



UNICA

UNIVERSITÀ
DEGLI STUDI
DI CAGLIARI



Università di Cagliari

UNICA IRIS Institutional Research Information System

This is the Author's | **accepted : version of the following contribution:**

[inserire De Agostini Antonio; Caltagirone Claudia; Caredda Angela; Cicutelli Angela; Cogoni Annalena; Farci Domenica; Guarino Francesco; Garau Alessandra; Labra Massimo; Lussu Michele; Piano Dario; Sanna Cinzia; Tommasi Nicola; Vacca Andrea; Cortis Pierluigi., Heavy metal tolerance of orchid populations growing on abandoned mine tailings: A case study in Sardinia Island (Italy)., Ecotoxicology and Environmental Safety, 189, 2020, 110018

**The publisher's version is available at:
<https://doi.org/10.1016/j.ecoenv.2019.110018>**

When citing, please refer to the published version.

Manuscript Number:

Title: A population of *Epipactis helleborine* (L.) Crantz subsp. *tremolsii* (Orchidaceae) growing on mine tailings: a case of study in Sardinia (Italy).

Article Type: Research paper

Section/Category: Ecotoxicology

Keywords: Heavy metals; Orchids; Mycorrhiza; *Epipactis*; Soil pollution; Mining Area

Corresponding Author: Dr. Francesco Guarino,

Corresponding Author's Institution:

First Author: Antonio De Agostini

Order of Authors: Antonio De Agostini; Claudia Caltagirone; Alberto Caredda; Angela Cicutelli; Annalena Cogoni; Domenica Farci; Francesco Guarino; Alessandra Garau; Massimo Labra; Michele Lussu; Dario Piano; Cinzia Sanna; Nicola Tommasi; Andrea Vacca; Pierluigi Cortis

Abstract: A large population of the orchid *Epipactis helleborine* (L.) Crantz subsp. *tremolsii* grows on a tailing dump in the South-west of the Sardinia island (Italy). The ecological growth context is characterized by high levels of heavy metals and low organic matter content in the soil. To characterize the ecological features of this population growing in such extreme context, a morphological analysis was performed on twenty individuals, that have been then subjected to measures of heavy metals bioaccumulation (bioaccumulation factor) and translocation (translocation factor). Finally, the mycorrhizae associated to the roots of plants grown on contaminated site have been identified by mean of DNA barcoding. All data were compared to those obtained from individuals collected in a non-contaminated site (controls). Plants grown on contaminated site result to be smaller than controls, able to tolerate heavy metals in the soil and to accumulate and translocate them in their organs. Fungi belonging to the genus *Ilionectrya* and to the Ascomycota phylum were found as symbionts of plants both on contaminated or not sites, while an unidentified fungus was isolated from roots on contaminated site only. Results are discussed in terms of heavy metals resistance of orchid and of physiological and ecological mechanisms.

Suggested Reviewers: Stefano Castiglione
scastiglione@unisa.it

He is an expert of heavy metals accumulation in plant and phytoremediation.

Lorenzo Pecoraro
Tianjin University - China
lorenzo.pecoraro@gmail.com

Giovanni Scopece
University of Naples
giovanni.scopece@unina.it

Katarzyna Turnau
katarzyna.turnau@uj.edu.pl

Grażyna SZAREK-ŁUKASZEWSKA
g.szarek@botany.pl

Opposed Reviewers:

Research Data Related to this Submission

There are no linked research data sets for this submission. The following
reason is given:
Data will be made available on request

1 **A population of *Epipactis helleborine* (L.) Crantz subsp. *tremolsii* (Orchidaceae) growing on**
2 **mine tailings: a case of study in Sardinia (Italy).**

3 Antonio De Agostini¹, Claudia Caltagirone², Alberto Caredda¹, Angela Cicatelli³, Annalena Cogoni¹, Domenica Farci⁴,
4 Francesco Guarino*³, Alessandra Garau², Massimo Labra, Michele Lussu¹, Dario Piano¹, Cinzia Sanna¹, Nicola
5 Tommasi⁵, Andrea Vacca², Pierluigi Cortis¹.

6 ¹ *Department of Life and Environmental Sciences, University of Cagliari, Viale Sant'Ignazio 13, 09123, Cagliari (CA),*
7 *Italy;*

8 ² *Department of Chemical and Geological Sciences, University of Cagliari, Cittadella Universitaria (Blocco D) - S.S.*
9 *554 bivio per Sestu, 09042 Monserrato (CA), Italy;*

10 ³ *Department of Chemistry and Biology "A. Zambelli", University of Salerno, Via Giovanni Paolo II 132, 84084*
11 *Fisciano (SA), Italy.*

12 ⁴ *Department of Plant Physiology, Laboratory of Photobiology and Plant Physiology, University of Life Sciences,*
13 *Nowoursynowska Str. 159, 02-776 Warsaw, Poland.*

14 ⁵ *Department of biotechnology and bioscience, University of Milano-Bicocca, Piazza della Scienza 2, 20126 Milano*
15 *(MI), Italy.*

16 * Corresponding author Francesco Guarino, email address fguarino@unisa.it

17

18 **Abstract:**

19 A large population of the orchid *Epipactis helleborine* (L.) Crantz subsp. *tremolsii* grows on a
20 tailing dump in the South-west of the Sardinia island (Italy). The ecological growth context is
21 characterized by high levels of heavy metals and low organic matter content in the soil. To
22 characterize the ecological features of this population growing in such extreme context, a
23 morphological analysis was performed on twenty individuals, that have been then subjected to
24 measures of heavy metals bioaccumulation (bioaccumulation factor) and translocation
25 (translocation factor). Finally, the mycorrhizae associated to the roots of plants grown on
26 contaminated site have been identified by mean of DNA barcoding. All data were compared to those
27 obtained from individuals collected in a non-contaminated site (controls). Plants grown on
28 contaminated site result to be smaller than controls, able to tolerate heavy metals in the soil and to
29 accumulate and translocate them in their organs. Fungi belonging to the genus *Ilionectrya* and to the
30 Ascomycota phylum were found as symbionts of plants both on contaminated or not sites, while an
31 unidentified fungus was isolated from roots on contaminated site only. Results are discussed in
32 terms of heavy metals resistance of orchid and of physiological and ecological mechanisms.

