



UNICA

UNIVERSITÀ
DEGLI STUDI
DI CAGLIARI



Università di Cagliari

UNICA IRIS Institutional Research Information System

This is the Author's [*accepted*] manuscript version of the following contribution:

Lanzolla Giulia

Graves disease: latest understanding of pathogenesis and treatment options

Titolo della Rivista

NATURE REVIEWS. ENDOCRINOLOGY

The publisher's version is available at: Cod DOI

<https://dx.doi.org/10.1038/s41574-024-01016-5>

When citing, please refer to the published version.

AAM – (versione Post-print)

Graves' disease: latest understanding of pathogenesis and treatment options

Giulia Lanzolla^{1,2}, Michele Marinò¹, Francesca Menconi³

¹ Department of Clinical and Experimental Medicine, Endocrinology Unit II, University of Pisa and University Hospital of Pisa, Via Paradisa 2, 56124, Pisa, Italy.

² Department of Orthopaedic Surgery, University of Pennsylvania, Philadelphia, Stammler Hall, 3450 Hamilton Walk, PA, USA

³ U.O Endocrinologia II, Azienda Ospedaliero Universitaria Pisana, University Hospital of Pisa, Via Paradisa 2, 56124, Pisa, Italy.

*Corresponding Author

Francesca Menconi

U.O Endocrinologia II, Azienda Ospedaliero Universitaria Pisana, University Hospital of Pisa, Via Paradisa 2, 56124, Pisa, Italy.

E-mail: fmenconi.mail@gmail.com

Abstract

Graves' disease (GD) is the most common cause of hyperthyroidism in iodine-sufficient areas. The main responsible mechanism is related to autoantibodies that bind and activate the thyrotropin receptor (TSH-R). Although Graves' hyperthyroidism (GH) is relatively common, no causal treatment options are available. Established treatment modalities are anti-thyroid drugs (ATD), which reduce thyroid hormone synthesis, radioactive iodine (RAI), and surgery. However, emerging drugs that target the main autoantigen (monoclonal antibodies, small molecules, peptides) or block the immune pathway have been recently tested in clinical trials. GD can involve the thyroid exclusively, or it can be associated with extrathyroidal manifestations, among which Graves' orbitopathy (GO) is the most common. The presence of GO can change the management of patients. An established treatment for moderate-to-severe GO is intravenous glucocorticoids. However, recent advances in understanding GO pathogenesis have allowed the development of new target-based therapies, by blocking proinflammatory cytokine receptors, lymphocytic infiltration, or the insulin-like growth factor receptor (IGF-1R), with several clinical trials providing promising results. This article aims to review the new discoveries in the pathogenesis of GH and GO that are offering several important tools in patient management.

Introduction

Graves' Disease (GD) is an autoimmune disease characterized by an enlarged and overactive thyroid gland¹. The main mechanism responsible for Graves' hyperthyroidism (GH) is the activation of the thyroid-stimulating hormone receptor (TSH-R) by autoantibodies that act as agonists, causing thyrocyte proliferation and hyperfunction^{1,2}. Genetic and environmental factors contribute to loss of immune self-tolerance, ultimately triggering the pathogenic process^{1,3,4}. New insights into GD have been provided, including the role of microbiota and polymorphisms of elements involved in immune checkpoints and immune response, namely protein tyrosine phosphatase non-receptor type 22 (PTPN22), cytotoxic T lymphocyte antigen-4 (CTLA-4), cluster of differentiation 40 (CD-40), and Fc receptor-like protein 3 (FCRL3)⁵⁻¹⁴. Furthermore, antigen-specific immunotherapy with two TSH-R peptides has been proposed¹⁵. Although conventional treatments of GH with antithyroid drugs, surgery, and radioactive iodine (RAI) are commonly used in clinical practice, novel tools are on the way (Table 1) (Figure 1).

GD affects primarily the thyroid, but extrathyroidal manifestations can be present (Box 1)^{16,17}. Graves' orbitopathy (GO) is an autoimmune syndrome affecting orbital fibroadipose tissue, due to an interplay between cellular and humoral immunity against TSH-R and possibly other autoantigens expressed by thyroid epithelial cells (TEC) and orbital fibroblasts (OFs)¹⁸. Treatment of GO is currently changing due to new insights into its pathogenesis (Table 1) (Figure 2).

This review aims at providing an overview of epidemiology, pathogenesis, and conventional treatment of GH and GO, and discussing the most recent research acquisitions that have allowed novel treatment modalities. Relevant studies published between Jan 1, 2000, and Jan 1, 2023, were identified from the PubMed, EMBASE, Cochrane Library, and the ClinicalTrials.gov registry, using the search terms "Graves' disease treatment", "Graves' disease therapy", "Graves' disease management", "Graves' disease pathogenesis", "Graves' orbitopathy", "Graves' ophthalmopathy", "thyroid eye disease", "thyroid-associated ophthalmopathy", "thyroid ophthalmopathy", "endocrine ophthalmopathy". In vitro studies, preclinical studies, retrospective studies and randomized clinical trials performed in Europe and US were included. Only English language articles were included. Only peer-reviewed journal articles were included.

Epidemiology

The prevalence of GD is ~1.2% worldwide, with a female-to-male ratio of ~ 10:1. The incidence ranges from 20 to 40 new cases/100 000/year, with a lifetime risk of 3% for women and

0.5 % for men^{191,20}. Patients can be affected at any age, but the incidence peaks between 30 and 50 years^{1,19,20}.

GO is observed in ~25-30% of GH patients^{21,22} with an incidence of 0.54-0.9 cases/100.000/year in men and 2.67-3.3 cases/100.000/year in women²². Mild forms are more common, whereas severe GO accounts for 5-6% of cases^{18,22}, the latter being more common in men²³. A recent prospective, observational study provided new trends in clinical presentation and management of GO²²: patients were older in 2019 compared to 2012, with a reduced frequency of moderate-to-severe forms, suggesting improvement in management timing and treatment²².

Pathogenesis and molecular mechanisms

Graves' disease

Genes that confer susceptibility have been identified^{244,25} as well as hypermethylation of several genes, including those encoding TSH-R and proteins involved in T-cell signaling²⁵ (Box 2). As an autoimmune condition, GD arises from the breakdown of self-tolerance towards autoantigens, with TSH-R being the most prominently recognized. Antigen-presenting cells (APCs), such as B-cells, macrophages, dendritic cells, and thyroid cells, identify TSH-R peptides. The interaction between the T-cell receptor (TCR) and major histocompatibility complex (MHC) triggers T-cell activation, subsequently activating B-cells through the interplay of TCR and MHC, along with the binding of cluster of differentiation 40 (CD40) to its ligand CD40L. Following activation, B-cells differentiate into plasma cells, producing TSH-R autoantibodies (TRAbs) that stimulate thyroid cells, leading to increased proliferation and hormone secretion, ultimately resulting in hyperthyroidism.

TSH-R autoantibodies

TSH-R is a G-protein coupled receptor (GPCR) with seven transmembrane helices. The signal transduction of the TSH-R is intricate and is currently being refined. It encompasses beta-arrestin signaling, IGF-1R crosstalk, and engagement with various G proteins. The receptor exists in multiple configurations, and there are diverse signal pathways that may or may not be activated as the receptor conformation changes upon ligand binding. This dynamic process can persist even after the receptors are internalized²⁶. Recently, an elegant computer-based approach has been

introduced to gain insights into the hinge region, enabling the development of a comprehensive full-length model of the TSH-R²⁷.

IgG1 subclass autoantibodies binding TSH-R (TRAbs), originally referred to as long-acting thyroid stimulators²⁸⁻³⁰, play a central role in GD³¹. Immunization against the TSH-R is essential for the development of GD in *in vivo* models (Box3). Oligoclonal production of TRAbs mostly arises from intrathyroidal B cells, reflecting the role of primary immune response³². TRAbs act as TSH-R agonists, promoting thyrocyte proliferation and activity³³, leading to increased thyroid hormone production. TRAbs recognize epitopes on the TSH-R- α subunit, which can be secreted in a soluble form and may trigger and/or enhance the autoimmune reaction to the full-length receptor^{34,35}. TSH-R also exists as multimers or high-order complexes³⁶, and multimeric forms of the α subunit may affect the affinity maturation of TRAbs³⁷. Although stimulating TRAbs (TSAb) are the hallmark of GD, TSH-R-blocking (TSAb) and neutral antibodies rarely, have been detected³⁸. In these patients, thyroid function can fluctuate between hyper- and hypothyroidism according to the serum levels of TBAb or TSAbs³⁹.

Immune tolerance escape

Inflammatory reactions reflect delicate balance between T-helper (Th) and T-regulatory (Treg)cells. Tregs exert an immune-modulating action by reducing Th1 (involved in cytotoxic immune response) and Th2 cell activity (responsible for antibody-mediated response). A decrease in Treg-dependent immunosuppressive action has been reported in GD^{40,41}.

Thymic processing of immune cells provides deletion of autoreactive T-cells by clonal deletion and diversion⁴². T-cells selected to differentiate further, become Th1, Th2, or Th17, and each phenotype produces a unique pattern of cytokines⁴². Peripheral tolerance processing exists wherein self-reactive T-cells become unresponsive or are deleted after encountering self-antigens outside the thymus⁴². Autoreactive T-cells against TSH-R escape both central and peripheral tolerance. Thus, they recognize TSH-R peptides presented by APCs (Figure1). Intrathyroidal T-cells mostly display a Th2 phenotype⁴³. However, whether GD is driven by Th1 or Th2 response remains a matter of debate⁴⁴.

Both T and B-cells are involved in GD. In addition to acting as APCs, B cells differentiate into antibody-producing plasma cells through antigen-binding to B-cell receptor and a second signal provided by the costimulatory pathway CD40-CD40L (Figure 1).

An *in vivo* study in a mouse model suggested that the checkpoint regulator VISTA (T-type immunoglobulin domain-containing suppressor of T-cell activation) may play a role in autoimmunity development⁴⁵, and further investigations into this mechanism are warranted in GD.

The role played by TEC in GD development is not well recognized. Their ability to release chemokines suggests a potential involvement in recruiting immune cells⁴⁶. Furthermore, TEC in GD express MHC class II molecules and CD40, and may act as APCs.

Graves' orbitopathy

GO is an autoimmune condition characterized by immune-mediated inflammatory reactions against autoantigens shared by TEC and OFs¹⁸ (Figure 2). OFs include a subset of CD34⁺ bone-marrow-derived fibrocytes that are the primary target of autoimmunity^{1,47}. GO shows a multifactorial pathogenesis, with both genetic and environmental risk factors playing a role (Box 4).

TSH-R

After TSH-R expression in orbital tissue was first shown⁴⁸, immunohistochemical studies revealed TSH-R overexpression in GO orbital tissues⁴⁹. The expansion of orbital adipose tissue is promoted by TSH-R activation, which triggers OFs and preadipocyte differentiation into adipocytes⁵⁰. Mouse experimental models support the involvement of TSH-R⁵¹⁻⁵³. High levels of circulating TRAbs are associated with GO activity^{54,55} and TRAbs are an independent risk factor for GO⁵⁶.

IGF-1R

A key role of insulin-like growth factor 1 receptor (IGF-1R) in GO has been described^{57,58}. IGF-1R is overexpressed in both B-⁵⁹ and T-cells⁶⁰ from GD patients. GD-IgG binding to IGF-1R triggers OF proliferation and secretion of hyaluronan and cytokines⁶¹. IGF-1R and TSH-R are colocalized in OFs⁶². Whether IGF-1R activation occurs through binding of stimulatory IGF-1R antibodies (IGF-1R-Abs) or exclusively via the established cross-talk between TSH-R and IGF-1R is not yet elucidated. Studies on IGF-1R Abs have yielded conflicting results^{63,64}. A recent study suggests that IGF-1R Abs may protect against GO occurrence⁶⁵.

