

BRAIN COMMUNICATIONS

Lack of epistatic interaction of *SNCA* with *APOE* in synucleinopathies

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Two recent studies suggested that the *APOE* $\epsilon 4$ haplotype was associated with increased α -synuclein pathology in cell and mouse models. Genetic variants in the *SNCA* region have strong association with Parkinson's disease (PD), dementia with Lewy bodies (DLB) and idiopathic REM sleep behaviour disorder (iRBD), while *APOE* is a genetic risk determinant for only DLB. To determine if genetic-level interactions between *SNCA* and *APOE* exists that can explain the protein-level association, we investigated the genotypic interaction of *APOE* and *SNCA* in cohorts of PD, DLB and iRBD. We analysed genome-wide association study (GWAS) data from 5229 PD patients and 5480 controls, 2610 DLB patients and 1920 controls, and 1055 iRBD patients and 3667 controls. We used logistic regression interaction models across all three cohorts independently between the (i) top GWAS signals of *SNCA* single nucleotide polymorphisms (SNPs) and *APOE* haplotypes and (ii) SNP \times SNP and three-way SNP interaction across the entire coding region plus 200 kb flanking each gene. No significant interactions were found to be associated with any of the synucleinopathies after correction for multiple testing. Our results do not support a role for genetic interactions between *APOE* and *SNCA* across PD, DLB and iRBD. Since the tested genetic variants affect the expression and function of these proteins, it is likely that any interactions between them do not affect the risk of PD, DLB and iRBD.

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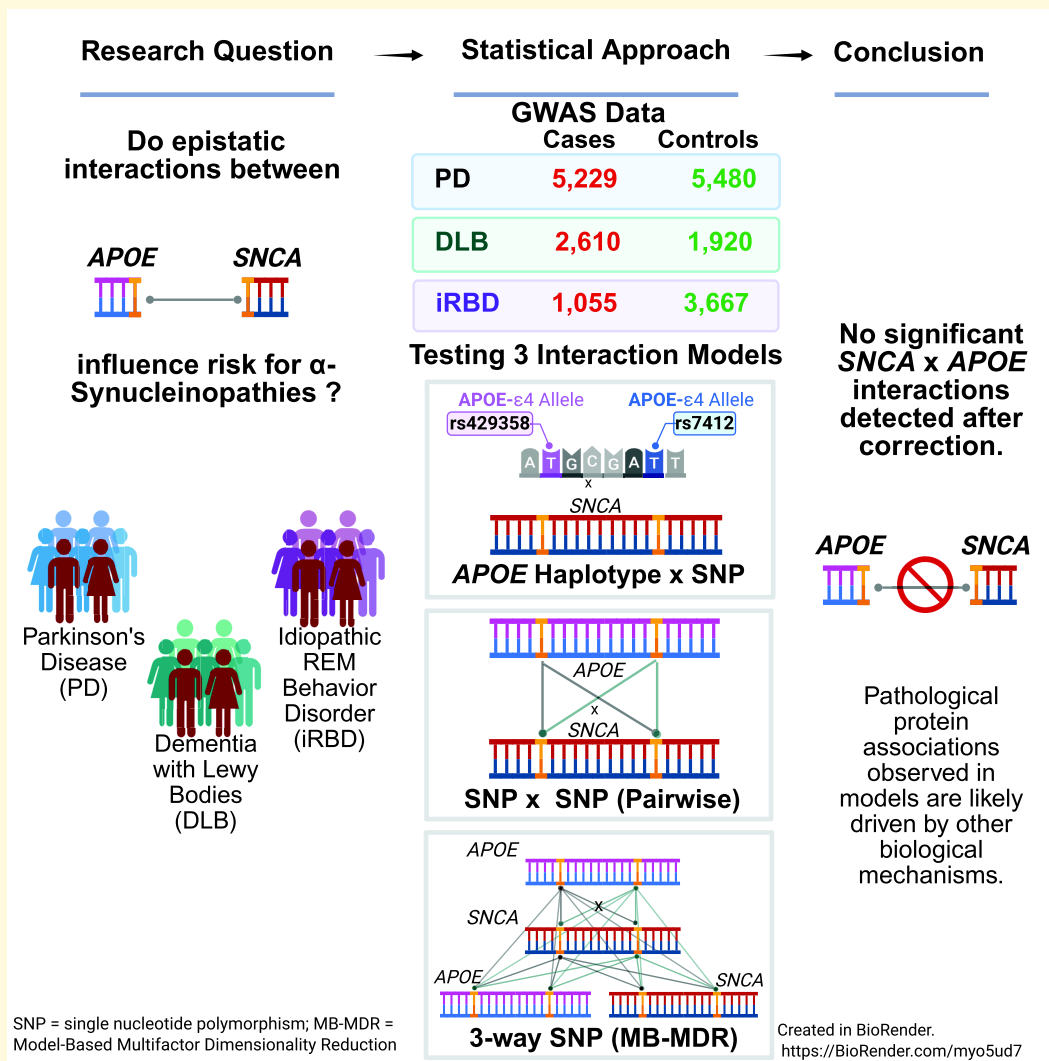
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Graphical Abstract