33 **Keywords:**

34 Heavy metals; Orchids; Mycorrhiza; *Epipactis*; Soil pollution.

35

36 **1. Introduction**

37 The island of Sardinia (Italy), located in a central position in the western Mediterranean basin, is
38 characterized by high geological, ecological and biogeographic complexity. It is considered a hot-
39 spot of biodiversity, and its flora counts 295 endemic *taxa* (Fenu et al. 2014). The *Orchidaceae*
40 family is well represented: in fact, in spite of the island small area (24100.2 Km²), Sardinia houses
41 68 orchid species (Lai, 2008) of which five are endemics (Lussu 2018; Gögler et al., 2015). As it
42 concerns the Italian peninsula, the orchid species are 197, while the species ascribed to the
43 European continent are 529 (GIROS, 2016).

44 Orchids are forced to establish a mycorrhizal symbiosis to guarantee the supply of nutrients to the
45 developing embryo, since their seeds, whose length ranges from 0.1 to 0.5 mm and weight only few
46 µg, lack of endosperm. Symbiosis with soil fungi plays fundamental ecological roles in adult
47 individuals too, both providing nutrients in those genera which are not able to photosynthesize,
48 totally or partially such as *Limodorum*, *Neottia* etc. (Scrugli et al., 1991). Moreover, fungal
49 symbiosis protects individuals when environmental pollutants are present, as heavy metals and
50 metalloids common in abandoned mine sites (Shefferson et al., 2008; Jurkiewicz et al., 2001).

51 Mining activity in Sardinia reached a considerable intensity in the first half of the nineteenth
52 century, and in many cases, it was carried out without an appropriate management of the mining by-
53 products. For this reason, the numerous abandoned mining areas still represent today sources of
54 environmental pollution (Bacchetta et al., 2018; Jiménez et al., 2011; Vacca and Vacca, 2001), since
55 they are characterized by high presence of metallic and metalloid pollutants as Cu, Pb, Zn, Cd, Cr,
56 As and Sb (Cidu et al. 2014; Vacca and Vacca, 2001; Fanfani et al., 2000; Frau, 2000).

57 Generally, the sources of contamination originating from previous mining activity are represented
58 by extended sterile, and tailing dumps (Bacchetta et al., 2015; Vacca and Vacca, 2001) not
59 adequately stored but accumulated in heaps. Those matrices are very reactive and mobile due to
60 their chemical nature (of sulphides and sulphates) and to the very fine dimensions of the waste
61 material [from the gravel to the silt granulometry, according to De Waele and Pisano, (1998)].

62 Generally, the contaminated heaps in abandoned mining areas are not suitable for the colonization
63 by the majority of the vascular flora because of i) pollutants are present in high levels, ii) they
64 include poor and non-consolidated soils, with very low organic matter content, iii) vegetation
65 canopy is absent or very rare (Bacchetta et al., 2018; Jiménez et al., 2011). Nevertheless, metal-
66 tolerant or metallophyte *taxa* are able to colonize and grow in that very harsh environments such as
67 the described ones.

68 This study is focused on the species *Epipactis helleborine* (L.) Crantz subsp. *tremolsii*
69 (*Orchidaceae*), an Eurasiatic orchid, present till the southern Europe and introduced in the recent
70 past in North America. The studied population of this orchid counts almost one hundred individuals
71 growing on a mine tailings dump resulting from an intense extraction of Zn, Cu, and argentiferous
72 Pb (De Waele and Pisano, 1998) in Domusnovas (South-West Sardinia). In order to investigate the
73 ability of *E. helleborine* subsp. *tremolsii* to accumulate heavy metals in its organs, all individuals
74 within this population were characterized through morphometric and ecological approaches. Data
75 obtained from the population of *E. helleborine* subsp. *tremolsii* growing on mine tailings dump
76 were compared to those obtained from a population (control) collected in a non-contaminated site.
77 To detect and characterize the specific mycorrhizal symbiosis of studied orchids, able to influence
78 population survival capacity, mycorrhizal fungi associated to the roots of plants grown on
79 contaminated or not sites were studied by culture methods. However, the symbiont mycorrhizae
80 were also molecularly characterized on plants from both populations.

81 Then, the research aims were to: i) detect morphological differences among orchids growing in
82 contaminated and non-contaminated areas and evaluate if they can be indicators of a stress
83 condition;

84 ii) estimate the content and compartmentation of heavy metals in plants' organs; iii) investigate the
85 role of soil fungi and mycorrhizal interactions in tolerance towards soil pollution and heavy metals.

86

87 **2. Materials and methods**

88 *2.1 Study area*

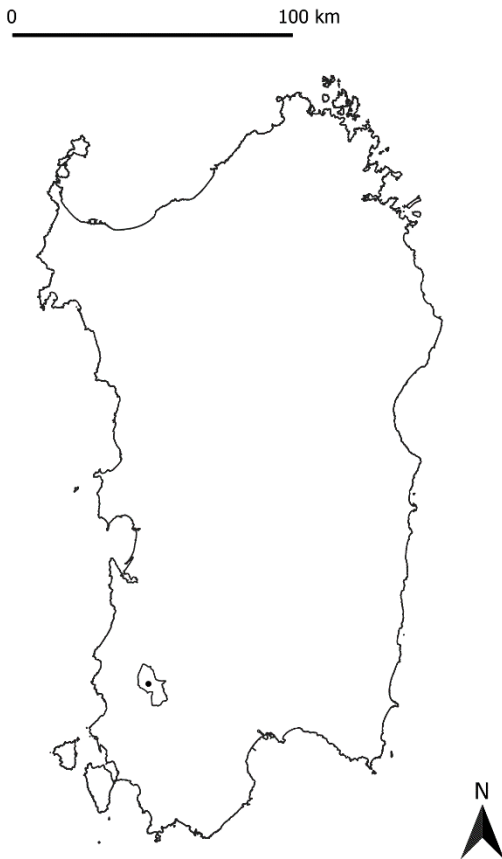
89 The abandoned mining site of “Barraxiutta” is located in the municipality of Domusnovas (South-
90 West Sardinia, Italy, Fig.1 A – B) where a mineralization of Sphalerite and Galena was exploited.

91 The heap where the studied population lives is located at 39°22'05.82" N, 8°36'28.46" E – WGS84;
92 while the control area is localized in the municipality of Nuoro, at coordinates 40°12'39.13" N,
93 8°41'14.43"E – WGS84.