The orbital infiltration: T and B-cells

Orbital immune infiltration changes over the natural course of GO, which is characterized by an early cell-mediated immune reaction followed by humoral immunity⁶⁶. In the first 18-24 months, Th1 cells are predominant and produce interleukin-2 (IL-2), IL-1, interferon- γ (IFN- γ), and tumor necrosis factor- α (TNF- α), which stimulate OFs to produce glycosaminoglycans. Later, the orbital infiltrate consists mainly of Th2 cells, which activate B-cells and antibody production. IL-4 produced by Th2 promotes collagen type 4 synthesis causing fibrosis. Th17 and Treg cells have also been reported to play a role, by inducing expansion of extracellular matrix and development of fibrosis^{67,68}. Besides T-cells, the orbital infiltrate in GO patients also consists of B-cells and both correlate with GO activity⁶⁹. B-cells work as APCs, interact with T-cells through CD40-CD40L costimulatory pathway, and differentiate into plasmacells that release TRAbs (Fig.2)^{1,70}.

Orbital fibroblasts

The interaction between T-cells, OFs, and TRAbs contributes to OF activation (Fig.2). Once activated, OFs proliferate, secrete inflammatory molecules and glycosaminoglycans and differentiate into adipocytes, leading to the tissue remodeling typical of GO¹. Since OFs are the target cells, their heterogeneity may affect tissue remodeling. In this regard, the surface marker CD90 has been reported to divide OFs into two subsets with different phenotypes and functions. Indeed, CD90⁺OFs promote fibrosis whereas CD90⁻ OFs promote adipogenesis⁷⁰.

Epigenetics: new acquisitions

Epigenetics, a new research area in GO pathogenesis, includes histone modification, DNA methylation, and non-coding RNA (ncRNA). Hypermethylation or hypomethylation of certain genes may occur in OFs in the setting of autoimmunity⁷¹, resulting in differential gene expression, which may affect cell proliferation and cell response to inflammation. NcRNA includes RNA transcripts lacking protein-coding functions, which can be classified into long non-coding RNA (lncRNA), circular RNA (circRNA), and microRNA (miRNA). The involvement of miRNA in GO is supported by several findings. miR-Let7-5p is higher in GD patients without GO compared to those with GO. Furthermore, miR-Let7-5p negatively correlates with the Clinical Activity Score (CAS) and the severity of GO⁷². Low serum miR-224-5p has been associated with reduced sensitivity to glucocorticoids⁷³. Finally, miR-146a expression is higher in OFs compared to controls and is related to the production of TGF- β -induced fibrosis markers⁷⁴.

Treatment

Graves' hyperthyroidism

Conventional treatments

Current treatment options for GH include antithyroid drugs (ATD), surgery and RAI. ATD, (methimazole and propylthiouracil) are the first-line treatment to restore euthyroidism⁷⁵. The most common side effects (reported in up to 13% of treated patients) are urticaria, pruritus, arthralgia and gastrointestinal symptoms⁷⁶. Major adverse events are rare (0.1-0.3% of patients), but might be life-threatening, and include agranulocytosis, hepatotoxicity, aplastic anemia, thrombocytopenia and vasculitis⁷⁷. Current guidelines recommend a course of ATD lasting 12–18 months. However, while a 12–18 month initial treatment period is reasonable for patients responding quickly to treatment and likely to achieve remission, the relapse rate after discontinuation of ATD is ~50%^{78,79}, and a more personalized approach is recommended⁸⁰. Patients with low chances of remission at the onset (e.g., high TRAb titres and smoking habit) or patients for whom relapse could be detrimental due to underlying conditions (e.g., patients with heart disease), may benefit from definitive therapy from the beginning or long-term ATD. RAI or surgery should be also considered in patients who experience GD relapse after ATD discontinuation, as a second ATD course rarely brings long-term remission.

RAI is a safe and well-tolerated option. Although it leads to hypothyroidism and lifelong replacement with levothyroxine, may be deemed necessary and can be open for discussion with patients, considering their perspectives and preferences. RAI is associated with *de novo* development or progression of GO²¹, which is prevented by steroids given after radioiodine^{21,81}. Rare RAI adverse events are sialadenitis and subacute thyroiditis, which may occur in ~1% of patients and can be treated with non-steroidal anti-inflammatory drugs or steroids^{80,82}.

Thyroidectomy can be considered in selected cases, namely large goiters, uncontrolled hyperthyroidism, severe GO, suspected thyroid cancer, patient preference⁸³.

Novel treatments

In patients with GH, the breakdown of immune tolerance triggers the recognition of self-antigens leading to a cascade of events that ultimately result in the production of TRAbs. None of the conventional treatments for GH currently available act on its pathogenesis. Recently, new

molecules that target different steps in the pathogenesis of GH, including early recognition of TSH-R peptides, T-cell activation, B-cell stimulation and survival, TRAbs production and TSH-R activation, have been tested with encouraging results (Figure 1) (Table 1)^{84,85}.

I-ATX-GD-59.

Antigen-specific immunotherapy with intact allergens has been successfully used in allergic disorders since 1911. Subsequently, the principle of tolerance induction has been applied to autoimmune diseases⁸⁶. Progress has been made with the discovery of synthetic peptides termed “apitopes” (antigen-processing independent epitopes) that mimic naturally processed CD4+ T-cell epitopes. Apitopes can induce immune tolerance without the risk of an immune response to the administered antigen. ATX-GD-59, a TSH-R apitope, is a combination of two TSH-R synthetic peptides. ATX-GD-59 has been shown to suppress CD4+ T-cell and T-dependent antibody responses to TSH-R in GD-associated HLA class II (*DRB1*0301*) transgenic mice⁸⁷. Immunotherapy with ATX-GD-59 has been tested in a Phase I Study, in GD patients with newly diagnosed and untreated hyperthyroidism¹⁵. Ten patients were treated with ATX-GD-59 in 10 i.d. injections over 18 weeks and followed for 12 weeks. In 50% of patients, normalization of free triiodothyronine was observed. Given the involvement of TSH-R, this treatment may also have a beneficial effect on GO¹⁵. ATX-GD-59 was well tolerated, with only mild local reactions in injection sites.

K1-70

K1-70 is a human monoclonal antibody blocking TSH-R. It was isolated from a patient with thyroid autoimmune disease and high levels of serum TRAbs⁸⁸. Among several TSH-R-blocking antibodies (i.e.: CS-17, M22, Mab-B2, 5C9), K1-70 is the most effective inhibitor of TSH-R *in vitro*⁸⁹ and it has been shown to reduce free and total T4 concentrations in rats⁸⁹. The Rees Smith group, in 2022, published the results of a phase I clinical trial aimed at testing safety, tolerability, pharmacokinetics and pharmacodynamics⁹⁰. Eighteen GD patients with hyperthyroidism controlled with ATD were treated with a single intramuscular or intravenous dose of K1-70. At the different doses tested, the drug was well tolerated with mild to moderate adverse events, among which fatigue, lethargy and diarrhea were the most common, and no serious adverse events. At the highest doses (25 mg im, 50 mg IV and 150 mg IV), evolution to hypothyroidism occurred in 100% of subjects. Improvement of GH was reported and an improvement in exophthalmos (>2

mm) was registered. These promising results make K1-70 a very interesting candidate among the new molecules to treat GH and GO⁹⁰.

Small molecule TSH-R antagonists

Small molecule agonists and antagonists can directly activate or inhibit TSH-R signaling. ANTAG-3, the best-studied inverse agonist, inhibits basal and agonist-induced signaling by decreasing TSH-R activity^{91,92}. These compounds inhibited TSH-stimulated cAMP production *in vitro* and reduced thyroid hormone levels in mice treated with the thyroid-stimulating monoclonal antibody M22, suggesting they may prevent TRAb-induced GH. VA-K-14 and S37a, two additional TSH-R antagonists, reduce TSH- and TRAb-induced signaling *in vitro*^{93,94}. These small molecules could lead to novel therapies for GD and GO.

Iscalimab

CD40 is a tumor necrosis factor receptor expressed on APCs. CD40 binds CD40L on T cells and has a critical role in antigen presentation, by activation of a costimulatory pathway that leads to B-cell proliferation, Ig class switching and germinal centre formation⁹⁵. Furthermore, CD40 is one of the major susceptibility genes for GD³. Blocking CD40 activation in animal models suppresses experimental GD⁹⁶, whereas overexpression of CD40 in the thyroid leads to increased antibody production and a more severe experimental GD⁹⁷. Iscalimab (CFZ533) is a fully human, non-depleting, monoclonal antibody that blocks CD154 (or CD40L) binding to CD40 and the subsequent activation pathway (Figure 1). A clinical phase two, open-label, single-arm study was conducted to test the effect of Iscalimab in GH¹¹. Fifteen hyperthyroid patients were treated with five doses of Iscalimab (10 mg/kg) intravenously over 12 weeks and observed for 24 weeks. In 7/15 (47%) patients, free and total T3 and T4 normalized at week 20, and 2/15 (13.3%) patients had normalization of TSH as well. Six patients (40%) were considered non-responders, since they required ATD to normalize thyroid function during the follow-up. TRAb levels decreased across the study. Intriguingly, all patients with TSH-R-Ab < 20IU/L were responders. While the limited sample size precludes definitive conclusions, this evidence could be taken into consideration to assess the optimal population that stands to gain the most from the administration of Iscalimab. No serious adverse events were observed; however, 80% of patients had mild adverse events that did not require withdrawal of study medication (except one case of moderate cystitis). In conclusion, the efficacy of Iscalimab seems similar to that of a 12-month course of ATD with a

good safety profile. In order to understand why about half of the patients were non-responders, it was investigated whether a specific genetic background could interfere and predict response. Seven single nucleotide polymorphisms (SNP) of CD40 gene were analyzed in 13 patients treated with Iscalimab and three haplotype profiles were found. Two were associated with higher CD40 mRNA levels and response to treatment, and the third was related to low CD40 mRNA levels and no response to treatment¹⁰. These data suggest that certain genetic variants of the CD40 gene are associated with higher expression of CD40 and good response to Iscalimab.

Belimumab

Belimumab is a fully humanized IgG1 monoclonal antibody that targets B-cell activating factor (BAFF) and prevents its interaction with BAFF receptors on the B cell surface, ultimately resulting in B cell failure to mature, expand, and differentiate into antibodies-producing plasmacells. Increased circulating BAFF levels and a significant positive correlation between TRAbs and BAFF levels have been found in GH patients⁹⁸. Genetic variants in *BAFF* seem to increase the susceptibility to GD⁹⁹. The role of BAFF/BAFF-R interactions in GD pathogenesis is supported by their increased expression in infiltrating immune cells and thyrocytes from GH patients¹⁰⁰ and by the evidence that blocking BAFF by a BAFF-specific receptor-Fc in a GD mouse model reduced the resulting hyperthyroidism¹⁰¹. Further clinical studies are warranted to investigate the potential efficacy of Belimumab for treating GH.

Graves' orbitopathy

Conventional treatments

Intravenous glucocorticoids (ivGCs) have been used as the first-line treatment of moderate-to-severe, active GO since approximately 30 years. However, the recent European Group of Graves' Orbitopathy (EUGOGO) guidelines suggest the addition of mofetil mycophenolate to methylprednisolone²¹. Mycophenolate acts by inhibiting T and B-cells proliferation, and antibody and adhesion molecule production¹⁰². A multicenter randomized clinical trial has been conducted entailing the enrollment of 164 patients randomized to receive methylprednisolone (500 mg once/week for 6 weeks followed by 250 mg once/week for 6 weeks) or methylprednisolone plus oral mycophenolate (720 mg daily for 24 weeks). Even though there was no significant difference at 12 weeks (primary endpoint), the combination therapy provided a greater overall response rate

and improvement in the individual GO features compared to methylprednisolone monotherapy at 24 and 36 weeks¹⁰³.

Orbital radiotherapy is an effective second-line treatment for moderate-to-severe and active GO, in combination with glucocorticoids, mainly in patients with diplopia and/or restriction of extraocular motility²¹. Orbital decompression is used for ivGC-resistant sight-threatening GO or rehabilitative purposes. Squint and eyelid surgery can be used for rehabilitative and/or aesthetic purposes²¹.

