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28 131 \pm 1 mgNO₃ -N L⁻¹d⁻¹), although a progressive decrease in desalination efficiency (from 77 ± 13 to $12\pm2\%$) was observed. Nitrate concentration and salinity close to threshold limits indicated by the World Health Organization for drinking water, as well as significant 31 chlorine production were achieved with an HRT of 4.9 ± 0.4 h. At such HRT, specific energy 32 consumption was low $(6.8 \cdot 10^{-2} \pm 0.3 \cdot 10^{-2} \text{ kWh } \text{g}^{-1} \text{NO}_3\text{-N}_{\text{removed}})$, considering that the supplied energy supports three processes simultaneously. A logarithmic equation correlated well with nitrate removal rates at the applied HRTs and may be used to predict BES behaviour with different HRTs. The bacterial community of the bio-cathode under galvanostatic mode was dominated by a few populations, including the genera *Rhizobium, Bosea, Fontibacter* and *Gordonia*. The results provide useful information for the scale-up of BES treating multi-contaminated groundwater.

 Keywords: circular economy; denitrification; microbial electrochemical technology; saline groundwater; value-added products; water recovery.

1. INTRODUCTION

 Groundwater is a critical freshwater reservoir, fundamental for global water and food security. Since the spread of contaminants in groundwater can limit its use as drinking water, actions must be taken to ensure its safe supply (Janža, 2022). Bioelectrochemical systems (BES) have emerged as sustainable alternatives to conventional bioremediation technologies for treating contaminated groundwater. Such systems are based on the ability of electroactive microorganisms to perform oxidation and reduction reactions by exchanging electrons with an electrode (Pous et al., 2018; Wang et al., 2020). Therefore, they are particularly suitable for groundwater treatment, as they promote bioremediation without needing chemicals as electron acceptors/donors.

 Since groundwater may be simultaneously exposed to different sources of pollution, one of the most intriguing challenges that researchers are currently facing is the application of BES to the bioremediation of multi-contaminated groundwater (Ceballos-Escalera et al., 2021; Wang et al., 2021; Cruz Viggi et al., 2022; Tucci et al., 2021, Puggioni et al., 2021). Among contaminants, nitrate is often found in groundwater due to inefficient farming practices and careless management of livestock activities (Kwon et al., 2021; Serio et al., 2018), thus hindering the exploitation of such important water reservoirs since it is related to severe health risks (Carrey et al., 2021; Ward et al., 2018). BES proved to be a promising solution for the remediation of nitrate-contaminated groundwater (Li et al., 2019; Pous et al., 2018), and they have demonstrated the possibility of achieving complete nitrate conversion into dinitrogen gas via autotrophic denitrification at the bio-cathode, with no or negligible production of intermediates such as nitrite and nitrous oxide (Ceballos-Escalera et al. 2021; Puig et al. 2011; Desloover et al., 2011).

 Besides nitrates, the occurrence of salinity in groundwater is often related to the over- exploitation of water reservoirs in coastal areas, which causes the alteration of the hydrodynamic balance between seawater and freshwater. The consequent seawater intrusion and salinisation of the aquifer limit the potential use of groundwater for human consumption (Liu et al., 2020). BES-based desalination through Microbial Desalination Cells (MDC) has been successfully applied, especially for seawater desalination. In such systems, the electric potential gradient created by the exoelectrogenic bacteria in the presence of organic matter allows water desalination by driving ion transport through a series of ion-exchange membranes (Ramírez-Moreno et al., 2019; Sevda et al., 2015; Kim et al., 2013).

 Though the combination of high nitrate concentrations and salinity levels severely affects groundwater quality in many countries worldwide (Troudi et al., 2020; Alfarrah et al., 2018; Gounari et al., 2014), only a few studies concerning their simultaneous removal using BES have been reported in the literature so far. Zhang et al. (2013) successfully tested a 2-compartment submerged microbial desalination-denitrification cell (SMDDC) for the treatment of synthetic groundwater affected by high salinity and nitrate concentration 80 (0.9-2.2 mS cm⁻¹ and 20 mgNO₃ -N L⁻¹, respectively). A readily degradable organic substrate 81 (i.e., sodium acetate, $800 \text{ mg } L^{-1}$) was spiked in the anode to sustain the process. The system achieved salinity and nitrate removal efficiencies of up to 60% and 99%, respectively, at the highest ionic strength tested. Recently, our research group designed and tested a proof-of-concept configuration based on a 3-compartment BES for the simultaneous 85 removal of nitrate $(30 \text{ mgNO}_3\text{-N L}^{-1})$ and salinity $(3.3\pm0.3 \text{ mS cm}^{-1})$ from synthetic saline groundwater, without any addition of organic substrates, and with the concomitant production of a value-added chemical (i.e., free chlorine, a disinfectant commonly used in water and wastewater treatment plants) (Puggioni et al., 2021). In this reactor, the electroactive biofilm attached to the bio-cathode carried out full autotrophic denitrification. At the same time, desalination took place in the central compartment thanks to the electrochemically driven migration of ions across two ion-exchange membranes. Part of the accumulated chloride was converted into chlorine in the anode compartment. Different operating strategies were tested, and the galvanostatic operation (applied current, 10 mA) 94 with pH control $(< 9$) in the bio-cathode compartment resulted in high nitrogen and salinity 95 removal efficiencies $(69\pm2\%$ and $63\pm5\%$, respectively) and significant recovery of free chlorine. Standard quality requirements for drinking water in terms of nitrate concentration 97 (91/767/EU, 11.3 mgNO₃ -N L⁻¹) and electrical conductivity (98/83/CE, 2.5 mS cm⁻¹) were successfully met at high hydraulic retention time (HRT) with no apparent limitations to biological nor physical-chemical processes involved. The proposed 3-compartment configuration still showed ample room for improvement in terms of nitrate removal rates and specific energy consumption.

