process, while an appropriate delivery formulation could mitigate this limitation.

Therefore, to further rationalize these structure activity observations, molecular docking and post-docking energy optimization experiments were carried out considering the most promising compound **3d**.

The docking protocol has been validated by running re-docking and cross-docking studies considering the following reversible ligand-MAO-B complexes available in the PDB⁴⁰: 1OJ9¹³, 2C67⁴¹, 1OJA⁴², 2BK3⁴³, 2V5Z⁴⁴, 2V60⁴⁴, 2V61⁴⁴, 6FW0⁴⁵.

Among all the examined protocols (see the methods part) the best was the QPDL-SP⁴⁶, likely due to the incorporation of QM treatment which helped achieve better RMSD. Indeed, the described

settings could reproduce the experimental binding modes of considered co-crystallized ligands with an RMSD value below 2.5 Å.

The QPLD⁴⁶ protocol is a very accurate docking method that combines the quantum mechanical (QM) approach in the binding region of the ligand and in the pocket binding, while the rest is treated according to the molecular mechanic's laws. The docking protocol validation allows relying qualitatively on the docking results and to analyze the ligand putative binding mode. After the molecular docking, the obtained complexes have been energy minimized allowing the residues of the cavity to adapt to the ligand simulating the induced fit effect. The resulting best binding mode is depicted in Figure 2.

It can be observed that the ligand shape is optimal to occupy both the entrance and the catalytic cavity. Ile199 and Tyr326 are considered “gatekeepers” of the MAO-B catalytic cavity”, while the MAO-A isoform does not have an entrance cavity. Therefore, this could explain the preference for MAO-B binding affinity and the selectivity. Concerning the complex **3d**-MAO-B stabilizing interactions, a number of van der Waals contacts can be observed with the aromatic cage residues Tyr435, Tyr398, Phe343, and Tyr188, as well as several hydrophobic residues in the entrance cavity Ile198, Ile199, Tyr326, Leu164, Leu171, Phe168. The MAO B binding site was analyzed using SiteMap,⁴⁷ which not only assesses the suitability of a site to host a ligand but also identifies the areas where specific moieties preferentially interact. The

SiteMap study highlighted a large hydrophobic area occupied by the diaryl-propenone derivative (Figure 3). A further molecular interaction fields (MIFs) analysis was considered. After removing the inhibitor, the interaction capabilities of the MAO-B active site were investigated by means of Cl probe as implemented in the program GRID^{31, 48}. The resulting isocontour maps were inspected taking into account the docking reported pose of the **3d** inhibitor into the hMAO-B binding. MIFs indicated strong interaction between the target and organic chlorine (Figure S1). Nevertheless, the presence of well-defined areas where hydrogen bonding could be established with the ligand was evidenced. Thus, the introduction of hydrogen bond acceptors and donors in the ligand structure, pointing to these specific areas, may result in increased activity and specificity toward the target MAO-B isozyme. All together these data indicate that 1-(4-nitrophenyl)-3-arylprop-2-en-1-ones may represent a valid structural simplification of previously reported heterocyclic MAO-B inhibitors. Indeed, all the synthesized compounds were selective nanomolar inhibitors of the MAO-B enzyme, while their potency on the MAO-A isozyme was significantly lower. This selectivity can be attributed to differences in the binding sites, including shape, size^{13, 49} and calculated energetically favored maps (Figure S2). Consequently, the binding energy contributions reflect these differences and explain the observed activity. For instance, we present the docking data for compound **3d** with both isoenzymes A and B (Table S5).