The conventional approach to treating mild Graves Orbitopathy (GO) primarily involved local measures, with major treatments reserved for cases significantly impacting quality of life. Given the role of oxidative stress in GO, the European Group on Graves' Orbitopathy (EUGOGO) conducted a randomized, placebo-controlled clinical trial in selenium-deficient European countries to evaluate the effects of selenium and pentoxifylline in mild GO¹⁰⁴. The study included 159 patients with mild GO who were randomized to receive sodium selenite, pentoxifylline, or placebo for six months, followed by a six-month follow-up period. The study assessed the overall eye outcomes, including exophthalmometry, clinical activity score (CAS), eyelid aperture, diplopia, and visual acuity. Selenium treatment showed a significant outcome, with 61% of patients experiencing GO improvement compared to 36% in the placebo group. Notably, selenium-treated patients had better quality of life scores, including visual functioning and appearance, with persistent benefits even at the 12-month follow-up visit. Even though the study had limitations, such as the lack of baseline and end-of-treatment selenium concentration measurements, which leaves uncertainty about whether selenium deficiency correction contributed to the observed benefits, these promising results opened the possibility of the clinical use of selenium in patients with mild GO. Indeed, a 6-month selenium supplementation should be given to patients with mild and active GO of recent onset, according to the most recent EUGOGO guidelines²¹.

Novel treatments

Teprotumumab

Teprotumumab is a humanized monoclonal antibody that blocks IGF-1R. In 2017, in a double-masked, multicenter, randomized trial, 88 patients with moderate-to-severe active GO were assigned to receive placebo or teprotumumab (eight intravenous infusions at 3-week intervals, with a starting dose of 10 mg/kg, which was later increased to 20 mg/kg)¹⁰⁵. The primary endpoint was

a composite evaluation which included reduction of CAS by at least 2 points and decrease in exophthalmos by at least 2 mm. At 24 weeks, a greater proportion of responders (69%) was observed in the teprotumumab group. The most notable effect was reduction of exophthalmos, with an average decrease of ~ 2.5 mm¹⁰⁵. Subsequently, a phase 3 clinical trial was performed. The study included 41 patients treated with teprotumumab and 42 patients with placebo. The primary outcome was a decrease in exophthalmos by at least 2 mm⁵⁷. The response rate to teprotumumab was 83%, compared to 10% in the placebo group. Additionally, secondary outcomes such as CAS, diplopia, and quality of life (QoL) were better in patients given teprotumumab⁵⁷. The results led to the approval of teprotumumab by the US Food and Drug Administration in January 2020 for GO, regardless of activity and severity. A pooled data analysis of these two studies with an off-treatment follow-up has been performed to define long-term responses¹⁰⁶. At 72 weeks following the discontinuation of Teprotumumab, 33% of patients observed a deterioration in exophthalmos, while 31% experienced diplopia relapse. Moreover, based on composite evaluation, 17% of patients were no longer considered responders in the long-term term¹⁰⁶. In a continuation of the second clinical trial teprotumumab was administered to non-responders or those who experienced a flare-up¹⁰⁷. The majority of them achieved a response to treatment¹⁰⁷.

Unfortunately, there are some limitations to using teprotumumab¹⁰⁸. Costs of treatment are quite high and the safety profile suggests caution. Thus, although teprotumumab was found to be generally safe in clinical trials^{57,105}, it carries hyperglycemia risk, especially in patients with pre-existing diabetes^{57,105,109}, and real-world data indicate hearing impairment as a relatively frequent event (12%)¹¹⁰. Ulcerative colitis^{111,112} and amyloid encephalopathy¹¹³ have also been reported during the post-marketing period¹⁰⁸.

Rituximab

Rituximab is a chimeric human/mouse monoclonal antibody that binds CD-20 positive B-cells inducing their depletion¹¹⁴. In addition, rituximab reduces IGF1-R positive T-cells¹¹⁵. Following early uncontrolled studies indicating a potential benefit in moderate-to-severe, active GO^{116,117}, two randomized clinical trials were conducted resulting in conflicting results^{118,119} that can be explained, at least in part, by the longer GO duration in one study¹²⁰. According to a meta-analysis including 12 studies, rituximab may have a beneficial effect on GO activity, measured by CAS, and low efficacy in improving proptosis¹²¹. Recently, a single 100 mg infusion of rituximab has

been reported to achieve GO inactivation in 15/17 GC-resistant patients with moderate-to-severe active GO. Additionally, around 60% of patients showed improvement in GO severity, and no flares were seen¹²². For the time being, rituximab may be considered as a second-line treatment for moderate-to-severe active GO^{21,108}. Despite its efficacy, the safety profile of rituximab still carries concerns. The most frequent adverse events include nose and throat itches, infusion reactions, nausea, and fever, but severe infections, progressive multifocal leukoencephalopathy, vasculitis and gastrointestinal complications have been described^{21,118}. The use of rituximab has been also tested in GH. A prospective study with a small cohort of patients suggested a beneficial role of rituximab in mild relapsing GH¹²³. A prospective study by El Fassi et al. compared short-term methimazole treatment with or without rituximab in 20 GH patients¹²⁴. Treatment with rituximab resulted in sustained remission in 4/10 patients after 24 months compared to 0/10 in the control group, and it was most successful in those with low TRAb levels at presentation (<5 IU/L)¹²⁴. Intriguingly, rituximab was found to enhance the remission rate of GH in young patients aged 12-20 years when combined with a short course of ATD. In a recent multicenter, single-arm, phase 2 trial, 27 patients received a single 500-mg dose of rituximab and 12 months of ATD titrated according to thyroid function. ATDs were discontinued after 12 months, and the primary outcome was assessed at 24 months using A'Hern design to distinguish an encouraging remission rate (40%) from an unacceptable rate (20%). Thirteen out of 27 patients (48%) achieved GH remission, providing evidence of a promising remission rate. However, the lack of a control group underscores the need for a randomized clinical to validate the efficacy of rituximab¹²⁵.

Tocilizumab

Tocilizumab is a humanized monoclonal antibody that targets IL-6R, representing a valid second-line treatment for moderate-to-severe, active GO²¹. In a small clinical trial, 32 GC-resistant patients with moderate-to-severe, active GO were randomized to receive iv tocilizumab (8mg/kg, four doses with a 4-week interval) or placebo, showing a significantly greater rate of inactivation in patients given tocilizumab, with limited efficacy on exophthalmos and diplopia. However, placebo-treated patients also had a surprisingly high rate of inactivation, suggesting that the improvement was affected by the delayed effect of previous GC therapy or GO natural history¹²⁶. Larger multicenter randomized clinical trials are warranted.

Batoclimab

Batoclimab is a fully human anti-FcRn monoclonal antibody that blocks FcRn-IgG interactions and accelerates autoantibody degradation. Its efficacy has been reported in other IgG-mediated autoimmune diseases¹²⁷. A proof-of-concept randomized trial in patients with GO was recently published¹²⁸. The trial was prematurely interrupted because of an unexpected increase in serum LDL-cholesterol reflecting a reduction in albumin levels. However, the efficacy of batoclimab was evaluated in 65 of the 77 planned patients. All patients who received 6 weekly subcutaneous injections of batoclimab had a significant reduction in TRAbs and orbital muscle volume decreased compared to placebo group. The improvement in exophthalmos and CAS did not reach the statistical significance, probably because of the small number of patients¹²⁸.

Linsitinib

Linsitinib is an orally available dual small-molecule inhibiting IGF-1R. Encouraging results have been recently reported in an experimental GO mouse model¹²⁹. Gulbins et al. showed that linsitinib can prevent hyperthyroidism development when administered in the early phase of the disease. In the chronic phase, linsitinib reduced immune infiltration (T-cell and macrophages) of orbital tissues as well as orbital inflammation¹²⁹. *In vitro* studies have shown that linsitinib, in combination with a small molecule TSH-R inhibitor, ANTAG3, inhibits hyaluronan secretion by cultured OFs¹³⁰. Taken together, these data suggest that linsitinib may be a very promising drug for GO.

Sirolimus

Sirolimus, also known as rapamycin, is an antiproliferative, antifibrotic, immunosuppressive drug used to prevent organ rejection and treat lymphangioliomyomatosis¹³¹. Its molecular target is the serine-threonine kinase known as “mammalian target of rapamycin” (mTOR) which plays a role in regulating cell growth, proliferation, motility, and survival¹³¹. Through mTORC1 inhibition, sirolimus counteracts T cell activation, adipogenesis, and fibroblast migration and transition into myofibroblasts, which are important mechanisms involved in GO¹³¹⁻¹³³. A retrospective investigation has been recently performed to assess the efficacy of sirolimus (2 mg orally on the first day, followed by 0.5 mg/day for 12 weeks) as a second-line therapy in patients with moderate-to-severe active GO compared to methylprednisolone¹³⁴. Thirty patients (15 per group) previously treated with methylprednisolone with no beneficial effects were included. The primary endpoint was the overall outcome of GO at 24 weeks, based on a composite evaluation.

The proportion of GO responders, at 24 weeks was higher in patients treated with sirolimus (86.6%) compared with those given methylprednisolone (26.6%). The GO QoL and the proportion of proptosis (80% vs 13.3%) and CAS responders (86.6% vs 33.3%) were greater in the sirolimus group compared to methylprednisolone group¹³⁴. Given the study design and the small sample size, further randomized clinical trials are needed.

Atorvastatin

Given the knowledge that statins counteract the occurrence of GO in GD^{135,136}, and elevated LDL-C is a risk factor for GO^{137,138}, it is reasonable that statins may improve the response rate in GO. The STAGO study was a phase II randomized clinical trial¹³⁹. The purpose was to assess the efficacy of atorvastatin on the overall response of GO to ivGCs. Eighty-eight patients with moderate-to-severe, active GO and high LDL-C serum levels were included and randomly assigned to statin (ST) or no-statin (NST) treatment. Patients in both groups received ivGCs. Patients in the ST group were treated with atorvastatin 20 mg/daily for 24 weeks. The primary outcome was the overall response of GO at 24 weeks, assessed by a composite evaluation. A greater proportion of GO responders was observed in the ST compared to NST group both at 12 and 24 weeks. Moreover, the improvement in QoL was higher in patients treated with atorvastatin both at 12 and 24 weeks. The incidence of GO relapse was significantly higher in NST. Interestingly, there was no difference in LDL-C levels between responders and non-responders within the statin group at 12 and 24 weeks, suggesting that the beneficial effect of atorvastatin was likely due to its pleiotropic actions rather than to lowering cholesterol¹³⁹.

Conclusions

The evolving landscape of GD and GO demands a closer examination of potential treatment modalities, given the current absence of treatments addressing the underlying disease etiology. Despite advancements in delivery and safety, current treatment methods for GD carry significant limitations, such as high relapse rates upon discontinuation of ATD and the occurrence of hypothyroidism following RAI or surgery. The journey from laboratory discoveries to clinical applications holds promise for transforming the care paradigm. New potential treatments for GD have emerged, employing diverse mechanisms of action. These include the restoration of immune tolerance through immunomodulatory TSH-R peptides and the counteraction of TSH-R signaling using activation with rituximab, iscalimab, and belimumab. As in other autoimmune disease, the

inhibition of IgG recycling through neonatal Fc receptor blockade with rozanolixizumab and efgartigimod may result in a new therapeutic approach in GD as well. Clinical studies on these treatments are predominantly open-label phase 1 and 2 trials. Further randomized controlled studies are warranted to confirm the efficacy and safety of the new compounds.

The occurrence of GO further complicates the treatment landscape, necessitating a nuanced approach. Major advances in the pathogenesis of GO have been made over the last few years, unlocking novel avenues for patient management. Although the FDA has approved teprotumumab as first-line therapy for the treatment of moderate-to-severe GO, its use has not been sanctioned in other countries, where the most commonly used treatment is still methylprednisolone. This discrepancy generates disparities in the guidelines among different scientific societies and in day-to-day clinical practice across the world, making patient management even more complicated.

The increasing understanding of GO pathogenesis has given rise to the suggestion of additional novel medications, such as linsitinib and tocilizumab. The eagerly awaited results from large, randomized multicenter trials will be crucial to confirming the promising yet preliminary findings.