94

95 Fig.1 A Map of Sardinia island, Italy.



96

97 Fig.1 B Map of the mining district of "Barraxiutta", municipality of Domusnovas.

98

99 *2.2 Soil sample collection and analyses*

100 In different sites of the study area (a flotation tailings dump), three selected topsoils (0-25 cm) were
101 described according to standard procedures of soil description (Schoeneberger et al., 2012). Soils
102 were sampled for physical and chemical analyses. The bulk soil samples were air dried and crushed
103 to pass a 2 mm sieve. Sand (2.00–0.05 mm), silt (0.050–0.002 mm) and clay (<0.002 mm) fractions
104 were separated by the sieve and pipette methods after the removal of organic matter by H₂O₂
105 treatment and dispersion aided by Na-hexametaphosphate. The organic carbon content was
106 determined by C elementary analyser (Leco, USA). Soil pH was measured by potentiometry in
107 soil/solution suspensions of 1:2.5 H₂O. The sieved samples for the determination of the metal total
108 content (Fe, As, Cd, Cu, Cr, Pb, Zn, Ni e Mn) were digested in concentrated HNO₃ according to the
109 EPA 3050-B method. For the determination of the metal bioavailable fractions, the Community

110 Bureau of Reference (BCR) extraction method (acetic acid 0.11 M) was used. The soil extracts were
111 analysed by an Inductively Coupled Plasma (ICP-OES 5110 Agilent).

112 *2.3 Plant sample collection and analysis*

113 The sampling of plant material was performed during the late spring 2018 from two populations of
114 *Epipactis helleborine* (L.) Crantz subsp. *tremolsii*: one localized on the tailing heap and the other
115 localized on the non-contaminated control area. From the first population a group of 20 random
116 individuals were selected to be compared with five individuals chosen from the control population.
117 Three morphometric parameters namely plant total height, inflorescence dimensions, length and
118 width of the bigger leaf were *in vivo* measured with metric tape, and after that, the individuals were
119 explanted. After removal, roots, leaves and stems from each individual were separated, cleaned and
120 dried in oven at 75° C up to a complete dehydration (24 hours). Subsequently, dried samples were
121 weighed, pulverized with liquid nitrogen and then digested with an acid mixture of 65% nitric acid
122 (HNO₃) and 50% fluoridric acid (HF) in a 2:1 ratio (v/v). Digestion was enhanced in a microwave
123 oven (Ethos, Milestone). Fe, Cu, Zn, Cd and Pb concentrations were determined by an Inductively
124 Coupled Plasma – Optical Emission Spectrometry (ICP-OES) on an Optima 7000DV (PerkinElmer)
125 and data were compared to a standard reference material (1575a Pine Needles; NIST, 2004) in order
126 to verify the accuracy. Standard solutions of each metal were also used in order to generate
127 calibration curves of emission readings vs concentrations.

128 *2.4 Bioaccumulation and translocation factors*

129 The Bioaccumulation factor (BAF) was calculated as the ratio between the concentration of a given
130 metal species in the root and the total or bioavailable fraction of the same metal species in the soil.
131 The values of this index indicate the capacity of the plant to accumulate (BAF > 1) or not (BAF <
132 1) metals in the roots. The translocation factor (TF) was also estimated in order to evaluate the
133 ability of the plant to translocate metals from the root to different epigeal parts. The translocation
134 factor (TF) is the ratio between the metal concentration in the epigeal portion of the plant and the
135 concentration of the same metal in the root. This index indicates the ability of the plant to

136 translocate metals from the root to the different epigeal parts such as stem, leaves, fruits etc. (TF >
137 1).

138 *2.5 Mycorrhizal fungi collection, cultivation and barcoding*

139 Mycorrhizal fungi were isolate from roots of different plants. Roots were collected, washed in
140 water, and sterilized with a solution of sodium ipochloride (1.15%) for 5 minutes. These steps
141 allowed to remove any possible contamination due to microorganisms present in the soil. After the
142 treatment, roots were longitudinally sectioned, and the cut-exposed surface was put on agarized
143 growth media. For this first step of fungal isolation a Potatoe Dextrose Agar media, added with
144 antibiotic (chloramphenicol 200 mg/mL), was used. Mycorrhizal fungi were grown for 7 days at
145 25°C before to be transferred on fresh culture media. Fungi were inoculated and grown on a
146 Sabouraud Dextrose Agar media added with antibiotic (chloramphenicol 200mg/mL). After 7-10
147 days of growth at 25°C, the plates were stored at -20°C.

148 The three isolated and cultivated mycorrhizal fungi were analysed trough DNA barcoding (Herbert
149 et al. 2003) analysis. Genomic DNA was isolated starting from 20 mg of culture medium from each
150 sample using Chelex® 100 Molecular Biology Grade Resin. Amplification of the nuclear internal
151 transcribed spacer region (ITS). was performed using puReTaq Ready-To-Go PCR beads
152 (Amersham Bioscience, Italy) in a 25-µL reaction according to the manufacturer's instructions and
153 primers ITS1F (CTTGGTCATTTAGAGGAAGTAA) and ITS4R (TCCTCCGCTTATTGATATGC)
154 from Luo et al (2002). PCR cycles consisted of an initial denaturation of 5 min at 95 °C followed by
155 32 cycles of denaturation (30 s at 95 °C) annealing (30 s at 58 °C) and extension (60 s at 72 °C) and
156 a final extension at 72 °C for 10 min. The obtained amplicons were isolated trough agarose gel
157 electrophoresis (1.5%) and purified from agarose using MinElute PCR Purification Kit (Qiagen,
158 Germany). Sequencing was performed by Macrogen Inc., Korea. Sequences were edited manually
159 and taxonomically assigned using blastn algorithm on GenBank (NCBI). Each sequence was
160 taxonomically assigned to the fungal taxon considering the nearest match (maximum identity >99%
161 and query coverage of 100%) according to Bruni et al. (2015). In case of multiple match with the

162 same threshold values, the sequence was assigned to the genus level.

163 2.6 Statistical analysis on plant morphometric data and metal concentration

164 A preliminary test to assay the Normal/Gaussian distribution, homogeneity variance and
165 homoscedasticity were performed on data in Rstudio through shapiro test, levene test and Bartlett
166 test, respectively. After that, morphometric data, metal concentration, and element accumulation in
167 different plant organs (in relation to the treatments) were tested in Rstudio by Kruskal and Wallis
168 one-way analysis of variance by ranks, followed by post hoc Nemenyi test (pgirmess package).

169

170 3. Results

171 3.1 Pedological and physicochemical soil features

172 The analyses on topsoils of dump revealed that they are characterised by a ^A horizon with sandy
173 texture (87.2% sand, 10.8% silt, and 2% clay), weak very fine and fine subangular blocky structure
174 with a tendency to single grain, soft, nonsticky, and nonplastic consistence, strong effervescence
175 after 1 NHCl application, organic carbon content of 0.53%, and pH (H₂O) equal to 7.8. Due to the
176 fact that they have formed on materials created by humans, as part of a mine process (mine spoils),
177 these topsoils belong to soils that are classified as Spolic Technosols (IUSS Working Group WRB,
178 2015).