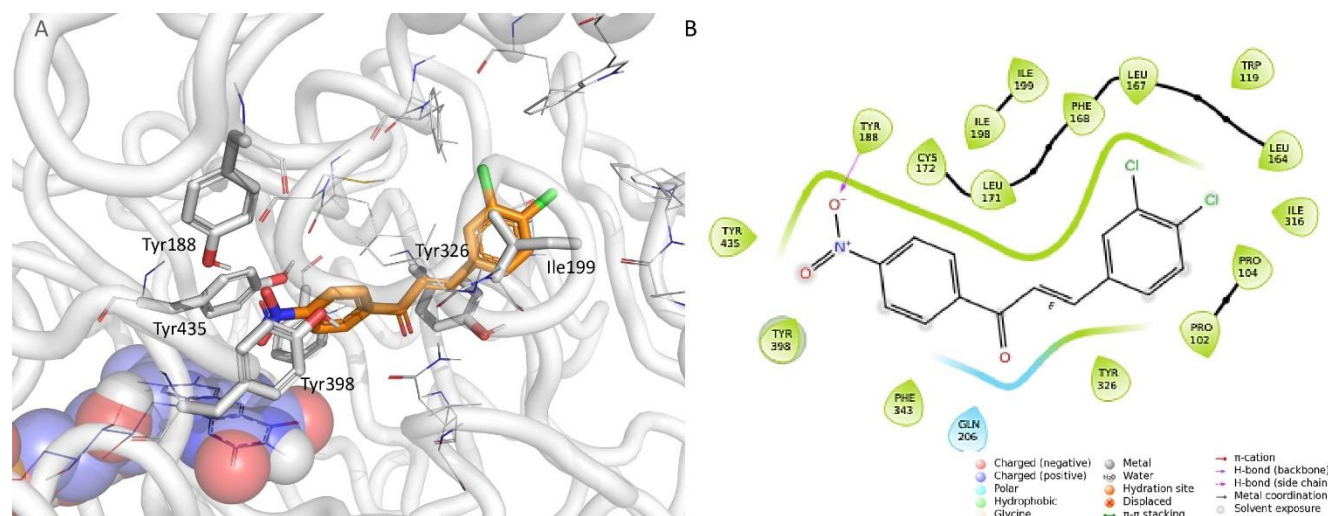


Figure 2. A) Representation of the molecular docking result of **3d** after subsequent complex energy minimization and B) 2D representation of the compound and interacting residues: in light green the hydrophobic, in blue the polar ones. In magenta arrow the hydrogen bond acceptor.