Introduction

Alpha-synucleinopathy is an umbrella term to describe several neurodegenerative diseases that have a common defining pathological feature, characterized by neuronal or glial inclusions of aggregated alpha-synuclein, known as Lewy bodies, Lewy neurites or glial cytoplasmic inclusions in the brain.¹ Disorders that are collectively referred to as alpha-synucleinopathies include Parkinson's disease (PD), dementia with Lewy bodies (DLB) and multiple system atrophy (MSA). Furthermore, there is an increased presence of alpha-synucleinopathy in the prodromal condition idiopathic/isolated REM sleep behaviour disorder (iRBD), which can convert to either PD, DLB or MSA in more than 80% of cases.²

Alpha-synuclein is encoded by the *SNCA* gene, and genetic variants in the *SNCA* locus are associated with PD, DLB and iRBD risk in genome-wide association studies (GWASs).³⁻⁸

Specifically, some variants of *SNCA* are strongly associated with PD (rs356182 and rs2870004), while others are associated with DLB (rs7681440 and rs7680557)^{7,9} and iRBD (rs2870004).¹⁰ The top *SNCA* association in PD is independent and different than the top associations in DLB⁷ and iRBD,⁸ raising the hypothesis that there could be differential effects of *SNCA* variants on the expression of alpha-synuclein in different brain regions.⁸

Overlapping neuropathologic features associated with Alzheimer's disease (AD) are seen in the brains of many patients with PD¹¹ and dementia including amyloid plaques composed of amyloid-beta ($A\beta$) plaques and neurofibrillary tangles containing the tau protein and may contribute to clinical features of disease.^{12,13}

Coding variants in apolipoprotein E (*APOE*) produce 3 common alleles, ϵ 2, ϵ 3 and ϵ 4. The ϵ 4 allele of *APOE* alters lipid metabolism regulation and cholesterol transport and

known to be a major genetic risk determinant for sporadic, late-onset Alzheimer's disease¹⁴ and Lewy body dementia.^{7,9} Dose effects by allele have demonstrated a 3.7-fold risk of developing AD while homozygosity increases the risk by up to 12-fold.⁷ From numerous GWASs, *APOE* does not alter the risk for PD, yet the $\epsilon 4$ allele has been described as a potential risk factor for cognitive decline and development of dementia in PD patients.^{15,16}

Two studies in mice demonstrated that the *APOE** $\epsilon 4$ genotype was associated with increased alpha-synuclein pathology, independent of the amyloid β deposition.^{17,18} These two studies emphasize a potential molecular mechanism of *APOE** $\epsilon 4$ on α -synuclein protein aggregation. However, beyond A53T, they have not evaluated disease specific variants of *SNCA* that have functional molecular consequences (e.g. E46K).¹⁹ Analysis of these variants could be insightful in understanding molecular association between *SNCA* variants and *APOE** $\epsilon 4$. Furthermore, while the *SNCA* locus is associated with all synucleinopathies, *APOE* is a genetic risk factor for DLB only. *SNCA* and *APOE* variants may affect the expression/function of the proteins encoded by them.²⁰ Therefore, if a true interaction exists at the protein level as suggested by the studies mentioned above, then there plausibly should be evidence of some association at the genetic level.