Competing interests

Michele Marinò is a member of the Advisory Board of Horizon Pharma.

'The remaining authors declare no competing interests'

Figure legends

Figure 1

Pathogenesis of Graves' hyperthyroidism and mechanisms of action of the most promising recently proposed therapies

In loss of self-tolerance, thyroid stimulating hormone receptor (TSH-R) peptides are recognized by antigen-presenting cells (APCs), namely B-cells, macrophages, dendritic cells, and thyroid cells. The interaction between T-cell receptor (TCR) and major histocompatibility complex (MHC) promotes activation of T-cells, which in turn activate B-cells through interaction between TCR and MHC and binding of cluster of differentiation 40 (CD40) to its ligand CD40L. Once activated, B-cells differentiate into plasma cells, producing TSH-R autoantibodies (TRAbs), which stimulate thyroid cells to proliferate and secrete hormones, resulting in hyperthyroidism. Five mechanisms and sites of action of novel treatments for Graves' hyperthyroidism are represented. (a) K1-70 is a human monoclonal antibody which binds and blocks TSH-R. (b) Iscalimab is an anti-CD40 monoclonal antibody that blocks CD40-CD40L costimulatory signaling. (c) ATX-GD-59 is an epitope that can prevent TRAbs production and restore immune tolerance. (d) Belimumab is a recombinant, fully human, monoclonal antibody against B-cell activating factor (BAFF), which prevents BAFF from interacting with its receptors. Belimumab inhibits primary humoral immune responses by depleting naïve B-cells that are dependent on BAFF for their survival; (e) Small molecules bind the transmembrane domain of the TSH-R, blocking its activation. Created with BioRender.com

Figure 2

Pathogenesis of Graves' orbitopathy (GO) and mechanisms of action of the most promising recently proposed therapies

The autoimmune reaction against thyroid stimulating hormone receptor (TSH-R) leads to autoreactive T- and B-lymphocytes that infiltrate orbital tissues, triggering fibroblast proliferation and differentiation into adipocytes. Activated CD34⁺ fibroblasts and the autoreactive T- and B-cells promote the release of pro-inflammatory cytokines and glycosaminoglycans, especially hyaluronic acid, contributing the typical connective tissue remodeling. Four mechanisms and sites of action of the novel treatment procedures for GO are represented. (a) Mycophenolate has antiproliferative activity on both T- and B-cells; (b) Rituximab is a chimeric mouse-human monoclonal antibody that causes B-cell depletion by binding cluster of differentiation 20 (CD20),

a cell-surface antigen expressed on their surface; (c) Teprotumumab is a monoclonal antibody against insulin-like growth factor 1 receptor (IGF-1R) that disrupts activation of orbital fibroblasts; (d) Tocilizumab is an anti-interleukin 6 receptor (IL-6) humanized monoclonal antibody. IL-6 is a proinflammatory cytokine overexpressed in orbital fibroblasts and capable, among other actions, of stimulating TSH-R expression in orbital fibroblasts. Created with BioRender.com

Box 1. Graves' disease and Graves' orbitopathy: clinical presentation.

Patients with GH may exhibit various clinical features ranging from subclinical presentations to overt hyperthyroidism. Tachycardia, shortness of breath, tremor, and heat intolerance are the most common symptoms. Atrial fibrillation occurs in ~10% of patients, and elderly patients can experience heart failure. Neurogenic symptoms predominate in patients < 50 years old. GH patients may experience weight loss, and bone turnover can be affected, resulting in osteopenia and osteoporosis. Menstrual bleeding may be light, decreased in frequency, or absent¹⁴⁰.

Although GD primarily involves the thyroid, patients may also experience extrathyroidal manifestations, including GO, acropachy and pretibial myxedema (PTM). PTM occurs in 0.5-4.3 % of patients¹⁴¹ and is a localized skin thickening due to hyaluronan deposition^{17,142}. PTM typically occurs on the lateral or anterior side of the legs, but it can involve other skin surfaces. Clinical manifestations range from nonpitting edema to nodular, plaque, and polypoid forms. About 1% of GD patients are affected by thyroid acropachy, with clubbing of toes and fingers, periosteal proliferation of the phalanges shafts and soft tissue swelling^{17,142}.

The natural history of GO is biphasic, with an active phase lasting 6-18 months followed by an inactive phase¹⁴³. Clinical manifestations of GO can be ascribed to three features: inflammation of soft tissues, glycosaminoglycans overproduction, and increase in adipose tissue. Inflammation and exophthalmos are typical of the early phase, whereas diplopia occurs when inflammation becomes organized resulting in fibrosis^{143,144}. Extraocular muscle enlargement results in impaired ocular motility and diplopia and, in the most severe cases, dysthyroid optic neuropathy (DON) due to compression of the optic nerves at the orbital apex^{144,145}. GO can be classified into active or non-active, based on inflammation extent, commonly assessed by CAS^{21,146}. Severity is established by the extent of proptosis, lid retraction, diplopia, and corneal and optic nerve involvement²¹. Mild GO rarely progresses to more severe forms¹⁴⁷. Sight-threatening GO is quite

rare, due to corneal breakdown (because of severe exophthalmos and/or lagophthalmos) or DON, and requires urgent treatment²¹.

Box 2. Genetic susceptibility to Graves' disease, and environmental risk factors.

The Human leukocyte antigens (HLA) complex increases the risk of GD occurrence by 2-4 fold, contributing 5% of the estimated genetic susceptibility to GD¹⁴⁸. An association between TSH-R gene polymorphism and the risk of GD has been described¹⁴⁹. Several elements involved in the immune checkpoint and immune response regulation, have been also reported to play a role. Forkhead box P3 gene (FOXP3), alternatively referred to as Scurfin, encodes a crucial transcription factor involved in the development of regulatory T (Treg) cells, thereby affecting the etiopathogenesis of autoimmune thyroid diseases. Polymorphisms and abnormal acetylation of FOXP3 gene are associated with the risk of developing GD¹⁵⁰⁻¹⁵². Fc receptor-like protein 3 (FCRL3) protein is involved in the regulation of the immune system by affecting B cell signaling. FCRL3 polymorphisms may increase the susceptibility to GD⁵⁻⁸. Other immune-regulatory genes, including cluster of differentiation 40 (CD-40), protein tyrosine phosphatase non-receptor type 22 (PTPN22), and cytotoxic T lymphocyte antigen-4 (CTLA-4), have been linked to GD⁹, but their impact varies, depending on several factors, especially ethnicity¹⁵³. Through the interaction with CD40 ligand (CD40L), which is expressed by activated CD4⁺ T cells, CD40 is essential to trigger B cells. The CD40-CD40L co-stimulatory pathway plays an important role in the pathogenesis of GD by promoting autoreactive B-cell activation. The CD40 gene is a key susceptibility gene for GD¹⁰. Recently, Bufalo et al. published a case-control study that confirms the crucial role of CTLA4 polymorphisms in GD susceptibility and demonstrates the role of PTPN22 polymorphisms in patients' clinical features, suggesting that these genes may influence the severity of the disease¹².

Sex, psychological stress, iodine, irradiation, infections, post-partum period, radioiodine (RI) treatment for other conditions are recognized risk factors¹⁴⁵. Immune-modulating agents, especially alemtuzumab, have also been reported as potential triggers. Infections, including SARS-CoV-2^{154,155} can induce GD through several mechanisms¹⁵⁶.

Box 3. Animal models of Graves' disease.

TSH-R is extensively preserved within the species, making the immune system highly tolerant. Over the past few decades, several attempts have been made to overcome tolerance and create an appropriate animal model. The most successful have been achieved by injecting TSH-R-expressing adenoviruses or plasmid encoding TSH-R to introduce the human TSH-R extracellular domain into the body. However, these models require repeated immunization thereby producing inconsistent results^{35,157-159}.

In 2023 Bao and colleagues proposed a new Cre-loxP system-based mouse model able to induce the expression of the human TSH-R-A subunit mimicking GD with an 81.25% frequency¹⁶⁰. The Cre-loxP system-based mouse was established by inserting the CAG-loxP-STOP-loxP-hTSH-R A subunit cassette into the Rosa 26 locus of the mouse genome. Conditional expression of the hTSH-R-A subunit was successfully achieved by a single intramuscular injection of the transactivator of transcription-Cre recombinase. Twenty-four (8 control and 16 GD) mice were investigated. The development of hyperthyroidism was investigated by measuring thyroid hormone and TSH-R Ab levels which were significantly higher in GD mice compared to controls at weeks 8 and 12. Histological analysis showed thyroid hypertrophy and diffuse enlargement in GD group. Before euthanasia, mice were subjected to MRI to investigate GO occurrence. Even though there was no significant proptosis, GD mice had a significant increase in retroorbital tissue volume compared to control mice. These findings were confirmed by histology. In addition, F4/80+ staining and Masson's trichrome staining of orbital tissues showed macrophage infiltration and muscle fibrosis in the GD group. This novel mouse model opens new perspectives for investigation in GD and GO animal models¹⁶⁰.

Recently, new immunomodulatory therapies have been also assessed and small cyclic peptides from the antigenic region of the extracellular subunit of the TSH-R were tested to induce immune tolerance and prevent GD occurrence^{161,162}.

Box 4. Genetic susceptibility to Graves' orbitopathy, and environmental risk factors.

Twin and family studies and family clustering indicate that inheritance predisposes to GD¹⁶³. Several genes have been identified and twin studies show a 30% concordance rate, indicating modest penetrance^{4,163}. However, the genetic profile of GD is unclear^{9,164}. Epigenetic factors, namely histone modification, DNA methylation, and noncoding RNAs may contribute GO

pathogenesis⁷¹. In addition, the gut microbiota is believed to play a crucial role in the development of GD and GO. Changes in gut microbiota have been reported in GD patients with or without GO¹³ and perturbation of gut microbiota in GO patients compared to healthy controls, with increased levels of *Lactobacillus*, *Prevotella*, and *Veillonella* often found in GD patients, has opened an intriguing topic^{14,165}. Treatment with the antibiotic vancomycin was shown to be promising in reducing disease severity, whereas fecal material transfer (FMT) from GD/GO patients has been observed to exacerbate the condition in mouse models. Common therapeutic agents for GD and GO can also impact the gut microbiota. Dysbiosis in GD and GO is primarily attributed to antigenic mimicry and an imbalance of T helper 17 cells (Th17) and regulatory T cells (Tregs). Several interventions, such as antibiotics, probiotics, and dietary modifications, have been actively explored in preclinical models and, to some extent, in clinical settings¹⁶⁵. Future research is expected to unveil molecular pathways linking gut and thyroid functions and their influence on orbital autoimmunity. Microbiota-targeting therapeutics are likely to emerge as a crucial strategy in the management of GD/GO in the coming years.

Risk factors associated with GO include sex, age, smoking, radioiodine, and inadequate control of thyroid dysfunction¹⁴⁵. A role of oxidative stress and selenium deficiency has been also reported^{166,167}. A predictive score for the occurrence and progression of GO in GD has been proposed, with determinants such as smoking, duration of hyperthyroidism, the GO CAS and TSH-binding inhibitor immunoglobulins (TBII)¹⁶⁸.

An association between diabetes mellitus (DM) and GO has been recently described. DM enhances systemic inflammation, thereby promoting molecular mechanisms responsible for GO¹⁶⁹. Since 2015, the role of low-density lipoprotein cholesterol (LDL-C) has been investigated. Two large retrospective studies^{135,136} found a lower incidence of GO in GD patients treated with statins. In a cross-sectional investigation, high cholesterol correlated with the occurrence of GO¹³⁷, CAS was higher in patients with high cholesterol, with a correlation between CAS, total and LDL-C¹³⁷. A retrospective analysis confirmed these results¹³⁸. A retrospective investigation demonstrated that high LDL-C was associated with poor response to intravenous glucocorticoid (ivGC)¹⁷⁰. Accordingly, the EUGOGO guidelines suggest correction hypercholesterolemia²¹.