179 Table 1 reports the total and the bioavailable concentration of Fe, As and heavy metals of the
180 collected topsoils. Zn and Pb are the elements with higher absolute values. With respect to the total
181 content, 60% of Cd, 49% of Pb, and 37% of Zn are bioavailable.

Elements	Mean concentration (mg g ⁻¹) ± Std. Error	Mean bioavailability (mg g ⁻¹) ± Std. Error
[Cr]	0.01 ± 0.01	l.o.d.
[Mn]	1.24 ± 0.10	0.22 ± 0.03
[Fe]	55.98 ± 3.24	0.05 ± 0.01

[Ni]	0.02 ± 0.01	l.o.d.
[Cu]	0.79 ± 0.08	0.01 ± 0.01
[Zn]	13.10 ± 0.9	4.87 ± 0.80
[Cd]	0.15 ± 0.01	0.09 ± 0.02
[Pb]	5.21 ± 0.35	2.57 ± 0,40
[As]	0.19 ± 0.05	l.o.d.

Tab. 1: Mean soil metal concentrations and bioavailability. l.o.d. = limit of detection.

182 *3.2 Morphometric parameters*

183 Individuals from the contaminated site (20) and controls (5) were analysed and compared in order
 184 to define morphometric differences. The plant height, inflorescence size, leaf length and width of
 185 individuals collected in non-contaminated site resulted significantly greater than those measured in
 186 the case of individuals sampled in the contaminated area (Tab. 2).

Morphometric parameters	Mean (cm) ± Std. Error (Contaminated soil)	Mean (cm) ± Std. Error (Control soil)	p-values
Height	20.65 ± 1.0	37.7 ± 1.2	< 0.05
Inflorescence size	3.575 ± 0.3	5.04 ± 0.4	< 0.05
Leaf length	6.535 ± 0.2	7.66 ± 0.2	> 0.05
Leaf width	3.36 ± 0.1	4.58 ± 0.2	< 0.05

Tab. 2: Morphometric data and comparison among orchids grown on contaminated or not-contaminated soils.

187 *3.3 Heavy metals content in plant organs, accumulation and translocation*

188 Fe, Cu, Zn, Cd, and Pb concentrations were detected in the organs of orchids grown on the tailing
 189 dump and control soils. Although metal concentrations were not very high, metal concentrations,
 190 mainly Fe and Zn, were higher in the organs of plants grown on dump than those grown on control
 191 soil (Tab. 3). Furthermore, the highest concentration of Fe and Zn, in fact, was measured in the

192 roots of orchids grown on polluted soil. In general, metal concentration in organs of plants grown in
193 the contaminated soil were significantly greater (even one magnitude higher) of those collected
194 from non-contaminated soil (Tab. 3).

Elements	Metal concentration in epigeal organs Mean (mg g ⁻¹) +/- Std. Error contaminated vs control individuals			Metal concentration in ipogea organ Mean (mg g ⁻¹) +/- Std. Error contaminated vs control individuals		
	S+L (Contaminated soil)	S+L (Control)	p-values	R (Contaminated soil)	R (Control)	p-values
Fe	15.36±0.3	2.71±0.20	8.081e-07 (t-test, Welch approx.)	43.08±0.19	17.09±0.04	< 0.005
Cu	0.26±0.05	0.02±0.01	0.5743 ('Mann-Whitney' test)	0.09±0.02	0.01±0.01	< 10e-05
Zn	8.52±0.19	0.23±0.02	7.123e-06 (t-test, Welch approx.)	23.28±0.9	0.25±0.03	< 10e-05
Cd	0.01±0.01	l.o.d.	//	0.09±0.02	l.o.d.	//
Pb	1.66±0.04	l.o.d.	//	4.10±0.10	l.o.d.	//

195 Legend: S = stem; L = leaves; R = root; S+L = stem + leaves; l.o.d. = limit of detection.

196 Tab. 3. Mean concentrations of Fe, Cu, Zn, Cd, and Pb in plant organs.

197 Bioaccumulation factors were calculated for plants collected in the contaminated area considering
 198 both the available metal soil concentration ($BAF_{bioav.}$), or the total one ($BAF_{tot.}$) (Tab. 4).
 199 In the case of BAF_{tot} the values were lower than 1 for all the detected metals, with the exception of
 200 Zn. Considering BAF_{bioav} the values were all greater than 1, with the exception of Cd. The Fe
 201 BAF_{bioav} was extremely high because of the very low Fe availability in the contaminated soil.

Elements	Mean $BAF_{tot.} \pm$ Std. Error	Mean $BAF_{bioav.} \pm$ Std. Error
Fe	0.77 ± 0.1	783.25 ± 52.7
Cu	0.12 ± 0.02	9.16 ± 1.54
Zn	1.78 ± 0.26	4.77 ± 0.71
Cd	0.58 ± 0.11	0.97 ± 0.18
Pb	0.79 ± 0.12	1.59 ± 0.24

202
 203 Tab. 4. BAF values considering total and bioavailable fractions of metals in the soil.

204 TF values for plants collected in the tailing dump, was shown to be >1 if we consider the epigeous
 205 portion of individuals (with the exception of Fe) (Tab. 6), also in the case of plant collected from
 206 non-contaminated soil certain TF values were higher than one, in particular in the case of Zn and
 207 Pb, whilst TF of Fe was lower than 1.

Elements	Mean TF \pm Std. Error					
	Epigeous		Leaves		Stem	
	Contaminated soil	Control	Contaminated soil	Control	Contaminated soil	Control
Fe	0.67 ± 0.29	0.15 ± 0.03	0.69 ± 0.39	0.06 ± 0.01	0.33 ± 0.16	0.10 ± 0.02
Cu	1.56 ± 1.34	1.10 ± 0.16	0.65 ± 0.43	0.56 ± 0.07	0.83 ± 0.75	0.54 ± 0.08
Zn	1.21 ± 0.86	2.20 ± 0.73	1.00 ± 0.57	0.95 ± 0.30	0.34 ± 0.11	1.25 ± 0.44
Cd	1.45 ± 1.25	//	0.69 ± 0.45	//	0.73 ± 0.64	//
Pb	1.93 ± 1.61	3.59 ± 0.93	1.60 ± 1.07	1.49 ± 0.36	0.32 ± 0.16	2.10 ± 0.60

208 Tab. 6. Translocation of metals in contaminated and control individuals (epigeous = stem + leaves).

209 3.4 Fungal barcoding

210 DNA extraction was carried out for all the samples and the whole amplification products showed a
 211 clear single band after electrophoresis (min-max length 502-550 bp). All the PCR products were
 212 sequenced, and high-quality bidirectional sequences were obtained. One of the sequences was
 213 taxonomically assigned to an unidentified endophyte fungus (GenBank reference sequence
 214 accession number AF373050.1) isolated from *Rosmarinus officinalis* L. roots (Girlanda et al. 2002).