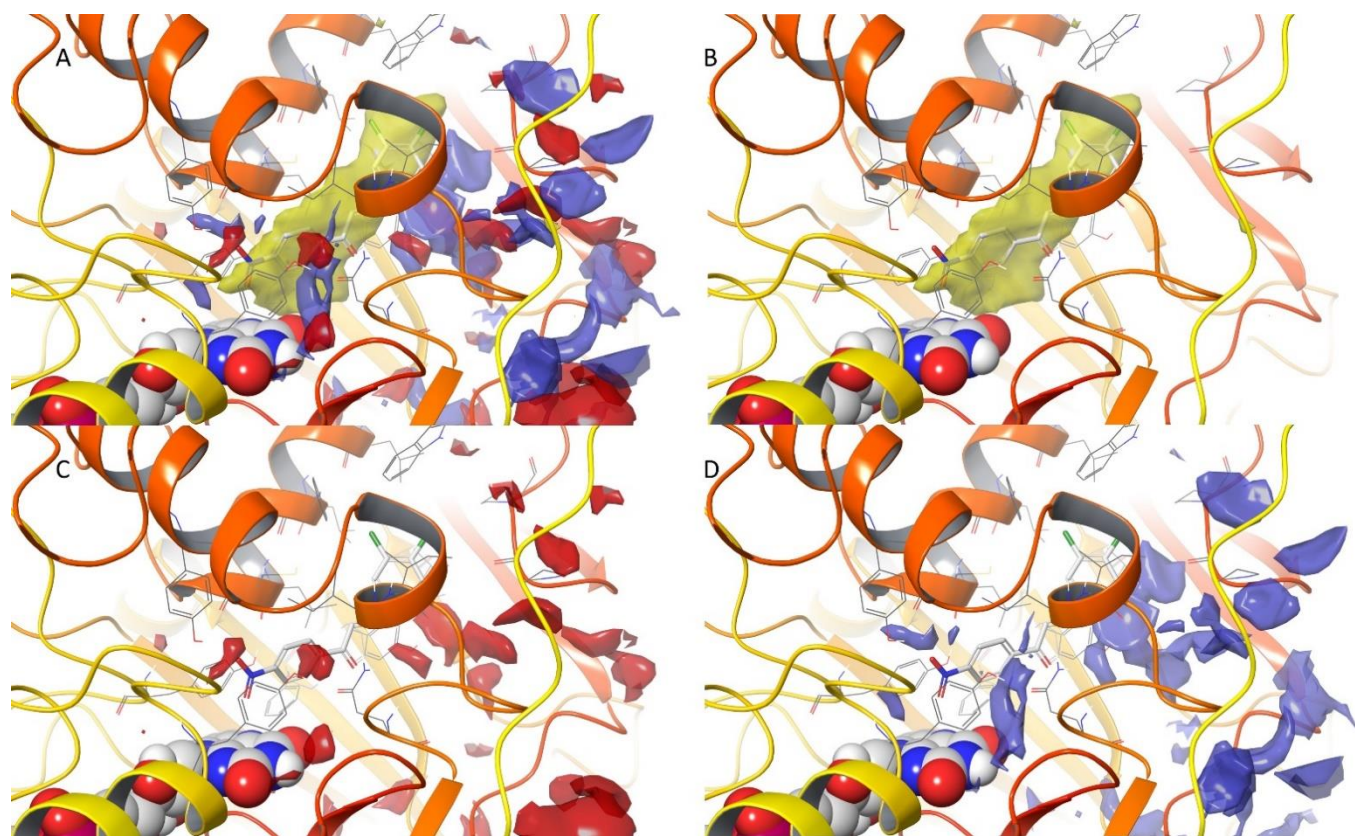


Figure 3. A) Visualization of binding site cavity favorable maps obtained with SiteMap: energetically favored maps are displayed all at once, then split in B, C, D. In yellow are shown the areas where the hydrophobic moieties are favored, in blue the hydrogen bond donors and in red the acceptors.

Both SARs and molecular modelling provided clear indications for further optimization of these derivatives that may positively influence the ability to inhibit the monoamine oxidase enzyme's B isoform selectively. Noteworthy, the new derivatives exhibited a structure-activity profile similar to the previously investigated dihydropyrazoles, dihydrooxadiazoles, and dihydroisoxazoles, suggesting potential further structural options regarding the spacer between the two aromatic moieties. Interestingly, although all compounds were potent MAO-B inhibitors, the relevant role of a 3,4-dichlorophenyl moiety at the 3-position of the propenone for inhibitory potency was pointed out. These data prompt us to further explore the structural requirements for the selective and potent inhibition of MAO-B. Concerning molecular modeling studies, the presence and nature of favorable energy maps will be considered for future structural modifications of this set of molecules.

ASSOCIATED CONTENT

Supporting Information

Methods Chemical physical properties of compounds, ^1H NMR, ^{13}C NMR, HRMS spectra, HPLC chromatograms, biological assay and molecular modeling details, and drug like properties prediction. The Supporting Information is available free of charge on the ACS Publications website.

AUTHOR INFORMATION

Corresponding Authors

***Antonella Fais** – Department of Life and Environmental Sciences, Cittadella Universitaria, University of Cagliari, Cagliari 09124, Italy; orcid.org/0000-0002-3009-4226; e-mail: fais@unica.it,

***Elias Maccioni** – Department of Life and Environmental Sciences, Cittadella Universitaria, University of Cagliari, Cagliari 09124, Italy; orcid.org/0000-0003-2175-2802; e-mail: elias.maccioni@unica.it.

Authors

Rita Meleddu – Department of Life and Environmental Sciences, Cittadella Universitaria, University of Cagliari, Cagliari 09124, Italy; orcid.org/0000-0003-1629-7454

Benedetta Era – Department of Life and Environmental Sciences, Cittadella Universitaria, University of Cagliari, Cagliari 09124, Italy; orcid.org/0000-0002-4154-9740

Sonia Floris – Department of Life and Environmental Sciences, Cittadella Universitaria, University of Cagliari, Cagliari 09124, Italy; orcid.org/0009-0007-3140-7048