Genetic interactions refer to a combination of two or more genetic variants whose phenotypic contribution is amplified by their co-occurrence.²¹ To determine if genetic-level interactions between *SNCA* and *APOE* exist that can explain the protein-level association as described, we investigated the genotypic interaction of *APOE* and *SNCA* in three disease cohorts of PD, DLB and iRBD patients and controls, with a total of 8855 patients and 11 067 controls.

Materials and methods

Patient population

For PD, we used the International Parkinson's Disease Genomics Consortium (IPDGC) dataset that contained 10 709 subjects with 5229 cases and 5480 controls. For DLB, we used the most recent DLB GWAS dataset, which included 4530 subjects with 2610 cases and 1920 controls. The iRBD cohort was composed of 4742 individuals, including 1055 cases and 3667 controls with 1968 controls from the NeuroGenetics Research Consortium (NGRC) (dbGAP: phs000196.v2.p1) and 790 controls from National Institute of Neurological Disorders and Stroke (NINDS) (dbGAP: phs000089) added from external studies of Parkinson's patients in addition to the controls collected for the iRBD cohort. PD was diagnosed using UK Brain Bank criteria or Movement Disorders Society (MDS) criteria.⁴ DLB patients were diagnosed with pathologically definite or clinically probable disease according to consensus criteria.²² iRBD was diagnosed according to the International Classification of Sleep Disorders (2nd or 3rd Edition).^{23,24} Informed consent and

ethics approval was obtained from the appropriate institutional review boards at participating institutions as described in the original studies. The STREGA reporting guidelines were used for this study.¹⁷

Genetic analysis

We generated genotype calls of two *APOE* SNPs, rs429358 and rs7412, to determine the *APOE* haplotype status of each sample. The combination of genotypes for rs429358 (C/T) and rs7412 (C/T) defines the three *APOE* haplotypes: epsilon 2 ($\epsilon 2$), epsilon 3 ($\epsilon 3$) and epsilon 4 ($\epsilon 4$). These three haplotypes can produce six genotypes, $\epsilon 2/\epsilon 2$, $\epsilon 2/\epsilon 3$, $\epsilon 2/\epsilon 4$, $\epsilon 3/\epsilon 3$, $\epsilon 3/\epsilon 4$ and $\epsilon 4/\epsilon 4$.

Cohorts were collected, quality controlled, genotyped, and filtered on individual and variant level as previously described for PD,⁴ iRBD⁸ and DLB.⁷ *APOE* variants were analysed for the 200 kb region flanking both sides on chromosome 19, 44 705 791–45 109 393, and for *SNCA* on chromosome 4, 89 500 345–90 038 324. The DLB genotypic data was converted from GrCh38 to GrCh37 using Liftover.¹⁸ Because external controls were added to the iRBD cohort; the cases and control genotypes were filtered for minor allele frequency (MAF) > 0.01 to reduce imputation errors and imputed using Michigan Imputation Server and the Haplotype Reference Consortium¹⁹ r1.1 2016 reference panel (GRCh37/hg19). Only imputed genotypes with an $R^2 > 0.30$ were kept for analysis. Additionally, prior to analysis, further quality control for each cohort included removing duplicate samples, missing data including covariates; SNPs were filtered based on variant missingness (<0.05), genomic relatedness (>0.125), disparate missingness between cases and controls ($P > 1E-04$), missingness by haplotype ($P > 1E-04$), deviation from Hardy–Weinberg equilibrium ($P > 1E-04$), minor allele frequency (MAF) > 0.01 and LD pruned with r^2 at >0.5 with a 50 kb window using plink 1.9.²⁰

Statistical analysis

Descriptive measures of mean, standard deviations, frequencies and percentages were used to summarize the data. SNP and haplotype interaction were analysed using logistic regression controlling for age, sex and ancestry using the first five principal components. Epistasis model was defined as

$$Y \sim \beta_0 + \beta_1 A + \beta_2 B + \beta_3 AB + \epsilon$$

where β_0 represents the intercept, β_n represents the coefficients for each SNP and A and B represent allele dosage of each SNP and AB represents the interaction. The test for interaction was based on the coefficient β_3 and P -value < 0.05 as significant when testing interaction between top SNPs and Bonferroni threshold applied for SNP×SNP interactions. Detecting gene–gene interactions in complex diseases can be accomplished using a variety of epistasis-focused tools or packages in existing software packages. Model-based multifactor dimensionality