- 1 Smith, T. J. & Hegedus, L. Graves' Disease. *N Engl J Med***375**, 1552-1565, doi:10.1056/NEJMra1510030 (2016).
- 2 Adams, D. D. Pathogenesis of the Hyperthyroidism of Graves's Disease. *Br Med J***1**, 1015-1019, doi:10.1136/bmj.1.5441.1015 (1965).
- 3 Lee, H. J., Li, C. W., Hammerstad, S. S., Stefan, M. & Tomer, Y. Immunogenetics of autoimmune thyroid diseases: A comprehensive review. *J Autoimmun***64**, 82-90, doi:10.1016/j.jaut.2015.07.009 (2015).
- 4 Brix, T. H., Kyvik, K. O., Christensen, K. & Hegedus, L. Evidence for a major role of heredity in Graves' disease: a population-based study of two Danish twin cohorts. *J Clin Endocrinol Metab***86**, 930-934, doi:10.1210/jcem.86.2.7242 (2001).
- 5 Simmonds, M. J. *et al.* Contribution of single nucleotide polymorphisms within FCRL3 and MAP3K7IP2 to the pathogenesis of Graves' disease. *J Clin Endocrinol Metab***91**, 1056-1061, doi:10.1210/jc.2005-1634 (2006).
- 6 Inoue, N. *et al.* Associations between autoimmune thyroid disease prognosis and functional polymorphisms of susceptibility genes, CTLA4, PTPN22, CD40, FCRL3, and ZFAT, previously revealed in genome-wide association studies. *J Clin Immunol***32**, 1243-1252, doi:10.1007/s10875-012-9721-0 (2012).
- 7 Khong, J. J. *et al.* Pooled genome wide association detects association upstream of FCRL3 with Graves' disease. *BMC Genomics***17**, 939, doi:10.1186/s12864-016-3276-z (2016).
- 8 Zhao, S. X. *et al.* A refined study of FCRL genes from a genome-wide association study for Graves' disease. *PLoS One***8**, e57758, doi:10.1371/journal.pone.0057758 (2013).
- 9 Hasham, A. & Tomer, Y. Genetic and epigenetic mechanisms in thyroid autoimmunity. *Immunol Res***54**, 204-213, doi:10.1007/s12026-012-8302-x (2012).
- 10 Faustino, L. C. *et al.* Precision Medicine in Graves' Disease: CD40 Gene Variants Predict Clinical Response to an Anti-CD40 Monoclonal Antibody. *Front Endocrinol (Lausanne)***12**, 691781, doi:10.3389/fendo.2021.691781 (2021).
- 11 Kahaly, G. J. *et al.* A Novel Anti-CD40 Monoclonal Antibody, Iscalimab, for Control of Graves Hyperthyroidism-A Proof-of-Concept Trial. *J Clin Endocrinol Metab***105**, doi:10.1210/clinem/dgz013 (2020).
- 12 Bufalo, N. E. *et al.* Polymorphisms of the genes CTLA4, PTPN22, CD40, and PPARG and their roles in Graves' disease: susceptibility and clinical features. *Endocrine***71**, 104-112, doi:10.1007/s12020-020-02337-x (2021).
- 13 Shi, T. T. *et al.* Alterations in the intestinal microbiota of patients with severe and active Graves' orbitopathy: a cross-sectional study. *J Endocrinol Invest***42**, 967-978, doi:10.1007/s40618-019-1010-9 (2019).
- 14 Biscarini, F. *et al.* Gut Microbiome Associated With Graves Disease and Graves Orbitopathy: The INDIGO Multicenter European Study. *J Clin Endocrinol Metab***108**, 2065-2077, doi:10.1210/clinem/dgad030 (2023).
- 15 Pearce, S. H. S. *et al.* Antigen-Specific Immunotherapy with Thyrotropin Receptor Peptides in Graves' Hyperthyroidism: A Phase I Study. *Thyroid***29**, 1003-1011, doi:10.1089/thy.2019.0036 (2019).

- 16 Chin, Y. H. *et al.* Prevalence of thyroid eye disease in Graves' disease: A meta-analysis and systematic review. *Clin Endocrinol (Oxf)***93**, 363-374, doi:10.1111/cen.14296 (2020).
- 17 Fatourehchi, V. Thyroid dermopathy and acropachy. *Best Pract Res Clin Endocrinol Metab***26**, 553-565, doi:10.1016/j.beem.2011.10.001 (2012).
- 18 Bahn, R. S. Current Insights into the Pathogenesis of Graves' Ophthalmopathy. *Horm Metab Res***47**, 773-778, doi:10.1055/s-0035-1555762 (2015).
- 19 McLeod, D. S. & Cooper, D. S. The incidence and prevalence of thyroid autoimmunity. *Endocrine***42**, 252-265, doi:10.1007/s12020-012-9703-2 (2012).
- 20 Taylor, P. N. *et al.* Global epidemiology of hyperthyroidism and hypothyroidism. *Nat Rev Endocrinol***14**, 301-316, doi:10.1038/nrendo.2018.18 (2018).
- 21 Bartalena, L. *et al.* The 2021 European Group on Graves' orbitopathy (EUGOGO) clinical practice guidelines for the medical management of Graves' orbitopathy. *Eur J Endocrinol***185**, G43-G67, doi:10.1530/EJE-21-0479 (2021).
- 22 Schuh, A. *et al.* Presentation of Graves' orbitopathy within European Group On Graves' Orbitopathy (EUGOGO) centres from 2012 to 2019 (PREGO III). *Br J Ophthalmol*, doi:10.1136/bjo-2022-322442 (2023).
- 23 Tanda, M. L. *et al.* Prevalence and natural history of Graves' orbitopathy in a large series of patients with newly diagnosed graves' hyperthyroidism seen at a single center. *J Clin Endocrinol Metab***98**, 1443-1449, doi:10.1210/jc.2012-3873 (2013).
- 24 Tomer, Y. Mechanisms of autoimmune thyroid diseases: from genetics to epigenetics. *Annu Rev Pathol***9**, 147-156, doi:10.1146/annurev-pathol-012513-104713 (2014).
- 25 Limbach, M. *et al.* Epigenetic profiling in CD4+ and CD8+ T cells from Graves' disease patients reveals changes in genes associated with T cell receptor signaling. *J Autoimmun***67**, 46-56, doi:10.1016/j.jaut.2015.09.006 (2016).
- 26 Kleinau, G. *et al.* Structural-Functional Features of the Thyrotropin Receptor: A Class A G-Protein-Coupled Receptor at Work. *Front Endocrinol (Lausanne)***8**, 86, doi:10.3389/fendo.2017.00086 (2017).
- 27 Mezei, M., Latif, R. & Davies, T. F. Computational model of the full-length TSH receptor. *Elife***11**, doi:10.7554/eLife.81415 (2022).
- 28 Sanders, J., Miguel, R. N., Furmaniak, J. & Smith, B. R. TSH receptor monoclonal antibodies with agonist, antagonist, and inverse agonist activities. *Methods Enzymol***485**, 393-420, doi:10.1016/B978-0-12-381296-4.00022-1 (2010).
- 29 Morris, J. C. *et al.* Identification of epitopes and affinity purification of thyroid stimulating auto-antibodies using synthetic human TSH receptor peptides. *Autoimmunity***17**, 287-299, doi:10.3109/08916939409010669 (1994).
- 30 Tahara, K. *et al.* Epitopes for thyroid stimulating and blocking autoantibodies on the extracellular domain of the human thyrotropin receptor. *Thyroid***7**, 867-877, doi:10.1089/thy.1997.7.867 (1997).
- 31 Weetman, A. P. *et al.* Thyroid-stimulating antibody activity between different immunoglobulin G subclasses. *J Clin Invest***86**, 723-727, doi:10.1172/JCI114768 (1990).
- 32 Nakashima, M., Martin, A. & Davies, T. F. Intrathyroidal T cell accumulation in Graves' disease: delineation of mechanisms based on in situ T cell receptor analysis. *J Clin Endocrinol Metab***81**, 3346-3351, doi:10.1210/jcem.81.9.8784095 (1996).

- 33 Morshed, S. A., Latif, R. & Davies, T. F. Characterization of thyrotropin receptor antibody-induced signaling cascades. *Endocrinology***150**, 519-529, doi:10.1210/en.2008-0878 (2009).
- 34 Nagayama, Y., Wadsworth, H. L., Russo, D., Chazenbalk, G. D. & Rapoport, B. Binding domains of stimulatory and inhibitory thyrotropin (TSH) receptor autoantibodies determined with chimeric TSH-lutropin/chorionic gonadotropin receptors. *J Clin Invest***88**, 336-340, doi:10.1172/JCI115297 (1991).
- 35 Chazenbalk, G. D. *et al.* Thyroid-stimulating autoantibodies in Graves disease preferentially recognize the free A subunit, not the thyrotropin holoreceptor. *J Clin Invest***110**, 209-217, doi:10.1172/JCI15745 (2002).
- 36 Latif, R., Morshed, S. A., Zaidi, M. & Davies, T. F. The thyroid-stimulating hormone receptor: impact of thyroid-stimulating hormone and thyroid-stimulating hormone receptor antibodies on multimerization, cleavage, and signaling. *Endocrinol Metab Clin North Am***38**, 319-341, viii, doi:10.1016/j.ecl.2009.01.006 (2009).
- 37 Rapoport, B., Aliesky, H. A., Chen, C. R. & McLachlan, S. M. Evidence that TSH Receptor A-Subunit Multimers, Not Monomers, Drive Antibody Affinity Maturation in Graves' Disease. *J Clin Endocrinol Metab***100**, E871-875, doi:10.1210/jc.2015-1528 (2015).
- 38 McLachlan, S. M. & Rapoport, B. Thyrotropin-blocking autoantibodies and thyroid-stimulating autoantibodies: potential mechanisms involved in the pendulum swinging from hypothyroidism to hyperthyroidism or vice versa. *Thyroid***23**, 14-24, doi:10.1089/thy.2012.0374 (2013).
- 39 Kasagi, K. *et al.* Fluctuating thyroid function depending on the balance between stimulating and blocking types of TSH receptor antibodies: a case report. *Thyroid***3**, 315-318, doi:10.1089/thy.1993.3.315 (1993).
- 40 Mao, C. *et al.* Impairment of regulatory capacity of CD4+CD25+ regulatory T cells mediated by dendritic cell polarization and hyperthyroidism in Graves' disease. *J Immunol***186**, 4734-4743, doi:10.4049/jimmunol.0904135 (2011).
- 41 Pan, D., Shin, Y. H., Gopalakrishnan, G., Hennessey, J. & De Groot, L. J. Regulatory T cells in Graves' disease. *Clin Endocrinol (Oxf)***71**, 587-593, doi:10.1111/j.1365-2265.2009.03544.x (2009).
- 42 Xing, Y. & Hogquist, K. A. T-cell tolerance: central and peripheral. *Cold Spring Harb Perspect Biol***4**, doi:10.1101/cshperspect.a006957 (2012).
- 43 Martin, A., Schwartz, A. E., Friedman, E. W. & Davies, T. F. Successful production of intrathyroidal human T cell hybridomas: evidence for intact helper T cell function in Graves' disease. *J Clin Endocrinol Metab***69**, 1104-1108, doi:10.1210/jcem-69-6-1104 (1989).
- 44 Rapoport, B. & McLachlan, S. M. Graves' hyperthyroidism is antibody-mediated but is predominantly a Th1-type cytokine disease. *J Clin Endocrinol Metab***99**, 4060-4061, doi:10.1210/jc.2014-3011 (2014).
- 45 ElTanbouly, M. A. *et al.* VISTA is a checkpoint regulator for naive T cell quiescence and peripheral tolerance. *Science***367**, doi:10.1126/science.aay0524 (2020).
- 46 Armengol, M. P. *et al.* Chemokines determine local lymphoneogenesis and a reduction of circulating CXCR4+ T and CCR7 B and T lymphocytes in thyroid autoimmune diseases. *J Immunol***170**, 6320-6328, doi:10.4049/jimmunol.170.12.6320 (2003).