Simona Distinto – Department of Life and Environmental Sciences, Cittadella Universitaria, University of Cagliari, Cagliari 09124, Italy; orcid.org/0000-0003-1620-6225

Antonio Lupia – Department of Life and Environmental Sciences, Cittadella Universitaria, University of Cagliari, Cagliari 09124, Italy; Net4Science S.r.l, Università degli Studi “Magna Graecia” di Catanzaro, Catanzaro 88100, Italy; orcid.org/0000-0003-0870-2376

Filippo Cottiglia – Department of Life and Environmental Sciences, Cittadella Universitaria, University of Cagliari, Cagliari 09124, Italy; orcid.org/0000-0003-4659-9808

Alessia Onali – Department of Life and Environmental Sciences, Cittadella Universitaria, University of Cagliari, Cagliari 09124, Italy; orcid.org/0009-0004-7540-3901

Erica Sanna – Department of Life and Environmental Sciences, Cittadella Universitaria, University of Cagliari, Cagliari 09124, Italy; orcid.org/0009-0001-4895-970X

Daniela Secci – Department of Life and Environmental Sciences, Cittadella Universitaria, University of Cagliari, Cagliari 09124, Italy; Present Address: Faculty of Pharmacy, University of Ljubiana, 1000 Ljubljana, Slovenia; orcid.org/0000-0002-1628-1850

Giulia Atzeni – Department of Life and Environmental Sciences, Cittadella Universitaria, University of Cagliari, Cagliari 09124, Italy; orcid.org/0009-0005-1519-5431

Laura Demuru – Department of Life and Environmental Sciences, Cittadella Universitaria, University of Cagliari, Cagliari 09124, Italy; orcid.org/0009-0006-2424-3789

Pierluigi Caboni – Department of Life and Environmental Sciences, Cittadella Universitaria, University of Cagliari, Cagliari 09124, Italy; orcid.org/0000-0003-2448-3767

Donatella Valenti – Department of Life and Environmental Sciences, Cittadella Universitaria, University of Cagliari, Cagliari 09124, Italy; orcid.org/0000-0002-5081-0891

Present Addresses

[#]Faculty of Pharmacy, University of Ljubiana, Aškerčeva cesta 7, 1000 Ljubljana, Slovenia

Author Contributions

The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript. ‡These authors contributed equally. Conceptualization: R.M., A.F., and S.D.; Formal analysis: R.M., A.F., B.E., P.C. Investigation: R.M., S.D. B.E., S.F., A.L., F.C., A.O., E.S., D.S., G.A., L.D., D.V., and P.C; Methodology: R.M., A.F., and S.D.; Project administration: E.M., and A.F.; Resources: E.M. and A.F.; Supervision: R.M., S.D., F.C., and A.L.; Original draft preparation: R.M., S.D., and E.M.

Notes

All experiments involving hazardous materials were conducted in compliance with relevant safety protocols and guidelines. Appropriate safety measures, including the use of protective equipment and fume hoods, were employed for both synthetic and biochemical studies. The authors declare no competing financial interest.

ACKNOWLEDGMENT

Authors acknowledge the CeSAR (Centro Servizi d'Ateneo per la Ricerca) of the University of Cagliari, Italy for the ¹H NMR and ¹³C NMR experiments performed with Bruker Avance III HD 600.

ABBREVIATIONS

MAOs, Monoamine oxidases; DA, dopamine; NE, norepinephrine; 5HT, serotonin; AD, Alzheimer's disease; PD, Parkinson's disease; NDDs, neurodegenerative diseases; hMAO-A, human monoamine oxidase A; hMAO-B, human monoamine oxidase B, ADMET, absorption, distribution, metabolism, excretion, and toxicity; BBB, blood brain barrier; RMSD, root mean square deviation; MIFs, molecular interaction fields.