Table 1 Age and sex of each cohort in cases and controls

Demographic			Total	PD	DLB	iRBD
Sex	Cases	Male (%)	5845 (65.72)	3366 (64.37)	1642 (62.91)	837 (79.34)
		Female (%)	3010 (33.84)	1863 (35.63)	947 (36.28)	200 (18.96)
	Controls	Male (%)	7256 (65.56)	3055 (55.75)	972 (50.63)	1564 (42.65)
		Female (%)	3811 (34.44)	2425 (44.25)	948 (49.38)	2103 (57.34)
Age	Cases		65.7 (13.6)	61.2 (12.6)	74.7 (10.5)	-
	Controls		67.9 (16.2)	64.3 (14.8)	72.7 (16.8)	-

Values for age represent mean and standard deviation in parenthesis. Age information for iRBD was missing. Abbreviations: PD, Parkinson's disease; DLB, dementia with Lewy bodies; iRBD, idiopathic rapid eye movement (REM) sleep behaviour disorder.

reduction (MB-MDR) was implemented as a final screen strategy to detect any significant SNP×SNP and SNP×SNP×SNP interactions using the open-source MB-MDR v 4.4.1 software, as MB-MDR merges multi-locus genotypes exhibiting some significant evidence of High or Low risk, based on association testing into a new lower-order dimension. A new association test is subsequently performed per marker pair/triplet, by adopting a permutation-based strategy that corrects for multiple testing (over all marker pairs/triplets) and adequately controls family-wise error rate at $\alpha = 5\%$.²¹

Results

To explore possible interactions between *APOE* and *SNCA* in cases and controls, we first focused primarily on the top *SNCA* SNPs associated with PD, DLB and RDB (Table 1) and the *APOE* haplotypes (Table 2). We performed a SNP-haplotype logistic regression interaction controlling for age, sex and ancestry using the first five principal components. No significant interactions were found to be statistically significant (Table 3).

To further explore all potential genetic interactions between *APOE* and *SNCA*, we expanded the range to include all SNPs in *APOE* and *SNCA* plus 200 kb outside the region ends on both genes across all cohorts. We pruned SNPs that were in LD with an r^2 of 0.5 with 316 SNPs remaining in PD, 421 SNPs remaining in DLB and 198 SNPs remaining in iRBD. We applied PLINKs regression-based approach to model and test SNP×SNP interactions. After correction for multiple comparisons with a Bonferroni correction, we did not identify any interactions associated with any of the alpha-synucleinopathies (Supplementary Tables 1–3).

Lastly, a MB-MDR method was implemented as a final screen strategy to detect any significant SNP×SNP and SNP×SNP×SNP interactions using the open-source MB-MDR software.²² Here too, no significant interactions were found to be associated with PD, DLB or iRBD after correction for multiple testing using permutation testing across all three cohorts (Supplementary Tables 4–6).

Discussion

Our results do not support a role for genetic interactions between *APOE* and *SNCA* across PD, DLB and iRBD. The

Table 2 APOE haplotype frequencies of each cohort in cases and controls

Group	Haplotype	Controls	Cases	Total
DLB	e2e2	4 (0.21)	5 (0.19)	9
	e2e3	171 (8.91)	189 (4.17)	360
	e2e4	17 (0.89)	83 (3.18)	100
	e3e3	1273 (66.30)	1203 (46.09)	2476
	e3e4	419 (21.82)	916 (35.10)	1335
	e4e4	36 (1.88)	214 (8.20)	250
	Total		1920 (42.38)	2610 (57.62)
iRBD	e2e2	22 (0.61)	5 (0.48)	27
	e2e3	444 (12.27)	109 (10.4)	553
	e2e4	82 (2.27)	15 (1.43)	97
	e3e3	2241 (61.92)	671 (64.03)	2912
	e3e4	752 (20.78)	236 (22.52)	988
	e4e4	78 (2.16)	12 (1.15)	90
	Total		3619 (77.54)	1048 (22.46)
PD	e2e2	41 (0.75)	25 (0.48)	66
	e2e3	725 (13.23)	656 (12.55)	1381
	e2e4	106 (1.93)	75 (1.43)	181
	e3e3	3599 (65.68)	3498 (66.90)	7097
	e3e4	954 (17.41)	932 (17.82)	1886
	e4e4	55 (1.00)	43 (0.82)	98
	Total		5480 (51.17)	5229 (48.83)