- 47 Fernando, R. *et al.* Human fibrocytes coexpress thyroglobulin and thyrotropin receptor. *Proc Natl Acad Sci U S A***109**, 7427-7432, doi:10.1073/pnas.1202064109 (2012).
- 48 Feliciello, A. *et al.* Expression of thyrotropin-receptor mRNA in healthy and Graves' disease retro-orbital tissue. *Lancet***342**, 337-338, doi:10.1016/0140-6736(93)91475-2 (1993).
- 49 Hai, Y. P., Lee, A. C. H., Frommer, L., Diana, T. & Kahaly, G. J. Immunohistochemical analysis of human orbital tissue in Graves' orbitopathy. *J Endocrinol Invest***43**, 123-137, doi:10.1007/s40618-019-01116-4 (2020).
- 50 Kumar, S., Nadeem, S., Stan, M. N., Coenen, M. & Bahn, R. S. A stimulatory TSH receptor antibody enhances adipogenesis via phosphoinositide 3-kinase activation in orbital preadipocytes from patients with Graves' ophthalmopathy. *J Mol Endocrinol***46**, 155-163, doi:10.1530/JME-11-0006 (2011).
- 51 Zhang, M. *et al.* A Promising Mouse Model of Graves' Orbitopathy Induced by Adenovirus Expressing Thyrotropin Receptor A Subunit. *Thyroid***31**, 638-648, doi:10.1089/thy.2020.0088 (2021).
- 52 Holthoff, H. P. *et al.* Prolonged TSH receptor A subunit immunization of female mice leads to a long-term model of Graves' disease, tachycardia, and cardiac hypertrophy. *Endocrinology***156**, 1577-1589, doi:10.1210/en.2014-1813 (2015).
- 53 Moshkelgosha, S., So, P. W., Deasy, N., Diaz-Cano, S. & Banga, J. P. Cutting edge: retrobulbar inflammation, adipogenesis, and acute orbital congestion in a preclinical female mouse model of Graves' orbitopathy induced by thyrotropin receptor plasmid-in vivo electroporation. *Endocrinology***154**, 3008-3015, doi:10.1210/en.2013-1576 (2013).
- 54 Gerding, M. N. *et al.* Association of thyrotrophin receptor antibodies with the clinical features of Graves' ophthalmopathy. *Clin Endocrinol (Oxf)***52**, 267-271, doi:10.1046/j.1365-2265.2000.00959.x (2000).
- 55 Nicoli, F. *et al.* Correlation between serum anti-TSH receptor autoantibodies (TRAbs) and the clinical feature of Graves' orbitopathy. *J Endocrinol Invest***44**, 581-585, doi:10.1007/s40618-020-01353-y (2021).
- 56 Eckstein, A. K. *et al.* Thyrotropin receptor autoantibodies are independent risk factors for Graves' ophthalmopathy and help to predict severity and outcome of the disease. *J Clin Endocrinol Metab***91**, 3464-3470, doi:10.1210/jc.2005-2813 (2006).
- 57 Douglas, R. S. *et al.* Teprotumumab for the Treatment of Active Thyroid Eye Disease. *N Engl J Med***382**, 341-352, doi:10.1056/NEJMoa1910434 (2020).
- 58 Pritchard, J., Han, R., Horst, N., Cruikshank, W. W. & Smith, T. J. Immunoglobulin activation of T cell chemoattractant expression in fibroblasts from patients with Graves' disease is mediated through the insulin-like growth factor I receptor pathway. *J Immunol***170**, 6348-6354, doi:10.4049/jimmunol.170.12.6348 (2003).
- 59 Douglas, R. S. *et al.* B cells from patients with Graves' disease aberrantly express the IGF-1 receptor: implications for disease pathogenesis. *J Immunol***181**, 5768-5774, doi:10.4049/jimmunol.181.8.5768 (2008).
- 60 Douglas, R. S., Gianoukakis, A. G., Kamat, S. & Smith, T. J. Aberrant expression of the insulin-like growth factor-1 receptor by T cells from patients with Graves' disease may carry functional consequences for disease pathogenesis. *J Immunol***178**, 3281-3287, doi:10.4049/jimmunol.178.5.3281 (2007).

- 61 Smith, T. J. & Hoa, N. Immunoglobulins from patients with Graves' disease induce hyaluronan synthesis in their orbital fibroblasts through the self-antigen, insulin-like growth factor-I receptor. *J Clin Endocrinol Metab***89**, 5076-5080, doi:10.1210/jc.2004-0716 (2004).
- 62 Tsui, S. *et al.* Evidence for an association between thyroid-stimulating hormone and insulin-like growth factor 1 receptors: a tale of two antigens implicated in Graves' disease. *J Immunol***181**, 4397-4405, doi:10.4049/jimmunol.181.6.4397 (2008).
- 63 Minich, W. B. *et al.* Autoantibodies to the IGF1 receptor in Graves' orbitopathy. *J Clin Endocrinol Metab***98**, 752-760, doi:10.1210/jc.2012-1771 (2013).
- 64 Vwarewijk, A. J. *et al.* Circulating IgGs may modulate IGF-I receptor stimulating activity in a subset of patients with Graves' ophthalmopathy. *J Clin Endocrinol Metab***98**, 769-776, doi:10.1210/jc.2012-2270 (2013).
- 65 Lanzolla, G. *et al.* Putative protective role of autoantibodies against the insulin-like growth factor-1 receptor in Graves' Disease: results of a pilot study. *J Endocrinol Invest***43**, 1759-1768, doi:10.1007/s40618-020-01341-2 (2020).
- 66 Aniszewski, J. P., Valyasevi, R. W. & Bahn, R. S. Relationship between disease duration and predominant orbital T cell subset in Graves' ophthalmopathy. *J Clin Endocrinol Metab***85**, 776-780, doi:10.1210/jcem.85.2.6333 (2000).
- 67 Fang, S. *et al.* Regulation of Orbital Fibrosis and Adipogenesis by Pathogenic Th17 Cells in Graves Orbitopathy. *J Clin Endocrinol Metab***102**, 4273-4283, doi:10.1210/jc.2017-01349 (2017).
- 68 Ma, R. *et al.* PH20 Inhibits TGFbeta1-Induced Differentiation of Perimysial Orbital Fibroblasts via Hyaluronan-CD44 Pathway in Thyroid-Associated Ophthalmopathy. *Invest Ophthalmol Vis Sci***60**, 1431-1441, doi:10.1167/iovs.18-26268 (2019).
- 69 Rotondo Dottore, G. *et al.* Association of T and B Cells Infiltrating Orbital Tissues With Clinical Features of Graves Orbitopathy. *JAMA Ophthalmol***136**, 613-619, doi:10.1001/jamaophthalmol.2018.0806 (2018).
- 70 Bahn, R. S. Graves' ophthalmopathy. *N Engl J Med***362**, 726-738, doi:10.1056/NEJMra0905750 (2010).
- 71 Rotondo Dottore, G. *et al.* Insights Into the Role of DNA Methylation and Gene Expression in Graves Orbitopathy. *J Clin Endocrinol Metab***108**, e160-e168, doi:10.1210/clinem/dgac645 (2023).
- 72 Martinez-Hernandez, R. *et al.* A MicroRNA Signature for Evaluation of Risk and Severity of Autoimmune Thyroid Diseases. *J Clin Endocrinol Metab***103**, 1139-1150, doi:10.1210/jc.2017-02318 (2018).
- 73 Shahraki, K. *et al.* Non-coding RNA-mediated epigenetic alterations in Grave's ophthalmopathy: A scoping systematic review. *Noncoding RNA Res***8**, 426-450, doi:10.1016/j.ncrna.2023.04.001 (2023).
- 74 Jang, S. Y. *et al.* Role of microRNA-146a in regulation of fibrosis in orbital fibroblasts from patients with Graves' orbitopathy. *Br J Ophthalmol***102**, 407-414, doi:10.1136/bjophthalmol-2017-310723 (2018).
- 75 De Leo, S., Lee, S. Y. & Braverman, L. E. Hyperthyroidism. *Lancet***388**, 906-918, doi:10.1016/S0140-6736(16)00278-6 (2016).

- 76 Sundaresh, V. *et al.* Comparative effectiveness of therapies for Graves' hyperthyroidism: a systematic review and network meta-analysis. *J Clin Endocrinol Metab***98**, 3671-3677, doi:10.1210/jc.2013-1954 (2013).
- 77 Cooper, D. S. Antithyroid drugs. *N Engl J Med***352**, 905-917, doi:10.1056/NEJMra042972 (2005).
- 78 Abraham, P., Avenell, A., McGeoch, S. C., Clark, L. F. & Bevan, J. S. Antithyroid drug regimen for treating Graves' hyperthyroidism. *Cochrane Database Syst Rev***2010**, CD003420, doi:10.1002/14651858.CD003420.pub4 (2010).
- 79 Struja, T. *et al.* Can we predict relapse in Graves' disease? Results from a systematic review and meta-analysis. *Eur J Endocrinol***176**, 87-97, doi:10.1530/EJE-16-0725 (2017).
- 80 Ross, D. S. *et al.* 2016 American Thyroid Association Guidelines for Diagnosis and Management of Hyperthyroidism and Other Causes of Thyrotoxicosis. *Thyroid***26**, 1343-1421, doi:10.1089/thy.2016.0229 (2016).
- 81 Bartalena, L. *et al.* Use of corticosteroids to prevent progression of Graves' ophthalmopathy after radioiodine therapy for hyperthyroidism. *N Engl J Med***321**, 1349-1352, doi:10.1056/NEJM198911163212001 (1989).
- 82 Ross, D. S. Radioiodine therapy for hyperthyroidism. *N Engl J Med***364**, 542-550, doi:10.1056/NEJMct1007101 (2011).
- 83 Bartalena, L., Burch, H. B., Burman, K. D. & Kahaly, G. J. A 2013 European survey of clinical practice patterns in the management of Graves' disease. *Clin Endocrinol (Oxf)***84**, 115-120, doi:10.1111/cen.12688 (2016).
- 84 Ruslan, A. & Okosieme, O. E. Non-thionamide antithyroid drug options in Graves' hyperthyroidism. *Expert Rev Endocrinol Metab***18**, 67-79, doi:10.1080/17446651.2023.2167709 (2023).
- 85 Lane, L. C., Cheetham, T. D., Perros, P. & Pearce, S. H. S. New Therapeutic Horizons for Graves' Hyperthyroidism. *Endocr Rev***41**, 873-884, doi:10.1210/endrev/bnaa022 (2020).
- 86 Larche, M. & Wraith, D. C. Peptide-based therapeutic vaccines for allergic and autoimmune diseases. *Nat Med***11**, S69-76, doi:10.1038/nm1226 (2005).
- 87 Jansson, L., Vrolix, K., Jahraus, A., Martin, K. F. & Wraith, D. C. Immunotherapy With Apitopes Blocks the Immune Response to TSH Receptor in HLA-DR Transgenic Mice. *Endocrinology***159**, 3446-3457, doi:10.1210/en.2018-00306 (2018).
- 88 Evans, M. *et al.* Monoclonal autoantibodies to the TSH receptor, one with stimulating activity and one with blocking activity, obtained from the same blood sample. *Clin Endocrinol (Oxf)***73**, 404-412, doi:10.1111/j.1365-2265.2010.03831.x (2010).
- 89 Furmaniak, J., Sanders, J. & Rees Smith, B. Blocking type TSH receptor antibodies. *Auto Immun Highlights***4**, 11-26, doi:10.1007/s13317-012-0028-1 (2013).
- 90 Furmaniak, J., Sanders, J., Sanders, P., Li, Y. & Rees Smith, B. TSH receptor specific monoclonal autoantibody K1-70(TM) targeting of the TSH receptor in subjects with Graves' disease and Graves' orbitopathy-Results from a phase I clinical trial. *Clin Endocrinol (Oxf)***96**, 878-887, doi:10.1111/cen.14681 (2022).
- 91 Neumann, S. *et al.* A new small-molecule antagonist inhibits Graves' disease antibody activation of the TSH receptor. *J Clin Endocrinol Metab***96**, 548-554, doi:10.1210/jc.2010-1935 (2011).