REFERENCES

- (1) Shih, J. C.; Chen, K.; Ridd, M. J., Monoamine oxidase: from genes to behavior. *Annu. Rev. Neurosci.* **1999**, *22*, 197-217.
- (2) Singer, T. P.; Von Korff, R. W.; Murphy, D. L.; Editors, *Monoamine Oxidase: Structure, Function, and Altered Functions. [Papers from a Symposium Held at the Michigan Molecular Institute, Midland, on July 15-19, 1979]*. Academic Press: 1979; p 557 pp.
- (3) Dezsi, L.; Vecsei, L., Monoamine Oxidase B Inhibitors in Parkinson's Disease. *CNS Neurol. Disord.: Drug Targets* **2017**, *16* (4), 425-439.
- (4) Perez-Lloret, S.; Rascol, O., The safety and efficacy of safinamide mesylate for the treatment of Parkinson's disease. *Expert Rev. Neurother.* **2016**, *16* (3), 245-258.
- (5) Bhat, M. A., Mofegiline helps alleviate symptoms of neuropathy in diabetic rats. *EJBPS* **2016**, *3* (2), 376-380.
- (6) Kong, P.; Zhang, B.; Lei, P.; Kong, X.; Zhang, S.; Li, D.; Zhang, Y., Neuroprotection of MAO-B inhibitor and dopamine agonist in Parkinson disease. *Int. J. Clin. Exp. Med.* **2015**, *8* (1), 431-439.
- (7) Inaba-Hasegawa, K.; Akao, Y.; Maruyama, W.; Naoi, M., Rasagiline and selegiline, inhibitors of type B monoamine oxidase, induce type A monoamine oxidase in human SH-SY5Y cells. *J. Neural Transm.* **2013**, *120* (3), 435-444.
- (8) Naoi, M.; Maruyama, W.; Inaba-Hasegawa, K., Type A and B monoamine oxidase in age-related neurodegenerative disorders: their distinct roles in neuronal death and survival. *Curr. Top. Med. Chem.* **2012**, *12* (20), 2177-2188.
- (9) Naoi, M.; Maruyama, W.; Inaba-Hasegawa, K.; Akao, Y., Type A monoamine oxidase regulates life and death of neurons in neurodegeneration and neuroprotection. In *International Review of Neurobiology*, Youdim, M. B. H.; Douce, P., Eds. Academic Press: 2011; Vol. 100, pp 85-106.
- (10) Foley, P.; Gerlach, M.; Youdim, M. B. H.; Riederer, P., MAO-B inhibitors: multiple roles in the therapy of neurodegenerative disorders? *Parkinsonism Relat. Disord.* **2000**, *6* (1), 25-47.
- (11) Edmondson, D. E.; Binda, C.; Mattevi, A., Structural insights into the mechanism of amine oxidation by monoamine oxidases A and B. *Arch. Biochem. Biophys.* **2007**, *464* (2), 269-276.
- (12) Li, M.; Binda, C.; Mattevi, A.; Edmondson, D. E., Functional role of the "aromatic cage" in human monoamine oxidase B: structures and catalytic properties of Tyr435 mutant proteins. *Biochemistry* **2006**, *45* (15), 4775-84.
- (13) Binda, C.; Li, M.; Hubálek, F.; Restelli, N.; Edmondson, D. E.; Mattevi, A., Insights into the mode of inhibition of human mitochondrial monoamine oxidase B from high-resolution crystal structures. *Proc. Natl. Acad. Sci. U. S. A.* **2003**, *100* (17), 9750-9755.
- (14) Binda, C.; Mattevi, A.; Edmondson, D. E., Structural properties of human monoamine oxidases A and B. In *International Review of Neurobiology*, Youdim, M. B. H.; Douce, P., Eds. Academic Press: 2011; Vol. 100, pp 1-11.
- (15) Hudspith, L.; Shmam, F.; Dalton, C. F.; Princivalle, A.; Turega, S. M., Neurotransmitter selection by monoamine oxidase isoforms, dissected

in terms of functional groups by mixed double mutant cycles. *Org. Biomol. Chem.* **2019**, *17* (39), 8871-8877.

(16) Kurihara, K.; Mishima, T.; Fujioka, S.; Tsuboi, Y., Efficacy and safety evaluation of safinamide as an add-on treatment to levodopa for parkinson's disease. *Expert Opin. Drug Saf.* **2022**, *21* (2), 137-147.