 Among the operating parameters that can optimise the process, the HRT is known to influence denitrifying BES performance, which can be severely limited by the hydrodynamics and the corresponding distribution of nitrate/substrates in the system (Vilà- Rovira et al., 2015). Pous et al. (2015) reported that insufficient mixing could generate different gradients of nitrate and pH along the reactor, producing different environments and, thus, heterogeneity in microbial growth and activity. Ceballos-Escalera et al. (2021) 108 observed an increase in nitrate removal rates (from 166 ± 22 to 247 ± 22 mgNO₃-N L⁻¹d⁻¹) as 109 the HRT was progressively reduced from 7.5 ± 0.2 to 3.6 ± 0.2 h in a tubular BES treating 110 synthetic nitrate- and arsenic-contaminated groundwater $(28\pm6 \text{ mgNO}_3\text{-N L}^{-1}$ and 5 111 mgAs(III) L^{-1} , respectively). Pous et al. (2017) observed the enhancement of nitrate removal 112 rates (from 73 ± 5 to 849 ± 23 mgNO₃-N L⁻¹d⁻¹) with decreasing HRTs (from 10.89 \pm 0 to 0.46±0.01 h) in a tubular BES fed with synthetic nitrate-contaminated groundwater (33 114 mgNO₃ -N L^{-1}). Such behaviour was mainly attributed to the increase in bacterial activity rather than bacterial growth due to the reinforced water flowrate itself, and not due to the increase in nitrate availability (Pous et al., 2017). However, short HRT has been proved to have a detrimental effect on desalination technologies such as microbial desalination cells since it negatively influences biomass activities at the anode (Imoro et al., 2021) and current generation (Jingyu et al., 2017).

 Within this framework, the influence of the HRT on denitrification and desalination performance of the 3-compartment cell configuration developed by Puggioni et al. (2021) was investigated in this study. Since the overall effect of decreasing HRT is difficult to be predicted when biotic (e.g., nitrate removal) and abiotic (i.e., desalination, chloride removal

 and chlorine production) processes co-exist in the same reactor, the outcomes of this study will provide helpful information for maximising system's performance and finding its operational limits, with the perspective of fostering the scale-up of the system by reducing the capital and operating costs linked to the size of the reactors and the energy consumption, respectively.

2. MATERIALS AND METHODS

2.1 Reactor set-up

 Two identical 3-compartment cells made of polycarbonate were used (Puggioni et al., 2021). 132 Each cell consisted of a bio-cathode compartment $(8x8x2 \text{ cm}^3, \text{net volume } 110\pm9 \text{ mL})$, an 133 anode compartment $(8x8x2 \text{ cm}^3, \text{ net volume } 130\pm7 \text{ mL})$, and a thin central desalination 134 compartment ($8x8x0.5$ cm³, net volume 31 ± 2 mL). Such a peculiar geometric configuration was designed to minimise the distance between the electrodes and the membranes and consequently decrease the internal resistance of the system. The bio-cathode compartment 137 contained the graphite felt electrode $(64 \text{ cm}^2, \text{degree of purity } 99.9\% , \text{AlfaAesar}, \text{Germany}),$ and it was physically separated from the central compartment by a cation exchange membrane (CEM 7000, Membrane International Inc., USA). The anode compartment, 140 containing a titanium mesh electrode coated with mixed metals oxide (Ti-MMO, 15 cm^2 , NMT-Electrodes, South Africa), was physically separated from the central compartment by an anion exchange membrane (AEM 7001, Membranes International Inc., USA). A reference electrode (Ag/AgCl, +0.197 V vs SHE, mod. MF2052, BioAnalytical Systems, USA) was placed in the bio-cathode compartment. Cathode, anode, and reference electrodes were connected to a multichannel potentiostat (Ivium technologies, IviumNstat, NL). The 146 system was thermostatically controlled at 25 ± 1 °C.

2.2 Groundwater characteristics

 A synthetic medium mimicking nitrate concentration and salinity of groundwater from the nitrate vulnerable zone of Arborea (Sardinia, Italy) was fed to the bio-cathode compartment. 150 This medium contained 216.6 mg L⁻¹ KNO₃ (corresponding to 30.0 mgNO₃ -N L⁻¹); 10 mg L^{-1} NH₄Cl (corresponding to 2.6 mgNH₄⁺-N L⁻¹), 4.64 mg L⁻¹ KH₂PO₄; 11.52 mg L⁻¹ 152 K₂HPO₄; 350 mg L⁻¹ NaHCO₃; 2000 mg L⁻¹ NaCl and 100 μ L L⁻¹ of trace elements solution 153 (Patil et al., 2010). The resulting electrical conductivity and pH were 3.06 ± 0.5 mS cm⁻¹ and 8.2±0.3, respectively. The medium was prepared using distilled water and pre-flushed with 155 N_2 gas for 15 minutes to avoid any presence of oxygen.