Values represent frequency and percentage. Bold values represent totals. Abbreviations: PD, Parkinson's disease; DLB, dementia with Lewy bodies; iRBD, idiopathic rapid eye movement (REM) sleep behaviour disorder.

genetic variants that were tested influence the expression and function of these proteins, but it is unlikely that any interactions between them affect the risk of developing synucleinopathies. Although functional epistasis, in the form of biomolecular interaction, can determine biological pathways of disease progression, it may not always be detected through mathematical or statistical genetic interaction analysis. However, if longitudinal disease progression data is part of the analysis, then mixed effects models could be used to analyse within and between subject variability on the interplay between haplotype, *SNCA* risk variants and disease progression. The genetic interactions identified in our study, which represent three synucleinopathies, do not modify the risk of developing PD, DLB or iRBD in our clinical cohorts. This suggests that the pathological interactions between *APOE* and *SNCA* observed in model organisms or human synucleinopathies may not be driven by genetic epistasis influencing disease susceptibility. Therefore, these pathological associations may be driven by other mechanisms such as regulatory

Table 3 Top hits of SNCA and APOE haplotype interaction regression results

Disease	Gene	Chr	Location	Ref Allele	Alt Allele	rs	Interaction	Estimate	Standard Error	z-score	Pr(> z)
iRBD	SNCA	4	90471245	T	A	rs2870004	rs2870004×APOE Haplotype	−0.06359	0.06976	−0.912	0.362
		4	90626111	G	A	rs356182	rs356182×APOE Haplotype	0.03883	0.0629	0.617	0.537021
		4	90756550	C	G	rs7681440	rs7681440−APOE Haplotype	0.04838	0.05954	0.812	0.416257
PD	SNCA	4	90471245	T	A	rs2870004	rs2870004×APOE Haplotype	0.039526	0.038801	1.019	0.30835
		4	90626111	G	A	rs356182	rs356182×APOE Haplotype	0.027522	0.031979	0.861	0.389447
		4	90756550	C	G	rs7681440	rs7681440−APOE Haplotype	−0.016695	0.030556	−0.546	0.584807
DLB	SNCA	4	89550094	T	A	rs2870004	rs2870004×APOE Haplotype	−0.139925	0.118107	−1.185	0.2361
		4	89704960	G	A	rs356182	rs356182×APOE Haplotype	0.152286	0.110879	1.373	0.17
		4	89835399	C	G	rs7681440	rs7681440−APOE Haplotype	0.070889	0.100602	0.705	0.481

Abbreviations: PD, Parkinson's disease; DLB, dementia with Lewy bodies; iRBD, idiopathic rapid eye movement (REM) sleep behaviour disorder.

factor expression and/or post-translational modifications, protein–protein interactions or environmental factors.

Previous studies have shown that APOE $\epsilon 4$ is linked to DLB in both AD and non-AD cases.^{23,24} However, some studies have also shown that APOE $\epsilon 4$ is only associated with DLB when there is a significant amount of co-existing Alzheimer's pathology.^{25–27} This finding contradicts the idea that APOE $\epsilon 4$ independently drives α -synuclein pathology.

This study has several limitations. The lack of pathological confirmation of Lewy bodies and AD pathology in the cohorts does not allow for an analysis based on co-pathology. Such analysis would have been able to detect interactions that exist only in subpopulation of patients, for example, those who have both alpha-synuclein and amyloid pathology. Another limitation is that the study scope was limited to APOE and SNCA. Recently, a stratified GWAS of DLB uncovered an association only between GBA rs2230288 and pathologically confirmed DLB without AD pathology, but not in mixed pathological cases.²⁷ This same SNP has been identified as a significant risk variant in the most recent GWAS of DLB.⁷ It is possible that large-scale GWAS would uncover associations that encompass a broad spectrum of disease (e.g. DLB with no AD pathology, DLB with mixed PD and AD pathology, etc.), even though these associations may be driven by subsets of these genetic loci (e.g. GBA). Furthermore, it is plausible that different disease subtypes consist of distinct genetic combinations and interactions that have not yet been identified at a population level due to the limited size of current sample cohorts. Further studies should include more samples and implement alternative statistical or interrogative methods to leverage the current data to its fullest potential despite its small sample size.