(17) Nam, M.-H.; Park, J.-H.; Song, H. J.; Choi, J. W.; Kim, S.; Jang, B. K.; Yoon, H. H.; Heo, J. Y.; Lee, H.; An, H.; Kim, H. J.; Park, S. J.; Cho, D.-W.; Yang, Y.-S.; Han, S.-C.; Kim, S.; Oh, S.-J.; Jeon, S. R.; Park, K. D.; Lee, C. J., KDS2010, a Newly Developed Reversible MAO-B Inhibitor, as an Effective Therapeutic Candidate for Parkinson's Disease. *Neurotherapeutics* **2021**, *18* (3), 1729-1747.

(18) Jha, S. K.; Jha, N. K.; Kumar, D.; Ambasta, R. K.; Kumar, P., Linking mitochondrial dysfunction, metabolic syndrome and stress signaling in neurodegeneration. *Biochim. Biophys. Acta, Mol. Basis Dis.* **2017**, 1132-1146.

(19) Czerniczyniec, A.; Bustamante, J.; Lores-Arnaiz, S., Modulation of brain mitochondrial function by deprenyl. *Neurochem. Int.* **2006**, *48* (3), 235-41.

(20) Wu, Y.; Kazumura, K.; Maruyama, W.; Osawa, T.; Naoi, M., Rasagiline and selegiline suppress calcium efflux from mitochondria by PK11195-induced opening of mitochondrial permeability transition pore: a novel anti-apoptotic function for neuroprotection. *J Neural Transm (Vienna)* **2015**, *122* (10), 1399-407.

(21) Olufunmilayo, E. O.; Gerke-Duncan, M. B.; Holsinger, R. M. D., Oxidative Stress and Antioxidants in Neurodegenerative Disorders. *Antioxidants* **2023**, *12* (2), 30.

(22) Nakamura, K., α -Synuclein and mitochondria: partners in crime? *Neurotherapeutics* **2013**, *10* (3), 391-9.

(23) Kang, S. S.; Ahn, E. H.; Zhang, Z.; Liu, X.; Manfredsson, F. P.; Sandoval, I. M.; Dhakal, S.; Iuvone, P. M.; Cao, X.; Ye, K., α -Synuclein stimulation of monoamine oxidase-B and legumain protease mediates the pathology of Parkinson's disease. *EMBO J.* **2018**, *37* (12).

(24) Nakamura, Y.; Arawaka, S.; Sato, H.; Sasaki, A.; Shigekiyo, T.; Takahata, K.; Tsunekawa, H.; Kato, T., Monoamine Oxidase-B Inhibition Facilitates α -Synuclein Secretion In Vitro and Delays Its Aggregation in rAAV-Based Rat Models of Parkinson's Disease. *J. Neurosci.* **2021**, *41* (35), 7479-7491.

(25) Tan, Y. Y.; Jenner, P.; Chen, S. D., Monoamine Oxidase-B Inhibitors for the Treatment of Parkinson's Disease: Past, Present, and Future. *J Parkinsons Dis* **2022**, *12* (2), 477-493.

(26) Meleddu, R.; Distinto, S.; Cirilli, R.; Alcaro, S.; Yanez, M.; Sanna, M. L.; Corona, A.; Melis, C.; Bianco, G.; Matyus, P.; Cottiglia, F.; Maccioni, E., Through scaffold modification to 3,5-diaryl-4,5-dihydroisoxazoles: new potent and selective inhibitors of monoamine oxidase B. *J. Enzyme Inhib. Med. Chem.* **2017**, *32* (1), 264-270.

(27) Distinto, S.; Meleddu, R.; Yanez, M.; Cirilli, R.; Bianco, G.; Sanna, M. L.; Arridu, A.; Cossu, P.; Cottiglia, F.; Faggi, C.; Ortuso, F.; Alcaro, S.; Maccioni, E., Drug design, synthesis, in vitro and in silico evaluation of selective monoaminoxidase B inhibitors based on 3-acetyl-2-

dichlorophenyl-5-aryl-2,3-dihydro-1,3,4-oxadiazole chemical scaffold. *Eur. J. Med. Chem.* **2016**, *108*, 542-552.