2.3 Experimental procedure

 The cells were already running from previous research, and the initial inoculum consisted of the supernatant of activated sludge liquor drawn from the municipal wastewater treatment plant of Cagliari (Italy) and the effluent from a parent electro-denitrifying system in a 60:40 (v:v) mixture (Puggioni et al., 2021). The bio-cathode compartment was continuously fed with groundwater, and the effluent was sent into the central compartment to achieve desalination. The anode compartment was filled with tap water (Pous et al., 2015) to minimise the use of chemicals and operated in batch mode with recirculation. Tap water was periodically replaced (about every 10 days) to avoid excessive chlorine accumulation. According to the best operating strategy defined in Puggioni et al. (2021), the potentiostat 166 was set in galvanostatic mode at a current of 10 mA (0.16 mA cm^2) _{membrane}), and pH control (< 9) was implemented to avoid excessive pH increase in the bio-cathode compartment, by dosing HCl (1 M) in the bio-cathode recirculation line. The probe for continuous pH measurement (Mettler Toledo, mod. InPro 3253i/SG/225, USA) was connected to a transmitter (Mettler Toledo, mod. M300, USA), which recorded data every 10 minutes.

 Since the enhancement of electro-bioremediation systems must be linked to the treatment capacity, the HRT was used as the operational parameter, as summarised in Table 1. Each HRT was maintained for about one month with constant nitrate influent concentration $(29.3 \pm 3.5 \text{ mgNO}_3 - \text{N L}^{-1}).$

¹ alculated considering the sum of volumes of the bio-cathode and central compartments.

178 $*$ $*$ calculated considering the volume of the central compartment.

 In particular, Test 7 was implemented as a repetition of Test 5 to restore process performance after observing the worsening of denitrification activity in Test 6. Hence, the results achieved during Test 7 were not included in graphical representations nor in the calculations to make the results easier to interpret.

2.4 Analytical methods

 Samples were periodically drawn from influent (once per week), effluent (three times per week), bio-cathode and anode compartments (three times per week) in order to evaluate overall cell performances. The same samples from the duplicate cell were taken once a week to confirm the process progress of the main cell. Liquid samples were analysed for 188 quantification of anions, i.e., chloride (Cl⁻), nitrite (NO₂⁻-N), nitrate (NO₃⁻-N), phosphate 189 ($PO₄³$), and sulphate ($SO₄²$), using an ion chromatograph (ICS-90, Dionex-Thermofisher,

 USA) equipped with an AS14A Ion-PAC 5 μm column. Samples were filtered (acetate membrane filter, 0.45 μm porosity) and properly diluted with grade II water. The 192 concentrations of the main cations, i.e., potassium (K^+) and sodium (Na^+) , were determined using an ICP/OES (Varian 710-ES, Agilent Technologies, USA): samples were filtered (acetate membrane filter, 0.45 μm porosity), acidified (nitric acid, 1% v:v) and diluted with grade I water.

 Electrical conductivity and pH were measured using a benchtop meter (HI5522, Hanna Instruments, Italy).

 The concentration of free chlorine was analysed using spectrophotometric techniques (DR1900, Hach Lange, Germany) and the DPD (N,N-diethyl-p-phenylenediamine) free chlorine method (DPD free chlorine reagent powder pillows Cat. 2105569, Hach Lange, Germany).

 Nitrous oxide (N2O) was measured using an N2O liquid-phase microsensor (Unisense, Den- mark) located in the effluent line of the reactors, thanks to a dedicated glass measuring cell. The resulting bio-cathode potentials were recorded every five minutes through the potentiostat (Ivium technologies, IviumNstat, NL). Cell potential was periodically checked using a multimeter (K2M, mod. KDM-600C, Italy).

2.5 Calculations

 Nitrate Removal Efficiency (N-RE) and Nitrate Removal Rate (N-RR) were calculated according to equations 1 and 2, respectively:

210
$$
N - RE\left[\%\right] = \frac{c_{NO_3^- - N(inf)} - c_{NO_3^- - N(eff)}}{c_{NO_3^- - N(inf)}} \times 100
$$
 (1)

211
$$
N - RR \left[mg \ N L^{-1} d^{-1} \right] = \frac{c_{N0} \overline{a} - N(inf) - C_{N0} \overline{a} - N(eff)}{HRT}
$$
 (2)

212 Where C_{NO3} $\bar{N}(inf)$ and C_{NO3} $\bar{N}(eff)$ [mgNO₃ -N L⁻¹] are nitrate concentrations in the influent and 213 the effluent, respectively, while *HRT* [d] is the hydraulic retention time considering the 214 volumes of the cathodic and central compartments.