Supplementary material

Supplementary material is available at *Brain Communications* online.

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Competing interests

Prabhjyot Saini—Nothing to declare; Eric Yu—Nothing to declare; Mehrdad A. Estiar—Nothing to declare; Lynne Krohn—Nothing to declare; Kheireddin Mufti—Nothing to declare; Uladzislau Rudakou—Nothing to declare; Jennifer A. Ruskey—Nothing to declare; Farnaz Asayesh—Nothing to declare; Sandra B. Laurent—Nothing to declare; Dan Spiegelman—Nothing to declare; Jean-François Trempe—Nothing to declare; Timothy G. Quinnell—Nothing to declare; Nicholas Oscroft—Nothing to declare; Isabelle Arnulf—I.A. was previously consultant for Idorsia Pharma and UCB Pharma. Jacques Y. Montplaisir—Nothing to declare; Jean-François Gagnon—Jean-François Gagnon; Alex Desautels—Alex Desautels received operating grants from CHIR, AASM and research grants from Eisai, Takeda and Canopy Growth; honoraria from serving on the scientific advisory board of Eisai, Paladin Labs and UCB, as well as honoraria from speaking engagements from Eisai, Jazz Pharma and Paladin Labs. None of the financial disclosures is relevant to the submitted work. Yves Dauvilliers—has served as a consultant or on advisory

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Data availability

The data that support the findings of this study are available from dbGaP. Parkinson's patients and controls (phs000918.v1.p1). DLB patients and controls are available from dbGaP (phs001963.v1.p1). iRBD patients and controls are available upon reasonable request from the corresponding author. Additional controls were also obtained from dbGaP; NGRC (phs000196.v2.p1) and NINDS (phs000089). Code used for analysis can be found on GitHub (https://github.com/gan-orlab/APOE_SNCA).

References

1. Calabresi P, Mechelli A, Natale G, Volpicelli-Daley L, Di Lazzaro G, Ghiglieri V. Alpha-synuclein in Parkinson's disease and other synucleinopathies: From overt neurodegeneration back to early synaptic dysfunction. *Cell Death Dis.* 2023;14(3):176.
2. Postuma RB, Iranzo A, Hu M, *et al.* Risk and predictors of dementia and parkinsonism in idiopathic REM sleep behaviour disorder: A multicentre study. *Brain.* 2019;142(3):744-759.
3. Foo JN, Chew EGY, Chung SJ, *et al.* Identification of risk loci for Parkinson disease in Asians and comparison of risk between Asians and Europeans: A genome-wide association study. *JAMA Neurol.* 2020;77(6):746.
4. Nalls MA, Blauwendraat C, Vallerga CL, *et al.* Identification of novel risk loci, causal insights, and heritable risk for Parkinson's disease: A meta-analysis of genome-wide association studies. *Lancet Neurol.* 2019;18(12):1091-1102.
5. Nalls MA, Pankratz N, Lill CM, *et al.* Large-scale meta-analysis of genome-wide association data identifies six new risk loci for Parkinson's disease. *Nat Genet.* 2014;46(9):989-993.
6. Chang D, Nalls MA, Hallgrímsdóttir IB, *et al.* A meta-analysis of genome-wide association studies identifies 17 new Parkinson's disease risk loci. *Nat Genet.* 2017;49(10):1511-1516.
7. Chia R, Sabir MS, Bandres-Ciga S, *et al.* Genome sequencing analysis identifies new loci associated with Lewy body dementia and provides insights into its genetic architecture. *Nat Genet.* 2021; 53(3):294-303.
8. Krohn L, Heilbron K, Blauwendraat C, *et al.* Genome-wide association study of REM sleep behavior disorder identifies polygenic risk and brain expression effects. *Nat Commun.* 2022; 13(1):7496.
9. Guerreiro R, Ross OA, Kun-Rodrigues C, *et al.* Investigating the genetic architecture of dementia with Lewy bodies: A two-stage genome-wide association study. *Lancet Neurol.* 2018;17(1):64-74.
10. Krohn L, Wu RYJ, Heilbron K, *et al.* Fine-mapping of SNCA in rapid eye movement sleep behavior disorder and overt synucleinopathies. *Ann Neurol.* 2020;87(4):584-598.
11. Dugger BN, Adler CH, Shill HA, *et al.* Concomitant pathologies among a spectrum of parkinsonian disorders. *Parkinsonism Relat Disord.* 2014;20(5):525-529.