215 Another sequence was assigned to the *Ascomycota* phylum identified by Vu et al. (2019) (GenBank
216 reference sequence accession number MH863168.1), while the last sequence was assigned to the
217 genus *Ilyonectria*.

218

219 **4. Discussion**

220 The habitat of the orchid *Epipactis helleborine* subsp. *tremolsii* is typically shady or mildly-shady
221 with deep and wet soils; nevertheless, this species can be found in parks, city gardens, and also in
222 ecologically-compromised sites interested by previous mining activity (Szarek-Łukaszewska, 2009;
223 Shefferson et al., 2008; Jurkiewicz et al., 2001; Richards and Swan, 1976).

224 In this study an orchid population growing on a soil derived from mining activity, characterized by
225 low organic matter and high metals concentration, was investigated in comparison with another
226 orchid population harvested in a non-contaminated soil. The contaminated site can be attributed to
227 the previous mining activity carried out in the area, that continued during almost one hundred years,
228 reaching production rates of 130 tons of *tout venant* with 60% in Pb (De Waele and Pisano, 1998).

229 The waste material and the flotation tailings produced during the mine activity were not properly
230 managed and still today present relevant contents of heavy metals and are characterized by high
231 environmental mobility (De Waele and Pisano, 1998). The total and the bioavailable content of Fe,
232 As, and heavy metals of the studied topsoils reflects the origin of the parent material. Zn and Pb are
233 very abundant in the whole area, the former being derived from sphalerite and the bulk of oxidised
234 products called “calamine”, and the latter being derived from both galena and from oxidation
235 minerals, like anglesite and cerussite. Cadmium is mostly related to Zn-minerals and follows its
236 abundance, with particular enrichments related to treatment plants and tailings areas. Arsenic is a
237 common element in some of the pyrites, especially those of the orebodies at the base of the
238 Cambrian carbonates. Consequently, as to be expected, total contents of Zn, Pb, and Cd in topsoil
239 samples are much higher than limits imposed by the Italian law (GURI 2006, D.lgs. 152) for sites of
240 commercial and industrial use (Zn = 1.5 mg g⁻¹, Pb = 1.0 mg g⁻¹, and Cd 0.015 mg g⁻¹). Higher

241 total values, with respect to the law limits, are also found for As and Cu, whose law limits are set at
242 0.05 mg g^{-1} and 0.6 mg g^{-1} , respectively. It should be noted that Zn and Pb total values in the
243 studied topsoils are also higher than the median values of stream sediments in the district (Zn = 1.2
244 mg g^{-1} and Pb = 0.95 mg g^{-1} , Boni et al., 1999), that can be taken as an indication of the local post-
245 mining geochemical baseline. Zinc was found to be the most bioavailable metal in the soil followed
246 by lead, and manganese. On the contrary, iron, despite the fact that is the most abundant metal in the
247 soil, is one of the less bioavailable. Some of the metals present in the soil such as iron, manganese,
248 nickel and copper are essential micronutrients for plants metabolism. However, concentrations
249 detected in the studied soil are significantly higher than in unpolluted soils, making a significant
250 stress factor for the majority of plants (Laghlimi et al., 2015). Other detected metals, aluminium,
251 cadmium, lead, and chromium are known only for their toxic effect on plants. In particular, their
252 phytotoxic effects bring to the alterations in photosynthesis, respiration, nutrient uptake, genic
253 expression, and membrane integrity (Rascio and Navari-Izzo, 2011; Laghlimi, 2015). In addition,
254 the high presence of cations in the soil could cause the saturation of the radical cation exchange
255 sites determining a reduced efficiency in the uptake of other important non-metal cations, such as
256 Ca^{2+} and Na^{+} . This fact in combination with the low amount of organic matter, drastically reduces
257 the ability of a balanced nutrient uptake for the plant.

258 It is evident that under these growth conditions the ecology of the studied population is expected to
259 be strongly affected. In fact, the results seem to detect a condition of stress in the orchid population
260 growing on the tailing dump, witnessed by the presence of smaller individuals (with respect to the
261 four morphometric parameters considered) and by the presence in plants' organs of metal pollutants.
262 The analysis of metal content in the individuals revealed the presence of iron, zinc and lead in roots,
263 while in stems and leaves are mainly accumulated iron and zinc, and lead. Those metals, that in the
264 case of iron and zinc are micronutrients (which are known to be toxic only at high levels), were
265 detected in the orchids grown on polluted soils together with lead, know for its phytotoxicity
266 (Amari et al., 2017; Lamhamdi etl al., 2011) indicating that the species *Epipactis helleborine* subsp.

267 *tremolsii* cannot avoid the uptake of metals present in the soils at high concentrations, and that their
268 concentrations in the organs are, into some extent, proportional to the concentration in the soil.
269 Tolerance to heavy metal pollution is common in different plant *taxa*, such as *Helianthus annuus* L.
270 (Cicatelli et al. 2017; Lin et al., 2003; Davies et al. 2001), *Zea mays* (Vigliotta et al. 2016; Li et al.,
271 2011; Tanyolaç et al., 2007), *Populus* (Di Lonardo et al. 2011; Cicatelli et al. 2010; Krpata et al.
272 2008), *Dittrichia* (Buscaroli et al. 2017; Guarino et al. 2017) etc. that, however, can be more or less
273 tolerant to soil heavy metal pollution depending on their genotype, the bioavailability of the
274 pollutants, the co-presence of elements in the substrate, radical symbiosis with fungi and bacteria
275 among others pedo-climatological, physical and chemical growth conditions.