215 The desalination performance was evaluated by calculating the electrical conductivity 216 removal efficiency (EC-RE, equation 3), the chloride removal efficiency (Cl-RE, equation 217 4), and the chloride removal rate (Cl⁻-RR, equation 5).

218
$$
EC - RE \, [\%] = \frac{EC_{(inf)} - EC_{(eff)}}{EC_{(inf)}} \times 100 \tag{3}
$$

219
$$
Cl^- - RE [\%] = \frac{c_{Cl^-}(inf)^{-C_{Cl^-}(eff)}}{c_{Cl^-}(inf)} \times 100
$$
 (4)

$$
220 \t Cl- - RR [mg L-1 d-1] = \frac{c_{Cl-(inf)- C_{Cl-(eff)} }{HRT_{des}}
$$
(5)

221 where $EC_{(eff)}$ [mS cm⁻¹] and $Cci_{(eff)}$ [mg L⁻¹] represent the effluent electrical conductivity and chloride concentration, respectively. $EC_{(inf)}$ and $C_{CI}(inf)}$ correspond to the electrical conductivity and chloride concentration of the solution in the bio-cathode compartment (i.e., the influent to the central compartment), respectively, to consider the chloride input due to the acid dosage in this compartment. *HRTdes* [d] is the hydraulic retention time of the central compartment.

227 The coulombic efficiency for nitrate reduction ($\epsilon N O_x$) was calculated according to equation 228 6 (Virdis et al., 2008):

$$
229 \quad \varepsilon N O_x[\%] = \frac{I}{n \Delta C_{NO_x} Q_{inf} F} \times 100 \tag{6}
$$

230 where *I* is the fixed current [A]; *n* is the number of electrons that can be accepted by 1 mol 231 of oxidised nitrogen compounds present in the bio-cathode compartment, assuming N_2 is the 232 final product; *ΔCNOx* is the difference between the nitrate concentration in the cathodic 233 influent and effluent $[molNO₃ - N L⁻¹]$; Q_{inf} is the influent flowrate $[L s⁻¹]$; *F* is Faraday's 234 constant $[96485 \text{ Ce}^{-} \text{mol}^{-1}]$.

 The current efficiency (CE) was expressed as the percentage of the charge associated with the chloride removed from the central compartment to the amount of electric charge transferred (ECT) across the membranes (Ramírez-Moreno et al., 2019). CE [%] and ECT $\left[\text{C m}^{-3} \right]$ were calculated using equations 7 and 8, respectively:

239
$$
CE\left[\% \right] = \frac{v\ z\ F\left(c_{Cl^{-}(inf)} - c_{Cl^{-}(eff)}\right)}{ECT} \times 100
$$
 (7)

$$
240 \quad ECT \left[C \ m^{-3} \right] = \frac{\int I \ dt}{V} \tag{8}
$$

241 where *v* and *z* represent the stoichiometric coefficient and the valence of the chloride ion, 242 respectively; $V \text{ [m}^{-3}$ is the volume of water treated; *dt* is the time [s].

243 The specific energy consumption (SEC) was calculated according to equation 9 (Jingyu et 244 al., 2017):

 [ℎ −3] = ∫ 245 **(9)**

246 where E is the cell potential [V].

247 **2.6 Analysis of bacterial community by NGS of 16S rRNA gene**

 The composition of the bacterial community in the cathodic biofilm was characterised. Samples of the biofilms formed on the bio-cathode were axenically collected during Test 5 (Table 1). Five cathode points were sampled, and the biomass was pooled into a composite sample to mitigate the effects of microscale heterogeneity on the bio-cathode. Biomass samples were stored at -20°C before DNA extraction. Genomic DNA was extracted from biomass samples (250 mg wet weight) using the DNeasy PowerSoil Pro Kit (QIAGEN), and DNA was subsequently purified using the DNeasy PowerClean Cleanup Kit (QIAGEN). The DNA quality and concentration were determined on agarose gel using a DNA quantitation standard. DNA samples were submitted to Bio-Fab Research Srl (Rome, Italy) for sequencing of the V3-V4 region of the bacterial 16S rRNA gene on an Illumina Miseq 258 platform (Illumina, San Diego, CA) using 2×300 bp paired-end reads. The primer pair was used S-D-Bact-0341-b-S-17/S-D-Bact-0785-a-A-21 (Klindworth et al., 2013).

 For data processing, raw sequences were demultiplexed by the sequencing facility. Reads were trimmed to remove primer sequences using the CutAdapt version 3.5. Sequences were imported into Quantitative Insights into Microbial Ecology (QIIME 2) version 2020-11 (Bolyen et al., 2019). Using the DADA2 pipeline (Callahan et al., 2016), reads with ambiguous and poor-quality bases were discarded, good-quality reads dereplicated and denoised, and the paired reads merged. Chimeras and singletons were identified and removed from the dataset. DADA2 was used to produce alternative sequence variants (ASVs), thus obtaining a filtered ASV-abundance table. For each ASV, a representative sequence was used for taxonomy assignment against the Silva database release 138 (Quast et al., 2013). Richness was estimated as the number of observed ASV by using the vegan R package (Oksanen et al., 2019). Read count data were normalised by Cumulative Sum Scaling (CSS) transformation using the metagenomeSeq package (Paulson et al., 2013). The Bray-Curtis similarity index between samples was calculated.