- 92 Neumann, S. *et al.* A selective TSH receptor antagonist inhibits stimulation of thyroid function in female mice. *Endocrinology***155**, 310-314, doi:10.1210/en.2013-1835 (2014).
- 93 Latif, R., Realubit, R. B., Karan, C., Mezei, M. & Davies, T. F. TSH Receptor Signaling Abrogation by a Novel Small Molecule. *Front Endocrinol (Lausanne)***7**, 130, doi:10.3389/fendo.2016.00130 (2016).
- 94 Marcinkowski, P. *et al.* A New Highly Thyrotropin Receptor-Selective Small-Molecule Antagonist with Potential for the Treatment of Graves' Orbitopathy. *Thyroid***29**, 111-123, doi:10.1089/thy.2018.0349 (2019).
- 95 Munroe, M. E. Functional roles for T cell CD40 in infection and autoimmune disease: the role of CD40 in lymphocyte homeostasis. *Semin Immunol***21**, 283-288, doi:10.1016/j.smim.2009.05.008 (2009).
- 96 Carayanniotis, G., Masters, S. R. & Noelle, R. J. Suppression of murine thyroiditis via blockade of the CD40-CD40L interaction. *Immunology***90**, 421-426, doi:10.1111/j.1365-2567.1997.00421.x (1997).
- 97 Huber, A. K. *et al.* Genetically driven target tissue overexpression of CD40: a novel mechanism in autoimmune disease. *J Immunol***189**, 3043-3053, doi:10.4049/jimmunol.1200311 (2012).
- 98 Lin, J. D. *et al.* Serum BAFF and thyroid autoantibodies in autoimmune thyroid disease. *Clin Chim Acta***462**, 96-102, doi:10.1016/j.cca.2016.09.004 (2016).
- 99 Lane, L. C. *et al.* Analysis of BAFF gene polymorphisms in UK Graves' disease patients. *Clin Endocrinol (Oxf)***90**, 170-174, doi:10.1111/cen.13872 (2019).
- 100 Campi, I. *et al.* B Cell Activating Factor (BAFF) and BAFF Receptor Expression in Autoimmune and Nonautoimmune Thyroid Diseases. *Thyroid***25**, 1043-1049, doi:10.1089/thy.2015.0029 (2015).
- 101 Gilbert, J. A. *et al.* Treatment of autoimmune hyperthyroidism in a murine model of Graves' disease with tumor necrosis factor-family ligand inhibitors suggests a key role for B cell activating factor in disease pathology. *Endocrinology***147**, 4561-4568, doi:10.1210/en.2006-0507 (2006).
- 102 Allison, A. C. Mechanisms of action of mycophenolate mofetil. *Lupus***14 Suppl 1**, s2-8, doi:10.1191/0961203305lu2109oa (2005).
- 103 Kahaly, G. J. *et al.* Mycophenolate plus methylprednisolone versus methylprednisolone alone in active, moderate-to-severe Graves' orbitopathy (MINGO): a randomised, observer-masked, multicentre trial. *Lancet Diabetes Endocrinol***6**, 287-298, doi:10.1016/S2213-8587(18)30020-2 (2018).
- 104 Marcocci, C. *et al.* Selenium and the course of mild Graves' orbitopathy. *N Engl J Med***364**, 1920-1931, doi:10.1056/NEJMoa1012985 (2011).
- 105 Smith, T. J. *et al.* Teprotumumab for Thyroid-Associated Ophthalmopathy. *N Engl J Med***376**, 1748-1761, doi:10.1056/NEJMoa1614949 (2017).
- 106 Kahaly, G. J., Douglas, R. S., Holt, R. J., Sile, S. & Smith, T. J. Teprotumumab for patients with active thyroid eye disease: a pooled data analysis, subgroup analyses, and off-treatment follow-up results from two randomised, double-masked, placebo-controlled, multicentre trials. *Lancet Diabetes Endocrinol***9**, 360-372, doi:10.1016/S2213-8587(21)00056-5 (2021).

- 107 Douglas, R. S. *et al.* Teprotumumab Efficacy, Safety, and Durability in Longer-Duration Thyroid Eye Disease and Re-treatment: OPTIC-X Study. *Ophthalmology***129**, 438-449, doi:10.1016/j.ophtha.2021.10.017 (2022).
- 108 Burch, H. B. *et al.* Management of Thyroid Eye Disease: A Consensus Statement by the American Thyroid Association and the European Thyroid Association. *Thyroid***32**, 1439-1470, doi:10.1089/thy.2022.0251 (2022).
- 109 Amarikwa, L., Mohamed, A., Kim, S. H., Kossler, A. L. & Dosiou, C. Teprotumumab-Related Hyperglycemia. *J Clin Endocrinol Metab***108**, 858-864, doi:10.1210/clinem/dgac627 (2023).
- 110 Bartalena, L., Marino, M., Marcocci, C. & Tanda, M. L. Teprotumumab for Graves' orbitopathy and ototoxicity: moving problems from eyes to ears? *J Endocrinol Invest***45**, 1455-1457, doi:10.1007/s40618-022-01791-w (2022).
- 111 Safo, M. B. & Silkiss, R. Z. A case of ulcerative colitis associated with teprotumumab treatment for thyroid eye disease. *Am J Ophthalmol Case Rep***22**, 101069, doi:10.1016/j.ajoc.2021.101069 (2021).
- 112 Ashraf, D. C. *et al.* New-Onset of Inflammatory Bowel Disease in a Patient Treated With Teprotumumab for Thyroid Associated Ophthalmopathy. *Ophthalmic Plast Reconstr Surg***37**, e160-e164, doi:10.1097/IOP.0000000000001943 (2021).
- 113 Hoang, T. D., Nguyen, N. T., Chou, E. & Shakir, M. K. Rapidly progressive cognitive decline associated with teprotumumab in thyroid eye disease. *BMJ Case Rep***14**, doi:10.1136/bcr-2021-242153 (2021).
- 114 Reff, M. E. *et al.* Depletion of B cells in vivo by a chimeric mouse human monoclonal antibody to CD20. *Blood***83**, 435-445 (1994).
- 115 McCoy, A. N. *et al.* Rituximab (Rituxan) therapy for severe thyroid-associated ophthalmopathy diminishes IGF-1R(+) T cells. *J Clin Endocrinol Metab***99**, E1294-1299, doi:10.1210/jc.2013-3207 (2014).
- 116 Salvi, M. *et al.* Treatment of Graves' disease and associated ophthalmopathy with the anti-CD20 monoclonal antibody rituximab: an open study. *Eur J Endocrinol***156**, 33-40, doi:10.1530/eje.1.02325 (2007).
- 117 Khanna, D. *et al.* Rituximab treatment of patients with severe, corticosteroid-resistant thyroid-associated ophthalmopathy. *Ophthalmology***117**, 133-139 e132, doi:10.1016/j.ophtha.2009.05.029 (2010).
- 118 Stan, M. N. *et al.* Randomized controlled trial of rituximab in patients with Graves' orbitopathy. *J Clin Endocrinol Metab***100**, 432-441, doi:10.1210/jc.2014-2572 (2015).
- 119 Salvi, M. *et al.* Efficacy of B-cell targeted therapy with rituximab in patients with active moderate to severe Graves' orbitopathy: a randomized controlled study. *J Clin Endocrinol Metab***100**, 422-431, doi:10.1210/jc.2014-3014 (2015).
- 120 Stan, M. N. & Salvi, M. MANAGEMENT OF ENDOCRINE DISEASE: Rituximab therapy for Graves' orbitopathy - lessons from randomized control trials. *Eur J Endocrinol***176**, R101-R109, doi:10.1530/EJE-16-0552 (2017).
- 121 Chen, J., Chen, G. & Sun, H. Intravenous rituximab therapy for active Graves' ophthalmopathy: a meta-analysis. *Hormones (Athens)***20**, 279-286, doi:10.1007/s42000-021-00282-6 (2021).

- 122 Vannucchi, G. *et al.* Efficacy Profile and Safety of Very Low-Dose Rituximab in Patients with Graves' Orbitopathy. *Thyroid***31**, 821-828, doi:10.1089/thy.2020.0269 (2021).
- 123 Heemstra, K. A. *et al.* Rituximab in relapsing Graves' disease, a phase II study. *Eur J Endocrinol***159**, 609-615, doi:10.1530/EJE-08-0084 (2008).
- 124 El Fassi, D., Nielsen, C. H., Bonnema, S. J., Hasselbalch, H. C. & Hegedus, L. B lymphocyte depletion with the monoclonal antibody rituximab in Graves' disease: a controlled pilot study. *J Clin Endocrinol Metab***92**, 1769-1772, doi:10.1210/jc.2006-2388 (2007).
- 125 Cheetham, T. D. *et al.* Adjuvant Rituximab-Exploratory Trial in Young People With Graves Disease. *J Clin Endocrinol Metab***107**, 743-754, doi:10.1210/clinem/dgab763 (2022).
- 126 Perez-Moreiras, J. V. *et al.* Efficacy of Tocilizumab in Patients With Moderate-to-Severe Corticosteroid-Resistant Graves Orbitopathy: A Randomized Clinical Trial. *Am J Ophthalmol***195**, 181-190, doi:10.1016/j.ajo.2018.07.038 (2018).
- 127 Menon, D. & Bril, V. Pharmacotherapy of Generalized Myasthenia Gravis with Special Emphasis on Newer Biologicals. *Drugs***82**, 865-887, doi:10.1007/s40265-022-01726-y (2022).
- 128 Kahaly, G. J. *et al.* Proof-of-Concept and Randomized, Placebo-Controlled Trials of an Fc γ n Inhibitor, Batoclimab, for Thyroid Eye Disease. *J Clin Endocrinol Metab*, doi:10.1210/clinem/dgad381 (2023).
- 129 Gulbins, A. *et al.* Linsitinib, an IGF-1R inhibitor, attenuates disease development and progression in a model of thyroid eye disease. *Front Endocrinol (Lausanne)***14**, 1211473, doi:10.3389/fendo.2023.1211473 (2023).
- 130 Place, R. F., Krieger, C. C., Neumann, S. & Gershengorn, M. C. Inhibiting thyrotropin/insulin-like growth factor 1 receptor crosstalk to treat Graves' ophthalmopathy: studies in orbital fibroblasts in vitro. *Br J Pharmacol***174**, 328-340, doi:10.1111/bph.13693 (2017).
- 131 Sehgal, S. N. Sirolimus: its discovery, biological properties, and mechanism of action. *Transplant Proc***35**, 7S-14S, doi:10.1016/s0041-1345(03)00211-2 (2003).
- 132 Zhang, L. *et al.* Possible targets for nonimmunosuppressive therapy of Graves' orbitopathy. *J Clin Endocrinol Metab***99**, E1183-1190, doi:10.1210/jc.2013-4182 (2014).
- 133 Roos, J. C. P., Eglitis, V. & Murthy, R. Inhibition of Fibrotic Contraction by Sirolimus (Rapamycin) in an Ex Vivo Model of Thyroid Eye Disease. *Ophthalmic Plast Reconstr Surg***37**, 366-371, doi:10.1097/IOP.0000000000001876 (2021).
- 134 Lanzolla, G. *et al.* Sirolimus as a second-line treatment for Graves' orbitopathy. *J Endocrinol Invest***45**, 2171-2180, doi:10.1007/s40618-022-01862-y (2022).
- 135 Stein, J. D. *et al.* Risk factors for developing thyroid-associated ophthalmopathy among individuals with Graves disease. *JAMA Ophthalmol***133**, 290-296, doi:10.1001/jamaophthalmol.2014.5103 (2015).
- 136 Nilsson, A., Tsoumani, K. & Planck, T. Statins Decrease the Risk of Orbitopathy in Newly Diagnosed Patients with Graves Disease. *J Clin Endocrinol Metab***106**, 1325-1332, doi:10.1210/clinem/dgab070 (2021).
- 137 Sabini, E. *et al.* High Serum Cholesterol Is a Novel Risk Factor for Graves' Orbitopathy: Results of a Cross-Sectional Study. *Thyroid***28**, 386-394, doi:10.1089/thy.2017.0430 (2018).