276 The BAF values varied a lot if considering the $BAF_{tot.}$ or the $BAF_{bioav.}$ Taking into account that the
277 $BAF_{bioav.}$ is a parameter ecologically more relevant than the $BAF_{tot.}$, since it considers the fraction
278 clearly available to the plant, the values of bioaccumulation significantly increase. $BAF_{bioav.}$ values
279 show that all the analysed elements (with the exception for Cd) are accumulated in roots. Generally,
280 plants hold heavy metals in roots in order to protect the photosynthetic tissues from the toxic effects
281 of pollutants (Rascio and Navari-Izzo, 2011); However, translocation up to the epigeal portion of
282 the plant, showed by values of TF greater than one for lead, copper, and zinc (in particular lead and
283 zinc are translocated to the leaves), suggests that *E. helleborine* subsp. *tremolsii* is in some cases
284 able to translocate heavy metals in its epigeal organs. Considering that *E. helleborine* subsp.
285 *tremolsii* is a geophyte that, as the others Mediterranean orchids, loses stems and leaves after seeds
286 dispersion (GIROS, 2016), the translocation of metal pollutants to the epigeal portion could be an
287 active process carried out with the aim of detoxifying the organism across the vegetative season,
288 and storing heavy metals in the perennial part of the plant, the hypogeaal one. To verify this
289 hypothesis further investigation are needed, in particular regarding the intracellular location of
290 pollutant and their possible compartmentation in stems and leaves vacuoles or cell walls.

291 In this scenario of soil pollution, it's opportune to briefly discuss the features of plant-soil fungi
292 symbiotic interactions, especially strong in the case of terrestrial orchids (GIROS, 2016). Soil fungi

293 are known to establish symbiotic relationships with the roots of several plant species: the so-called
294 mycorrhizal symbiosis. The ecto- and endo- mycorrhizal symbiosis (fungal hyphae located
295 exteriorly or internally with respect to the radical cortical cells respectively) plays a key role in
296 facilitating nutrients uptake, but it can also provide protection from different kind of stress,
297 including heavy metal stress. Soil fungi in fact can chelate metals (Gadd, 1993; Tobin et al., 1984)
298 thanks to cell walls' physical and chemical properties: this ability is found in ectomycorrhizae
299 (Turnau and Dexheimer, 1995; Denny and Wilkins, 1987), ericoid mycorrhizae (Bradley et al.,
300 1982) and arbuscular mycorrhizal fungi (Gonzales-Chavez et al., 2002; Joner et al., 2000). Some of
301 these symbionts could also help a precipitating metals out of the mycelium by the production of
302 organic acids of the acid phosphatase (Turnau and Dexheimer, 1995) or by the production of
303 melanin-like pigments that can reduce the mobility of metals, preserving the plant from their toxic
304 effects (Gadd and De Rome, 1988). Because of those properties, the root endophytes of the orchids
305 growing in the polluted and control areas have been identified by mean of DNA barcoding.
306 Sequence analysis allowed the identification of the genus *Ilyonectria* in both contaminated and
307 control individuals, of an unidentified fungus isolated from *Rosmarinus officinalis* roots (Girlanda
308 et al. 2002) associated to roots of orchids on contaminated site, and of an endophyte belonging to
309 the *Ascomycota* phylum on both sites. However, the genus *Ilionectrya* has been described as
310 symbiont in different orchids' genera like *Paphiopedilum* (Han et al., 2016), *Pterostylis* (Obase and
311 Matsuda, 2014), *Microtis* (Frericks, 2016), *Calanthe* (Park et al., 2018) and *Epipactis* (Obase and
312 Matsuda, 2014). In Shefferson et al. 2008, studying *Epipactis* sp. populations in heavy metal
313 disturbed sites, the genus *Ilionectrya* was not reported as endophyte, but on the contrary, every
314 studied population seems to have different fungal symbionts (*Trichophaea woolhopeia*, *Geopora*
315 *cooperi*, *Chalara dualis* etc.).

316

317 **Conclusion**

318 Present study revealed that the individuals of *Epipactis helleborine* subsp. *tremolsii* of the studied

319 populations are able to tolerate and grow on soils polluted by heavy metals and metalloids, and that
320 they can also accumulate and translocate those elements in their organs. The association of the same
321 symbiont *taxa* to the roots of plants grown on polluted or unpolluted soils, suggests that probably
322 the metal-tolerance of the orchids, of the studied population, is not due to a specific fungal
323 symbiont, but rather to features of the genus *Epipactis*. Furthermore, the only detected difference
324 between analyzed populations was relative to the dimensions. Despite the tolerance to such extreme
325 conditions and the ability to accumulate and translocate pollutant of the soil, this species cannot be
326 considered a valid element in phytoremediation and phytostabilization plans, due to the low
327 biomass productivity. Nevertheless, the mechanisms of tolerance should be better studied in this
328 species.

329 **Acknowledgments**

330 The authors are grateful to the Chemical Laboratory of Agris Sardegna for the organic carbon
331 analyses. We acknowledge RAS/FBS (grant number: F72F16003080002) for funding.

332 **5. Rereferences**

333 Amari, T., Ghnaya, T., Abdelly, C., 2017. Nickel, cadmium and lead phytotoxicity and potential of
334 halophytic plants in heavy metal extraction. *S. Afr. J. Bot.* 111, 99-110.
335 <https://doi.org/10.1016/j.sajb.2017.03.011>.

336 Bacchetta, G., Boi, M.E., Cappai, G., De Giudici, G., Piredda, M., Porceddu, M., 2018. Metal
337 Tolerance Capability of *Helichrysum microphyllum* Cambess. subsp. tyrrhenicum Bacch., Brullo
338 and Giusso: A Candidate for Phytostabilization in Abandoned Mine Sites. *B. Environ. Contam. Tox.*
339 101(6), 758-765. <https://doi.org/10.1007/s00128-018-2463-9>.

340 Bacchetta, G., Cappai, G., Carucci, A., Tamburini, E., 2015. Use of native plants for the remediation
341 of abandoned mine sites in mediterranean semiarid environments. *B. Environ. Contam. Tox.* 94(3),
342 326-333. <https://doi.org/10.1007/s00128-015-1467-y>.

343 Boni, M., Costabile, S., De Vivo, B., Gasparrini, M., 1999. Potential environmental hazard in the
344 mining district of southern Iglesias (SW Sardinia, Italy). *J. Geochem. Explor.* 67(1), 417-430.

345 [http://dx.doi.org/10.1016/S0375-6742\(99\)00078-3](http://dx.doi.org/10.1016/S0375-6742(99)00078-3).

346 Bradley, R., Burt, A.J., Read, D.J., 1982. The biology of mycorrhiza in the Ericaceae. VIII. The role
347 of mycorrhizal infection in heavy metal resistance. *New Phytol.* 91, 197-209.

348 Buscaroli, A., Zannoni, D., Menichetti, M., Dinelli, E., 2017. Assessment of metal accumulation
349 capacity of *Dittrichia viscosa* (L.) Greuter in two different Italian mine areas for contaminated soils
350 remediation. *J. Geochem. Explor.* 182(B), 123-131. <https://doi.org/10.1016/j.gexplo.2016.10.001>.