3. RESULTS AND DISCUSSION

3.1 Effect of the HRT on denitrification and desalination performances

- The system's enhancement was tested by increasing the influent flowrate and, thus, reducing
- 276 the HRT. Figure 1 shows the average $NO₃$ -N loading and removal rates (N-LR and N-RR,
- respectively) at different influent flowrates.

Figure 1. Average trend in N-LR and N-RR with increasing influent 280 flowrates (Q_{inf}) .

282 The N-LR was increased from 23.6 ± 1.8 mgNO₃ -N L⁻¹d⁻¹ (Test 1) to 261 ± 16 mgNO₃ -N 283 L⁻¹d⁻¹ (Test 6), and the N-RR increased as well (from 16.9 ± 1.3 mgNO₃-N L⁻¹d⁻¹ to 284 130.8 \pm 14.7 mgNO₃ -N L⁻¹d⁻¹), though it did not follow the same linear trend as the N-LR. Such an increase in the N-RR was likely ascribed to improved hydrodynamics conditions which minimised the occurrence of nitrate and pH gradients in the bio-cathode compartment, with the corresponding increase in the denitrifying activity (Pous et al., 2017; Vilà-Rovira et al., 2015). The increase in nitrate removal rate with decreasing HRT was also observed in previous studies. Figure 2 compares the trend of N-RR versus the HRT observed in the current study with those reported by Pous et al. (2017) and Ceballos-Escalera et al. (2021), exploiting tubular systems with hydraulically connected anode and cathode compartments. Although the systems were highly heterogeneous in terms of configuration (3-chamber plate cell vs tubular cells), electrode materials (graphite felt vs granular graphite), and operating conditions (galvanostatic vs potentiostatic modes), the same mathematical model was able to fit the observed N-RR vs HRT relationship.

296
297 Figure 2. Comparison of nitrate removal rate (N-RR) trend versus the HRT observed in the present study with those reported by Pous et al. (2017) and Ceballos-Escalera et al. (2021), and modelling of results.

 This result is interesting as it proves that, regardless of the type of configuration or operating conditions used, the process behaviour with different HRTs may be reasonably predicted, providing useful information in the perspective of reactor scale-up.

 Despite the increasing N-RR observed in our study with decreasing HRT, nitrate concentration in the effluent started to increase from Test 4 onward (Figure 3). However, it 305 remained below the threshold level of 11.3 mgNO₃-N L⁻¹ (Nitrate Directive 91/767/EU) 306 throughout the experiment except during Test 6 (Q_{inf} , 1.42 L d⁻¹; HRT, 2.4 \pm 0.2 h), when an 307 average concentration of 13.5 ± 2.8 mgNO₃⁻-N L⁻¹ (corresponding to an N-RE of 50 \pm 8%) was observed. Since the applied current was initially much higher than that theoretically required to remove the nitrate input (10 mA applied vs approx. 1.4 mA theoretically required in Test 1), the coulombic efficiency for nitrate removal was always above 100%, decreasing as the HRT decreased, and reaching values close to 100% during Test 6.

313
314

314 Figure 3. Trends of average $NO₃$ -N concentration and electrical 315 conductivity (EC) in the effluent with increasing Q_{inf} .

 During Test 6, nitrite and nitrous oxide were detected in the effluent at low concentrations $(0.22 \pm 0.08 \text{ mgNO}_2\text{-N L}^{-1}$ and up to 0.5 mgN₂O-N L⁻¹, respectively). Pous et al. (2013) observed the formation of such stable intermediates in a denitrifying MFC treating 319 groundwater, and their presence was linked to reduced bacterial $NO₂$ - and $N₂O$ -reduction activities due to a change in the operating conditions (in that case, the influent composition). 321 Virdis et al. (2009) observed lower N₂O reduction rates in an MFC performing carbon and 322 nitrogen removal, compared to $NO₃^-$ reduction rates. Further tests suggested a possible 323 limitation through which the cathodic biofilm was not able to reduce N_2O at the same rate as it was produced by the previous step of denitrification. In our study, the further increase in nitrate removal rate observed in Test 6 was not fully accompanied by a corresponding 326 increase in nitrite and nitrous oxide removal rate, but still, 97% of NO₃ removed was converted into N₂. Such behaviour was related to the worsening of the denitrification process, therefore process boundary conditions were considered to lie in an HRT range between approximately 5 h (Test 5) and 2.5 h (Test 6).

 A final test (Test 7) was carried out setting the same HRT as Test 5 (i.e., 4.9±0.4 h) to restore the denitrifying process and verify microbial activity. The performance in terms of nitrate 332 removal observed during Test 5 (N-RE = $77\pm3\%$ and N-RR = 96.7 ± 2.8 mgNO₃-N L⁻¹d⁻¹) 333 was immediately restored during Test 7 (N-RE = $89\pm4\%$ and N-RR = 112 ± 7.5 mgNO₃-N 334 L⁻¹d⁻¹), and the average effluent NO₃⁻-N concentration was 3±1 mgNO₃⁻-N L⁻¹, far below the threshold limit for drinking water. No nitrite or nitrous oxide were detected in the effluent during Test 7, indicating that the worsening of nitrate removal with the production of intermediates observed during Test 6 was reversible. The slight increase in process performance observed between Test 5 and Test 7 demonstrates that biomass growth may have played a small role in improving the denitrification rates.