12. Irwin DJ, Lee VM, Trojanowski JQ. Parkinson's disease dementia: Convergence of α -synuclein, tau and amyloid- β pathologies. *Nat Rev Neurosci*. 2013;14(9):626-636.
13. Irwin DJ, White MT, Toledo JB, et al. Neuropathologic substrates of Parkinson disease dementia. *Ann Neurol*. 2012;72(4):587-598.
14. Serrano-Pozo A, Das S, Hyman BT. APOE and Alzheimer's disease: Advances in genetics, pathophysiology, and therapeutic approaches. *Lancet Neurol*. 2021;20(1):68-80.
15. Morley JF, Xie SX, Hurtig HI, et al. Genetic influences on cognitive decline in Parkinson's disease. *Mov Disord*. 2012;27(4):512-518.
16. Pankratz N, Byder L, Halter C, et al. Presence of an APOE4 allele results in significantly earlier onset of Parkinson's disease and a higher risk with dementia. *Mov Disord*. 2006;21(1):45-49.
17. Little J, Higgins JP, Ioannidis JP, et al. Strengthening the REporting of Genetic Association Studies (STREGA)—an extension of the STROBE statement. *Genet Epidemiol*. 2009;33(7):581-598.
18. Kuhn RM, Haussler D, Kent WJ. The UCSC genome browser and associated tools. *Brief Bioinform*. 2013;14(2):144-161.
19. McCarthy S, Das S, Kretzschmar W, et al. A reference panel of 64,976 haplotypes for genotype imputation. *Nat Genet*. 2016;48(10):1279-1283.
20. Purcell S, Neale B, Todd-Brown K, et al. PLINK: A tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet*. 2007;81(3):559-575.
21. Mahachie John JM, Van Lishout F, Van Steen K. Model-based multifactor dimensionality reduction to detect epistasis for quantitative traits in the presence of error-free and noisy data. *Eur J Hum Genet*. 2011;19(6):696-703.
22. Cattaert T, Calle ML, Dudek SM, et al. Model-based multifactor dimensionality reduction for detecting epistasis in case-control data in the presence of noise. *Ann Hum Genet*. 2011;75(1):78-89.
23. Tsuang D, Leverenz JB, Lopez OL, et al. APOE ϵ 4 increases risk for dementia in pure synucleinopathies. *JAMA Neurol*. 2013;70(2):223-228.
24. Dickson DW, Heckman MG, Murray ME, et al. APOE ϵ 4 is associated with severity of Lewy body pathology independent of Alzheimer pathology. *Neurology*. 2018;91(12):e1182-e1195.
25. Prokopenko I, Miyakawa G, Zheng B, et al. Alzheimer's disease pathology explains association between dementia with Lewy bodies and APOE- ϵ 4/TOMM40 long poly-T repeat allele variants. *Alzheimers Dement (N Y)*. 2019;5:814-824.
26. Schaffert J, LoBue C, White CL 3rd, et al. Risk factors for earlier dementia onset in autopsy-confirmed Alzheimer's disease, mixed Alzheimer's with Lewy bodies, and pure Lewy body disease. *Alzheimers Dement (N Y)*. 2020;16(3):524-530.
27. Kaivola K, Shah Z, Chia R, et al. Genetic evaluation of dementia with Lewy bodies implicates distinct disease subgroups. *Brain*. 2021;145(5):1757-1762.