351 Cicatelli A., Guarino F., Baldan E., Castiglione S.. 2017. Genetic and biochemical characterization
352 of rhizobacterial strains and their potential use in combination with chelants for assisted
353 phytoremediation. *ESPR*, 24(9), 8866-8878. <http://10.1007/s11356-016-7982-5>

354 Cicatelli A., Lingua G., Todeschini V., Biondi S., Torrigiani P., Castiglione S.. 2010. Arbuscular
355 mycorrhizal fungi restore normal growth in a white poplar clone grown on heavy metal-
356 contaminated soil, and this is associated with upregulation of foliar metallothionein and polyamine
357 biosynthetic gene expression. *Ann. Bot.* 106, 791-802. <https://10.1093/aob/mcq170>

358 Cidu, R., Biddau, R., Dore, E., Vacca, A., Marini, L., 2014. Antimony in the soil-water-plant system
359 at the Su Suergiu abandoned mine (Sardinia, Italy): Strategies to mitigate contamination. *Sci. Total.*
360 *Environ.* 1, 497-498:319-331. <https://doi.org/10.1016/j.scitotenv.2014.07.117>.

361 Di Lonardo, S., Capuana, M., Arnetoli, M., Gabbrielli, R., Gonnelli, C., 2011. Exploring the metal
362 phytoremediation potential of three *Populus alba* L. clones using an in vitro screening. *Environ.*
363 *Sci. Pollut. Res.* 18(1), 82-90. <https://doi.org/10.1007/s11356-010-0354-7>

364 De Waele, J., Pisano, M., 1998. Interazione fra attività mineraria ed un acquifero carsico: l'esempio
365 di Barraxiutta (Sardegna sud-occidentale). In *Convegno Nazionale sull'Inquinamento delle Grotte e*
366 *degli Acquiferi carsici e possibili ricadute sulla collettività* (pp. 195-209).

367 Denny, H.J., Wilkins, D.A., 1987. Zinc tolerance in *Betula* spp. III Variation in response to zinc
368 among ectomycorrhizal associates. *New Phytol.* 106(3), 535-544. [https://doi.org/10.1111/j.1469-](https://doi.org/10.1111/j.1469-8137.1987.tb00158.x)
369 [8137.1987.tb00158.x](https://doi.org/10.1111/j.1469-8137.1987.tb00158.x).

370 Fanfani, L., Caboi, R., Cidu, R., Cristini, A., Frau, F., Lattanzi, P., Zuddas, P., 2000. Impatto
371 ambientale dell'attività mineraria in Sardegna: studi mineralogici e geochemici. Rendiconti Del
372 Seminario Della Facoltà Di Scienze Università Di Cagliari. 70, 249-264.

373 Fenu, G., Fois, M., Cañadas, E.M., Bacchetta, G., 2014. Using endemic-plant distribution, geology
374 and geomorphology in biogeography: The case of Sardinia (mediterranean basin). Syst. Biodivers.
375 12, 181-193. <https://doi.org/10.1080/14772000.2014.894592>.

376 Frau, F., 2000. The formation-dissolution-precipitation cycle of melanterite at the abandoned pyrite
377 mine of Genna Luas in Sardinia, Italy: environmental implications. Mineral Mag. 64(6), 995-1006.
378 <https://doi.org/10.1180/002646100550001>.

379 Frericks, J., 2016. The effects of endophytic fungi of NZ terrestrial orchids: developing methods for
380 conservation. Master in Ecology and Biodiversity. Victoria University of Wellington.

381 G.I.R.O.S., 2016. Orchidee d'Italia. Guida alle orchidee spontanee. Il Castello.

382 Gadd, G. M., De Rome, L., 1988. Biosorption of copper by fungal melanin. Appl. Microbiol.
383 Biotechnol. 29(6), 610-617. 10.1007/BF00260993.

384 Gadd, G. M., 1993. Interactions of fungi with toxic metals. New Phyt. 124(1), 25-60.
385 <https://doi.org/10.1111/j.1469-8137.1993.tb03796.x>.

386 Girlanda, M., Ghignone, S., Luppi, A.M., 2002. Diversity of sterile root- associated fungi of two
387 Mediterranean plants. New Phyt. 155(3), 481-498. [https://doi.org/10.1046/j.1469-](https://doi.org/10.1046/j.1469-8137.2002.00474.x)
388 [8137.2002.00474.x](https://doi.org/10.1046/j.1469-8137.2002.00474.x).

389 Gögler, J., Stökl, J., Cortis, P., Beyrle, H., Barone Lumaga, M.R., Cozzolino, S., Ayasse, M., 2015.
390 Increased divergence in floral morphology strongly reduces gene flow in sympatric sexually
391 deceptive orchids with the same pollinator. Evol. Ecol. 29(5), 703-717.
392 <https://doi.org/10.1007/s10682-015-9779-2>.

393 Gonzales-Chavez, C., Harris, P.J., Dodd, J., Meharg, A.A., 2002. Arbuscular mycorrhizal fungi
394 confer enhanced arsenate resistance on *Holcus lanatus*. New Phyt. 155(1), 163-171.
395 <https://doi.org/10.1046/j.1469-8137.2002.00430.x>.

396 Guarino F., Conte B., Improta G., Sciarrillo R., Castiglione S., Cicatelli A., Guarino C.. 2018.
397 Genetic characterization, micropropagation, and potential use for arsenic phytoremediation of
398 *Dittrichia viscosa* (L.) Greuter. *Ecotoxicol. Environ. Saf.* 148, 675-683,
399 <http://10.1016/j.ecoenv.2017.11.010>

400 GURI, 2006. Norme in materia ambientale, Decreto Legislativo 3 aprile 2006, n.152, Supplemento
401 Ordinario n.96, alla Gazzetta Ufficiale n.88, del 14 aprile 2006.

402 Han, J.Y., Xiao, H.F., Gao, J.Y., 2016. Seasonal dynamics of mycorrhizal fungi in *Paphiopedilum*
403 *spicerianum* (Rchb.f.) Pfitzer - A critically endangered orchid from China. *Glob. Ecol. Conserv.* 6,
404 327-338. <https://doi.org/10.1016/j.gecco.2016.03.011>.

405 Hebert, P.D.N., Cywinska, N.A., Ball, S.L., deWaard, J.R., 2003. Biological identifications through
406 DNA barcodes. *Proceedings of the Royal Society B: Biological Sciences* 270: 313–321

407 IUSS Working Group WRB., 2015. World Reference Base for Soil Resources 2014, update 2015,
408 International soil classification system for naming soils and creating legends for soil maps. World
409 Soil Resources Reports No. 106. FAO, Rome.