 An almost opposite behaviour was observed for the desalination process. The best 341 performance was observed during Test 1 when the average effluent conductivity was 2.2 ± 0.2 342 mS cm⁻¹, below the threshold level of 2.5 mS cm⁻¹ (98/83/CE Directive) (Figure 3). Figure 4 shows the trend of electrical conductivity in the influent and effluent of the central desalination compartment and the corresponding desalination efficiency (EC-RE). The decrease in overall EC-RE with decreasing HRT due to the reduced contact time was counterbalanced by the higher amount of influent treated per day. As expected, the overall 347 conductivity removal rate did not vary substantially throughout the experiment (23.4 ± 7.3) mS cm⁻¹d⁻¹). In fact, the salinity of simulated groundwater in our study was primarily related to different ions rather than nitrate (e.g., chloride, sodium, etc.), and desalination was mostly relying on physicochemical separation and electromigration rather than biological removal. Though the precipitation of cations such as Ca^{2+} and Mg^{2+} at high concentrations led to reduced power generation and desalination efficiency in MDCs treating saltwater and seawater (Luo et al., 2012; Rahman et al., 2021), much lower ions concentrations were involved in our study. Therefore the possible effects of ion precipitation on membrane ion-exchange capacity and overall desalination performance are reasonably expected to be much less significant, thus allowing a longer lifespan for the membranes. To enhance salinity removal and keep the conductivity below threshold levels, a valuable operating strategy may consist in increasing the applied current proportionally with the increase in influent flowrate.

Figure 4. Average trend of central compartment influent and effluent electrical conductivity, and desalination efficiency versus the influent flowrate. The red line 362 indicates the threshold limit for the electrical conductivity in freshwater (2.5 mS cm^{-1}) .

 The influent conductivity of the central desalination compartment (corresponding to the 364 effluent of the bio-cathode compartment) dropped from 8.7 ± 0.2 mS cm⁻¹ (Test 1) to 4.2 ± 0.6 365 mS cm^{-1} (Test 6), likely as a result of the increased influent flowrate which probably led to a faster turnover of the solution in the bio-cathode compartment, thus reducing the accumulation of chlorides dosed as HCl for pH control, and cations migrating from the central compartment through the CEM. The average chloride concentration measured in the cathode chamber (including the contribution of HCl dosage) decreased from 3622±443 (Test 370 1) to 1385 ± 56 mgCl⁻ L⁻¹ (Test 6), while sodium concentration decreased from 2355 ± 370 to 371 1040 ± 182 mgNa⁺ L⁻¹.

372 On the other hand, the effluent electrical conductivity slightly increased to 3.3 mS cm^{-1} , 373 resulting in the reduction of the overall desalination efficiency, which dropped from $77\pm13\%$ 374 (Test 1) to $12\pm2\%$ (Test 6). Coherently, the current efficiency related to the removal of 375 chloride in the central compartment decreased from $89\pm14\%$ (Test 1) to $59\pm15\%$ (Test 6). Such behaviour was related to the increase in the influent flowrate which resulted in too low 377 HRT_{des} (from 6.7 \pm 0.3 h in Test 1 to 0.5 \pm 0.02 h in Test 6). The theoretical quantity of chloride ions that can be transferred through the membranes by applying a current of 10 mA at the 379 different HRT tested ranges from 2.8 g L⁻¹ (Test 1) to 0.22 g L⁻¹ (Test 6), very close to those 380 actually observed (i.e., 2.5 ± 0.4 and 0.13 ± 0.03 gCl⁻removed L⁻¹ in Tests 1 and 6, respectively). Thus, the HRT decrease did not allow sufficient ions to migrate through the membranes to observe a significant reduction in effluent electrical conductivity. The adverse effect of low HRT on desalination performance was already demonstrated for MDCs by Jingyu et al. (2017), who reported that reducing the HRT leads to lower current generation and, consequently, to lower removal of total dissolved solids (TDS).

 Chlorine production in the anode compartment was monitored throughout the whole experimentation. An essential aspect of process monitoring is the durability of materials in contact with chlorine, as it is a powerful oxidant which may damage them. For this reason, it was decided to replace the solution in the anodic chamber periodically (about every 10 390 days, resulting in an average concentration of 16 ± 1 mgCl₂ L⁻¹). Higher values of chlorine 391 concentration (i.e., approximately 30 mgCl₂ L⁻¹) were obtained by Puggioni et al. (2021) without the periodic replacement of the solution, resulting in damage of reactors' materials.

3.2 Considerations on pH evolution during the process

 Increasing the influent flowrate also affected the pH trend in the different compartments. pH control plays a significant role in ensuring optimal denitrifying microbial activity, as a neutral pH is strictly necessary for this biological process (Clauwaert et al., 2009). Such control has become essential to optimise water desalination performance. Several studies demonstrated that the pH gradient between the anode and cathode compartments could lead to potential losses (Puig et al., 2012) that adversely affect the desalination efficiencies of MDCs (Jingyu et al., 2017).