(28) Cardia, M. C.; Sanna, M. L.; Meleddu, R.; Distinto, S.; Yanez, M.; Vina, D.; Lamela, M.; Maccioni, E., A novel series of 3,4-disubstituted dihydropyrazoles. Synthesis and evaluation for MAO enzyme inhibition. *J. Heterocycl. Chem.* **2013**, *50* (S1), E87-E92.

(29) Secci, D.; Carradori, S.; Bolasco, A.; Bizzarri, B.; D'Ascenzio, M.; Maccioni, E., Discovery and optimization of pyrazoline derivatives as promising monoamine oxidase inhibitors. *Curr. Top. Med. Chem.* **2012**, *12* (20), 2240-2257.

(30) Distinto, S.; Yanez, M.; Alcaro, S.; Cardia, M. C.; Gaspari, M.; Sanna, M. L.; Meleddu, R.; Ortuso, F.; Kirchmair, J.; Markt, P.; Bolasco, A.; Wolber, G.; Secci, D.; Maccioni, E., Synthesis and biological assessment of novel 2-thiazolyldrazones and computational analysis of their recognition by monoamine oxidase B. *Eur. J. Med. Chem.* **2012**, *48*, 284-295.

(31) Maccioni, E.; Alcaro, S.; Cirilli, R.; Vigo, S.; Cardia, M. C.; Sanna, M. L.; Meleddu, R.; Yanez, M.; Costa, G.; Casu, L.; Matyus, P.; Distinto, S., 3-Acetyl-2,5-diaryl-2,3-dihydro-1,3,4-oxadiazoles: A new scaffold for the selective inhibition of monoamine oxidase B. *J. Med. Chem.* **2011**, *54* (18), 6394-6398.

(32) Dunkel, P.; Balogh, B.; Meleddu, R.; Maccioni, E.; Gyires, K.; Matyus, P., Semicarbazide-sensitive amine oxidase/vascular adhesion protein-1: a patent survey. *Expert Opin. Ther. Pat.* **2011**, *21* (9), 1453-1471.

(33) Maccioni, E.; Alcaro, S.; Orallo, F.; Cardia, M. C.; Distinto, S.; Costa, G.; Yanez, M.; Sanna, M. L.; Vigo, S.; Meleddu, R.; Secci, D., Synthesis of new 3-aryl-4,5-dihydropyrazole-1-carbothioamide derivatives. An investigation on their ability to inhibit monoamine oxidase. *Eur. J. Med. Chem.* **2010**, *45* (10), 4490-4498.

(34) Chimenti, F.; Secci, D.; Bolasco, A.; Chimenti, P.; Granese, A.; Carradori, S.; Maccioni, E.; Cardia, M. C.; Yanez, M.; Orallo, F.; Alcaro, S.; Ortuso, F.; Cirilli, R.; Ferretti, R.; Distinto, S.; Kirchmair, J.; Langer, T., Synthesis, semipreparative HPLC separation, biological evaluation, and 3D-QSAR of hydrazothiazole derivatives as human monoamine oxidase B inhibitors. *Bioorg. Med. Chem.* **2010**, *18* (14), 5063-5070.

(35) Cerioni, G.; Maccioni, E.; Cardia, M. C.; Vigo, S.; Mocchi, F., Characterization of 2,5-diaryl-1,3,4-oxadiazolines by multinuclear magnetic resonance and density functional theory calculations. Investigation on a case of very remote Hammett correlation. *Magn. Reson. Chem.* **2009**, *47* (9), 727-733.

(36) Chimenti, F.; Maccioni, E.; Secci, D.; Bolasco, A.; Chimenti, P.; Granese, A.; Carradori, S.; Alcaro, S.; Ortuso, F.; Yanez, M.; Orallo, F.; Cirilli, R.; Ferretti, R.; La Torre, F., Synthesis, Stereochemical Identification, and Selective Inhibitory Activity against Human Monoamine Oxidase-B of 2-Methylcyclohexylidene-(4-arylthiazol-2-yl)hydrazones. *J. Med. Chem.* **2008**, *51* (16), 4874-4880.