- 138 Lanzolla, G. *et al.* Relationship between serum cholesterol and Graves' orbitopathy (GO): a confirmatory study. *J Endocrinol Invest***41**, 1417-1423, doi:10.1007/s40618-018-0915-z (2018).
- 139 Lanzolla, G. *et al.* Statins for Graves' orbitopathy (STAGO): a phase 2, open-label, adaptive, single centre, randomised clinical trial. *Lancet Diabetes Endocrinol***9**, 733-742, doi:10.1016/S2213-8587(21)00238-2 (2021).
- 140 Davies, T. F. *et al.* Graves' disease. *Nat Rev Dis Primers***6**, 52, doi:10.1038/s41572-020-0184-y (2020).
- 141 Kriss, J. P. Pathogenesis and treatment of pretibial myxedema. *Endocrinol Metab Clin North Am***16**, 409-415 (1987).
- 142 Fatourechi, V., Pajouhi, M. & Fransway, A. F. Dermopathy of Graves disease (pretibial myxedema). Review of 150 cases. *Medicine (Baltimore)***73**, 1-7, doi:10.1097/00005792-199401000-00001 (1994).
- 143 Bartley, G. B. Rundle and his curve. *Arch Ophthalmol***129**, 356-358, doi:10.1001/archophthalmol.2011.29 (2011).
- 144 Bartalena, L. & Fatourechi, V. Extrathyroidal manifestations of Graves' disease: a 2014 update. *J Endocrinol Invest***37**, 691-700, doi:10.1007/s40618-014-0097-2 (2014).
- 145 Bartalena, L. & Tanda, M. L. Current concepts regarding Graves' orbitopathy. *J Intern Med***292**, 692-716, doi:10.1111/joim.13524 (2022).
- 146 Mourits, M. P., Prummel, M. F., Wiersinga, W. M. & Koornneef, L. Clinical activity score as a guide in the management of patients with Graves' ophthalmopathy. *Clin Endocrinol (Oxf)***47**, 9-14, doi:10.1046/j.1365-2265.1997.2331047.x (1997).
- 147 Perros, P., Crombie, A. L. & Kendall-Taylor, P. Natural history of thyroid associated ophthalmopathy. *Clin Endocrinol (Oxf)***42**, 45-50, doi:10.1111/j.1365-2265.1995.tb02597.x (1995).
- 148 Tomer, Y., Barbesino, G., Greenberg, D. A., Concepcion, E. & Davies, T. F. Linkage analysis of candidate genes in autoimmune thyroid disease. III. Detailed analysis of chromosome 14 localizes Graves' disease-1 (GD-1) close to multinodular goiter-1 (MNG-1). International Consortium for the Genetics of Autoimmune Thyroid Disease. *J Clin Endocrinol Metab***83**, 4321-4327, doi:10.1210/jcem.83.12.5343 (1998).
- 149 Bufalo, N. E. *et al.* TSHR intronic polymorphisms (rs179247 and rs12885526) and their role in the susceptibility of the Brazilian population to Graves' disease and Graves' ophthalmopathy. *J Endocrinol Invest***38**, 555-561, doi:10.1007/s40618-014-0228-9 (2015).
- 150 Li, H. N., Li, X. R., Du, Y. Y., Yang, Z. F. & Lv, Z. T. The Association Between Foxp3 Polymorphisms and Risk of Graves' Disease: A Systematic Review and Meta-Analysis of Observational Studies. *Front Endocrinol (Lausanne)***11**, 392, doi:10.3389/fendo.2020.00392 (2020).
- 151 Shehjar, F., Afroze, D., Misgar, R. A., Malik, S. A. & Laway, B. A. Association of FoxP3 promoter polymorphisms with the risk of Graves' disease in ethnic Kashmiri population. *Gene***672**, 88-92, doi:10.1016/j.gene.2018.06.023 (2018).
- 152 Zhang, D. *et al.* MiR-23a-3p-regulated abnormal acetylation of FOXP3 induces regulatory T cell function defect in Graves' disease. *Biol Chem***400**, 639-650, doi:10.1515/hsz-2018-0343 (2019).

- 153 Simmonds, M. J. GWAS in autoimmune thyroid disease: redefining our understanding of pathogenesis. *Nat Rev Endocrinol***9**, 277-287, doi:10.1038/nrendo.2013.56 (2013).
- 154 Mateu-Salat, M., Urgell, E. & Chico, A. SARS-COV-2 as a trigger for autoimmune disease: report of two cases of Graves' disease after COVID-19. *J Endocrinol Invest***43**, 1527-1528, doi:10.1007/s40618-020-01366-7 (2020).
- 155 Lanzolla, G., Marcocci, C. & Marino, M. Graves' disease and Graves' orbitopathy following COVID-19. *J Endocrinol Invest***44**, 2011-2012, doi:10.1007/s40618-021-01576-7 (2021).
- 156 Davies, T. F. Infection and autoimmune thyroid disease. *J Clin Endocrinol Metab***93**, 674-676, doi:10.1210/jc.2008-0095 (2008).
- 157 Wiesweg, B., Johnson, K. T., Eckstein, A. K. & Berchner-Pfannschmidt, U. Current insights into animal models of Graves' disease and orbitopathy. *Horm Metab Res***45**, 549-555, doi:10.1055/s-0033-1343451 (2013).
- 158 Nagayama, Y. *et al.* A novel murine model of Graves' hyperthyroidism with intramuscular injection of adenovirus expressing the thyrotropin receptor. *J Immunol***168**, 2789-2794, doi:10.4049/jimmunol.168.6.2789 (2002).
- 159 Costagliola, S. *et al.* Genetic immunization of outbred mice with thyrotropin receptor cDNA provides a model of Graves' disease. *J Clin Invest***105**, 803-811, doi:10.1172/JCI7665 (2000).
- 160 Bao, Y. *et al.* Cre-loxP System-Based Mouse Model for Investigating Graves' Disease and Associated Orbitopathy. *Thyroid*, doi:10.1089/thy.2023.0299 (2023).
- 161 Fassbender, J., Holthoff, H. P., Li, Z. & Ungerer, M. Therapeutic Effects of Short Cyclic and Combined Epitope Peptides in a Long-Term Model of Graves' Disease and Orbitopathy. *Thyroid***29**, 258-267, doi:10.1089/thy.2018.0326 (2019).
- 162 Holthoff, H. P. *et al.* Cyclic Peptides for Effective Treatment in a Long-Term Model of Graves Disease and Orbitopathy in Female Mice. *Endocrinology***158**, 2376-2390, doi:10.1210/en.2016-1845 (2017).
- 163 Brix, T. H. & Hegedus, L. Twin studies as a model for exploring the aetiology of autoimmune thyroid disease. *Clin Endocrinol (Oxf)***76**, 457-464, doi:10.1111/j.1365-2265.2011.04318.x (2012).
- 164 Yin, X., Latif, R., Bahn, R. & Davies, T. F. Genetic profiling in Graves' disease: further evidence for lack of a distinct genetic contribution to Graves' ophthalmopathy. *Thyroid***22**, 730-736, doi:10.1089/thy.2012.0007 (2012).
- 165 Hou, J., Tang, Y., Chen, Y. & Chen, D. The Role of the Microbiota in Graves' Disease and Graves' Orbitopathy. *Front Cell Infect Microbiol***11**, 739707, doi:10.3389/fcimb.2021.739707 (2021).
- 166 Lanzolla, G., Marcocci, C. & Marino, M. Oxidative Stress in Graves Disease and Graves Orbitopathy. *Eur Thyroid J***9**, 40-50, doi:10.1159/000509615 (2020).
- 167 Lanzolla, G., Marino, M. & Marcocci, C. Selenium in the Treatment of Graves' Hyperthyroidism and Eye Disease. *Front Endocrinol (Lausanne)***11**, 608428, doi:10.3389/fendo.2020.608428 (2020).
- 168 Wiersinga, W. *et al.* Predictive score for the development or progression of Graves' orbitopathy in patients with newly diagnosed Graves' hyperthyroidism. *Eur J Endocrinol***178**, 635-643, doi:10.1530/EJE-18-0039 (2018).

- 169 Le Moli, R. *et al.*Type 2 diabetic patients with Graves' disease have more frequent and severe Graves' orbitopathy. *Nutr Metab Cardiovasc Dis***25**, 452-457, doi:10.1016/j.numecd.2015.01.003 (2015).
- 170 Naselli, A. *et al.*Evidence That Baseline Levels of Low-Density Lipoproteins Cholesterol Affect the Clinical Response of Graves' Ophthalmopathy to Parenteral Corticosteroids. *Front Endocrinol (Lausanne)***11**, 609895, doi:10.3389/fendo.2020.609895 (2020).

Treatment	<i>Mechanism of action</i>	<i>Dose and treatment protocol</i>	<i>Treatment indication</i>
<u>ATX-GD-59</u>	Inducing immune tolerance.	10 i.d. injections over a period of 18 weeks.	Phase 1 clinical trial ¹⁵ .
<u>K1-70</u>	Human monoclonal antibody blocking thyroid stimulating hormone receptor.	Single i.m or i.v injection at different dosage (from 0.2 mg to 150 mg) have been tested.	Phase 1 clinical trial ⁹⁰ .
<u>Small molecule TSH-R antagonists</u>	Direct inhibition of thyroid stimulating hormone receptor activity.	N/A	Mouse model ⁹² ; <i>In vitro</i> ⁹¹ .
<u>Iscalimab</u>	Inhibition of antigen presentation by blocking CD40.	5 intravenous injection (10 mg/kg) over 12 weeks.	Phase 2 clinical trial ¹¹ .
<u>Belimumab</u>	Prevention of B cell maturation, clone expansion and differentiation into plasmacells	N/A	Mouse model ¹⁰¹
<u>Teprotumumab</u>	Human monoclonal antibody blocking growth factor 1- receptor.	8 intravenous infusions over 24 weeks (once every 3 weeks; 10 mg/kg for the first infusion, then 20 mg/kg).	First-line treatment for GO regardless severity and activity in USA. Second-line treatment for moderate-to-severe active GO in Europe, according to availability and affordability ^{57,105} .
<u>Rituximab</u>	Induction of B cells depletion by binding CD20 on B cell surface.	Two infusions of 1000 mg 2 weeks apart or a single dose of 500 mg.	Second-line treatment for moderate-to-severe and active GO of recent onset (<12 months), as long as dysthyroid optic neuropathy is excluded ²¹ .
<u>Tocilizumab</u>	Target blocking of interleukin 6 receptor.	8 mg/kg every four weeks for 12 weeks.	Second-line treatment for moderate-to-severe and active GO ²¹ .
<u>Linsitinib</u>	Target inhibition of the insulin growth factor 1- receptor.	75-150 mg BID for 24 weeks	Mouse model ¹²⁹ In vitro ¹³⁰ A multicenter randomized clinical trial is ongoing to define the potential use (NCT05276063).
<u>Batoclimab</u>	Target inhibition of the neonatal crystallizable fragment receptor (FcRn).	Subcutaneous administration of 680 mg once a week for 12 weeks followed by 340 mg once a week for 12 weeks.	Proof-of-concept randomized trial ¹²⁸ A phase 3, multicenter, randomized, placebo-controlled, clinical trial is ongoing to define the potential use (NCT05517421).
<u>Sirrolimus</u>	Systemic immunosuppression and anti-inflammation, and anti-fibrosis properties through the inhibition of the mTOR pathway.	2 mg as initial dose, followed by 0.5 mg daily for 12 weeks.	Off-label therapy ¹³⁴ .
<u>Atorvastatin</u>	Systemic immunomodulation and anti-inflammation.	20 mg once per day for 24 weeks, plus methylprednisolone 500 mg once a week for 6 weeks followed by 250 mg once a week for an additional six weeks.	Off-label therapy ¹³⁹

Table 1. Novel treatments for Graves' disease and Graves' orbitopathy

i.d., intradermal; i.m, intramuscular; i.v, intravenous; TSH-R, thyroid-stimulating hormone receptor, CD40, Cluster of differentiation 40; GO, Graves' orbitopathy.