 During the whole experiment, the periodic dosage of acid to control the pH in the bio-cathode compartment remained constant. This resulted in a difference mainly in the effluent pH as a function of the influent flowrate. As shown in Figure 5, while the influent pH remained 404 almost constant, the effluent pH increased from near-acidic (i.e., 4.1 ± 1.2 in Test 1) to slightly alkaline (i.e., 7.8±0.3 in Test 6).

408 Figure 5. Average influent and effluent pH trend versus the influent 409 flowrate.

 Near-acidic pH values in the effluent corresponding to the first Tests may be due to the 411 higher HRT_{des} (e.g., 6.7 ± 0.3 h in Test 1) that allowed protons produced at the anode (pH 412 2.0 \pm 0.7) to pass through the AEM, because of their small size. By reducing the HRT_{des} down to 0.5±0.02 h (Test 6), the faster solution replacement led to a slower pH increase in the

 bio-cathode compartment, and to a lower passage of protons through the AEM into the 415 effluent. In addition, the resulting dosage of HCl per $m³$ of treated water was significantly 416 reduced (from 109 L m⁻³ to 8.4 L m⁻³) as the influent flowrate increased, thus implying lower operating costs in the perspective of process scale-up.

3.3 Bacterial community diversity on the bio-cathode of the 3-compartment BES

 Cathodic biomass was collected during Test 5. The number of observed ASVs in the bacterial community was 99 and its composition is shown in Figure 6. The most abundant phyla of Bacteria in the biomass were Proteobacteria (44.0%) followed by Actinobacteriota (16.0%), Firmicutes (11.8%), Bacteroidota (10.8%), Planctomycetota (5.1%) and Chloroflexi (4.8%). The other less abundant phyla were all below the 3%, while the unassigned sequences accounted for 1.1% in the composition of bacterial community (Figure 6A). At order level (Figure 6B), the most abundant taxa were Rhizobiales (17.0%), Corynebacteriales (7.4%), and Burkholderiales (6.6%), followed by Xanthomonadales (4.5%), Alteromonadales (4.3%), and Thermomicrobiales (4.2%). At genus level (Figure 6C), the seven most abundant taxa accounted for more than 20% of the total community, including the genera *Rhizobium* (3.9%) and *Bosea* (3.1%) in Rhizobiales, *Mycobacterium* (3.2%) and *Gordonia* (2.4%) in Corynebacteriales, *Fontibacter* (2.6%) in Cytophagales, *Clostridium sensu strictu* (2.4%) in Firmicutes as well as the uncultured JG30-KF-CM45 in Thermomicrobiales (3.2%).

 An active role in denitrifying biomass has been previously proposed for several bacteria dominating the bio-cathodic biomass. More specifically, isolates affiliated to Rhizobiales have been proved to denitrify under autotrophic and heterotrophic conditions (Vilar-Sanz et al., 2108), and the genus *Rhizobium* has been implied in denitrification in MFC system for treating saline wastewater (Xu et al., 2019). *Clostridium sensu strictu* has been detected at a high amount in MEC biomass and suggested to be responsible for autotrophic denitrification in a bioelectrochemically-assisted constructed wetland system (Sotres et al., 2015; Xu et al., 2017). Recently, the genus *Fontibacter* has been found to be enriched after long-term adaptation in a BES for nitrate removal from coke wastewater effluent (Tang et al., 2017) and a species of the genus, isolated from an MFC, has been proved to couple oxidation of organic matter to Fe(III) reduction (Zhang et al., 2013). On the contrary, other dominant populations in the biomass, such as Corynebacteriales, have been less extensively described, and their metabolic role in bioelectrochemical systems is far to be undiscovered.

447 Figure 6. Bacterial community composition of the biofilm formed on the bio-cathode of the 3-compartment 448 bio-electrochemical cells under galvanostatic mode. Bar plots showing the contribution at phylum (A) and bio-electrochemical cells under galvanostatic mode. Bar plots showing the contribution at phylum **(A)** and

3.4 Sustainability perspective on the application of BES for simultaneous denitrification and desalination

 In order to move towards scaling up the proposed technology for groundwater treatment, the system must be both technically and economically feasible. For this reason, a preliminary and simplified cost-benefit analysis was carried out comparing the main operational costs associated with the technology and the potential benefits obtained according to experimental data.

 The operating costs of a technology depend significantly on the energy consumption of the process. Figure 7 shows the profiles of the specific energy consumption (SEC) per 459 gram of $NO₃ - N$ removed and per volume of water treated as a function of influent flowrate, compared with the trend in N-RR. During the experiment, an improvement of process performance was observed in terms of nitrate removal and SEC, the latter being 462 significantly reduced from $35.2 \cdot 10^{-2} \pm 3.6 \cdot 10^{-2}$ kWh g⁻¹NO₃-N_{removed} and 6.1 \pm 0.4 kWh $\rm m^{-3}$ _{water treated} (Test 1) to 5.1 \cdot 10⁻² ±0.7 \cdot 10⁻² kWh g⁻¹NO₃⁻N_{removed} and 0.5±0.03 kWh m⁻³_{water} 464 treated (Test 6).