(37) Chimenti, F.; Maccioni, E.; Secci, D.; Bolasco, A.; Chimenti, P.; Granese, A.; Befani, O.; Turini, P.; Alcaro, S.; Ortuso, F.; Cardia, M. C.; Distinto, S., Selective Inhibitory Activity against MAO and Molecular

Modeling Studies of 2-Thiazolylhydrazone Derivatives. *J. Med. Chem.* **2007**, *50* (4), 707-712.

(38) Chimenti, F.; Maccioni, E.; Secci, D.; Bolasco, A.; Chimenti, P.; Granese, A.; Befani, O.; Turini, P.; Alcaro, S.; Ortuso, F.; Cirilli, R.; La Torre, F.; Cardia, M. C.; Distinto, S., Synthesis, molecular modeling studies, and selective inhibitory activity against monoamine oxidase of 1-thiocarbamoyl-3,5-diaryl-4,5-dihydro-(1H)- pyrazole derivatives. *J. Med. Chem.* **2005**, *48* (23), 7113-7122.

(39) Schrödinger *Release 2024-2 QikProp S*, New York, NY, 2024.

(40) Berman, H. M.; Westbrook, J.; Feng, Z.; Gilliland, G.; Bhat, T. N.; Weissig, H.; Shindyalov, I. N.; Bourne, P. E., The Protein Data Bank. *Nucleic Acids Res.* **2000**, *28* (1), 235-242.

(41) Binda, C.; Hubálek, F.; Li, M.; Herzig, Y.; Sterling, J.; Edmondson, D. E.; Mattevi, A., Binding of Rasagiline-Related Inhibitors to Human Monoamine Oxidases: A Kinetic and Crystallographic Analysis. *J. Med. Chem.* **2005**, *48* (26), 8148-8154.

(42) Binda, C.; Li, M.; Hubálek, F.; Restelli, N.; Edmondson, D. E.; Mattevi, A., Insights into the mode of inhibition of human mitochondrial monoamine oxidase B from high-resolution crystal structures. **2003**, *100* (17), 9750-9755.

(43) Hubálek, F.; Binda, C.; Khalil, A.; Li, M.; Mattevi, A.; Castagnoli, N.; Edmondson, D. E., Demonstration of Isoleucine 199 as a Structural Determinant for the Selective Inhibition of Human Monoamine Oxidase B by Specific Reversible Inhibitors. *J. Biol. Chem.* **2005**, *280* (16), 15761-15766.

(44) Binda, C.; Wang, J.; Pisani, L.; Caccia, C.; Carotti, A.; Salvati, P.; Edmondson, D. E.; Mattevi, A., Structures of Human Monoamine Oxidase B Complexes with Selective Noncovalent Inhibitors: Saffinamide and Coumarin Analogs. *J. Med. Chem.* **2007**, *50* (23), 5848-5852.

(45) Reis, J.; Manzella, N.; Cagide, F.; Mialet-Perez, J.; Uriarte, E.; Parini, A.; Borges, F.; Binda, C., Tight-Binding Inhibition of Human Monoamine Oxidase B by Chromone Analogs: A Kinetic, Crystallographic, and Biological Analysis. *J. Med. Chem.* **2018**, *61* (9), 4203-4212.

(46) Chung, J. Y.; Hah, J.-M.; Cho, A. E., Correlation between Performance of QM/MM Docking and Simple Classification of Binding Sites. *J. Chem. Inf. Model.* **2009**, *49* (10), 2382-2387.

(47) Halgren, T. A., Identifying and Characterizing Binding Sites and Assessing Druggability. *J. Chem. Inf. Model.* **2009**, *49* (2), 377-389.

(48) Goodford, P. J., A computational procedure for determining energetically favorable binding sites on biologically important macromolecules. *J. Med. Chem.* **1985**, *28* (7), 849-857.

(49) Edmondson, D. E.; Binda, C.; Wang, J.; Upadhyay, A. K.; Mattevi, A., Molecular and mechanistic properties of the membrane-bound mitochondrial monoamine oxidases. *Biochemistry* **2009**, *48* (20), 4220-30.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60