 Figure 7. Average trends of SEC per gram of nitrate-nitrogen removed and per 467 volume of water treated, and of N-RR with increasing Q_{inf} .

 Pous et al. (2015) reported a list of SECs for various technologies such as bioelectrochemical systems (BES), biofilm electrode reactor (BER), membrane bioreactor (MBR), electrodialysis (ED) and reverse osmosis (RO) (Zhao et al., 2011; Twomey et al., 2010; McAdam and Judd, 2008; Ortiz et al., 2008). Compared to the 472 reported values, the energy consumption per $m³$ of treated water was within the range reported for desalination technologies, i.e., electrodialysis and reverse osmosis (0.04 - 2.09 kWh m⁻³ treated water). The energy consumption per gram of nitrate removed obtained in the present study was in line with those of the technologies reported for nitrate removal, 476 mainly BES and BER $(0.16 \cdot 10^{-2} - 7 \cdot 10^{-2} \text{ kWh } g^{-1} \text{NO}_3$ -N_{removed}). More specifically, the 477 values obtained in this study are closer to those of BER (i.e., $7 \cdot 10^{-2}$ kWh g⁻¹NO₃⁻-N_{removed}), which are based on the application of a potential difference between the electrodes. This type of catalytical operation produces hydrogen in the cathode chamber, which is then used by bacteria to reduce nitrate. In the present study, the current was fixed, and the potential established at the cathode (approximately -1.3 V vs Ag/AgCl) was suitable for hydrogen production. According to Pous et al. (2015), fixing the cathode potential makes it possible to control the reduction of nitrate to the end products and implies less energy consumption. In the present study, however, the aim was not only to remove nitrate but also to reduce the electrical conductivity of groundwater, as well as the production of value-added products (i.e., chlorine). In fact, during the process, part of the chloride accumulated in the solution of the anode compartment is converted into free chlorine (Puggioni et al., 2021). Thus, the energy applied is used to carry out three processes simultaneously with consumption comparable to systems carrying out a single process 490 (i.e., only denitrification or desalination). At the best HRT tested (i.e., 4.9 ± 0.4 h, Test 5), 491 the total cost of energy consumption was as low as $0.23 \text{ } \in \text{m}^3$, assuming an energy cost 492 of 0.21 ϵ kWh⁻¹ (Eurostat, 2021). This value is competitive, considering that

493 Ceballos-Escalera et al. (2022) estimated an operating cost of 0.14 ϵ m⁻³ for the removal of only nitrate using BES.

 From an economic point of view, the production of chlorine also plays an important role. Chlorine is a disinfectant agent that is highly used in water treatment plants, and its market value is growing significantly due to the rising demand from the agrochemical and pharmaceutical industries. Moreover, the rising demand for water treatment applications combined with increased awareness of better hygiene practices resulting from the impact of the Sars-CoV-2 pandemic will drive the need for chlorine among industrialists. Greaves et al. (2022) demonstrated that Sars-CoV-2 is successfully eliminated by disinfection with free chlorine in both deionised water and wastewater. Web-based chlorine market data show a forecast growth of the chlorine value at a CAGR (compound annual growth rate) between 3.5 and 4.5% for the period 2021-2027.

505 In the present study, up to 0.17 gCl₂ per gCl_{removed} were produced, and this production can easily be increased by switching to a continuous operating mode in the anodic chamber or by stripping the chlorine produced. In fact, Puggioni et al. (2021) showed higher production rates at the start of the batch that gradually decreased to a plateau over long periods of operation. Therefore, switching to a continuous operating mode would increase production rates while avoiding chlorine accumulation and excessive concentrations in the anode chamber. Optimising the chlorine capture system seems essential to maximise its production and reduce the contact time with the materials in the bioelectrochemical cell.

4. CONCLUSIONS

 At higher flowrates (and lower HRT, between 7.3±0.6 and 2.4±0.2 h), an increase in 517 intrate removal rates up to 131 mgNO₃-N L⁻¹d⁻¹ was observed. At an HRT of 2.4 \pm 0.2 h

 (Test 6), an effluent nitrate concentration above threshold levels for human consumption (91/767/EU) and the presence of intermediates were observed, indicating a worsening of the denitrifying performance of the system. Desalination performance was reduced in 521 terms of EC-RE (from $77\pm13\%$ to $12\pm2\%$), but the effluent electrical conductivity remained close to the threshold levels for human consumption (98/83/CE). In addition, the bacterial community in the cathodic biomass under galvanostatic mode was dominated by few populations, which were previously demonstrated to have an active role in denitrifying biomass. The tests carried out in the present study demonstrate the economic potential of the proposed technology thanks to the possibility of considerably reducing specific energy consumption while simultaneously increasing denitrification performance. Such a result was achieved by acting on the treated flowrate (i.e., by reducing hydraulic retention times) and not on the reactor volumes, which would imply additional costs in terms of materials and space. Finally, chlorine production represents an enormous potential for possible real application as it would reduce the costs of any on-site disinfection or, in general, an economic return if resold.

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