410 Jiménez, M.N., Bacchetta, G., Casti, M., Navarro, F.B., Lallena, A.M., Fernández-Onoño, E.,
411 2011. Potential use in phytoremediation of three plant species growing on contaminated mine-
412 tailing soils in Sardinia. *Ecol. Eng.* 37(2), 392-398. <https://doi.org/10.1016/j.ecoleng.2010.11.030>.

413 Li, T., Liu, M.J., Zhang X.T., Zhang, H.B, Sha, T., Zhao, Z.W., 2011. Improved tolerance of maize
414 (*Zea mays* L.) to heavy metals by colonization of a dark septate endophyte (DSE) *Exophiala*
415 *pisciphila*. *Sci. Total Environ.* 409(6), 1069-1074. <https://doi.org/10.1016/j.scitotenv.2010.12.012>.

416 Lin, J., Jiang, W., Liu, D., 2003. Accumulation of copper by roots, hypocotyls, cotyledons and
417 leaves of sunflower (*Helianthus annuus* L.). *Bioresour. Technol.* 86(2), 151-155.
418 [https://doi.org/10.1016/S0960-8524\(02\)00152-9](https://doi.org/10.1016/S0960-8524(02)00152-9).

419 Joner, E.J., Briones, R., Leyval, C., 2000. Metal-binding capacity of arbuscular mycorrhizal
420 mycelium. *Plant soil.* 226, 227-234. [10.1023/A:1026565701391](https://doi.org/10.1023/A:1026565701391).

421 Jurkiewicz, A., Turnau, K., Mesjasz-Przybyowicz, J., Przybyowicz, W., Godzik, B., 2001. Heavy

422 metal localisation in mycorrhizas of *Epipactis atrorubens* (Hoffm.) Besser (Orchidaceae) from zinc
423 mine tailings. *Protoplasma*. 218(3-4), 117-124. <https://doi.org/10.1007/BF01306601>.

424 Krpata, D., Peintner, U., Langer, I., Fitz, W.J., Schweiger, P., 2008. Ectomycorrhizal communities
425 associated with *Populus tremula* growing on a heavy metal contaminated site. *Mycological*
426 *Research*. 112(9), 1069-1079. <https://doi.org/10.1016/j.mycres.2008.02.004>.

427 Laghlimi, M., Baghdad, B., El Hadi, H., Bouabdli, A., 2015. Phytoremediation Mechanisms of
428 Heavy Metal Contaminated Soils: A Review. *Open J. Ecol.* 5(8), 375-388.
429 <https://doi.org/10.4236/oje.2015.58031>.

430 Lamhamdi, M., Bakrim, A., Aarab, A., Lafont, R., Sayah, F., 2011. Lead phytotoxicity on wheat
431 (*Triticum aestivum* L.) seed germination and seedlings growth. *C. R. Biol.* 334(2), 118-126.
432 <https://doi.org/10.1016/j.crv.2010.12.006>.

433 Lai, R., 2008. Aggiornamento corologico, tassonomico, nomenclaturale della flora orchidologica
434 della Sardegna. Dottorato di Ricerca in Botanica Ambientale e Applicata. Ciclo XX. Università
435 degli Studi di Cagliari.

436 Luo, G., and Mitchell, T.G., 2002. Rapid identification of pathogenic fungi directly from cultures by
437 using multiplex PCR. *J. Clin. Microbiol.* 40(8), 2860-2865. 10.1128/JCM.40.8.2860-2865.2002.

438 Lussu, M., De Agostini, A., Marignani, M., Cogoni, A., Cortis, P., 2018. *Ophrys annae* and *Ophrys*
439 *chestermanii*: an impossible love between two orchid sister species. *Nord. J. Bot.* 36(10).
440 10.1111/njb.01798.

441 Obase, K., Matsuda, Y., 2014. Culturable fungal endophytes in roots of *Enkianthus campanulatus*
442 (Ericaceae). *Mycorrhiza*. 24(8), 635-644. <https://doi.org/10.1007/s00572-014-0584-5>.

443 Park, M.S., Eimes, J.A., Oh, S.H., Suh, H.J., Oh, S.Y., Lee, S., Park, K.H., Kwon, H.J., Kim, S.,
444 Lim, Y.W., 2018. Diversity of fungi associated with roots of *Calanthe* orchid species in Korea. *J.*
445 *Microbiol.* 56(1), 49-55. <https://doi.org/10.1007/s12275-018-7319-9>.

446 R Core Team (2019). R: A language and environment for statistical computing. R Foundation for
447 Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/>.

448 Rascio, N., Navari-Izzo, F., 2011. Heavy metal hyperaccumulating plants: How and why do they do
449 it? And what makes them so interesting? *Plant Sci.* 180(2), 169-181.
450 <https://doi.org/10.1016/j.plantsci.2010.08.016>.

451 Richards, A.J., Swan, G.A., 1976. *Epipactis leptochila* (Godfery) Godfery and *E. phyllanthes* G. E.
452 Srn. occurring in South Northumberland on lead and zinc soils. *Watsonia.* 11, 1-5.

453 Schoeneberger, P.J., Wysocki D.A., Benham E.C., and Soil Survey Staff., 2012. Field book for
454 describing and sampling soils, Version 3.0. Natural Resources Conservation Service. National Soil
455 Survey Center. Lincoln, NE.

456 Scrugli, A., Cogoni, A., Riess, S., 1991. Beitrag zur Kenntnis Mykorrhiza in der Gattung
457 *Limodorum* Boehmer in C. G. Ludwig (Orchidaceae): *Limodorum trabutianum* Battand. *Die*
458 *Orchidee.* 42(2), 99-103.

459 Shefferson, R.P., Kull, T., Tali, K., 2008. Mycorrhizal interactions of orchids colonizing Estonian
460 mine tailings hills. *Am. J. Bot.* 95(2), 156-164. <https://doi.org/10.3732/ajb.95.2.156>.

461 Szarek-Łukaszewska, G., 2009. Vegetation of reclaimed and spontaneously vegetated Zn-Pb mine
462 wastes in Southern Poland. *Pol. J. Environ.* 18(4), 717-733.

463 Tanyolaç, D., Ekmekçi, Y., Ünalın, S., 2007. Changes in photochemical and antioxidant enzyme
464 activities in maize (*Zea mays* L.) leaves exposed to excess copper. *Chemosphere.* 67(1), 89-98.
465 <https://doi.org/10.1016/j.chemosphere.2006.09.052>.

466 Tobin, J.M., Cooper, D.G., Neufeld, R.J., 1984. Uptake of metal ions by *Rhizopus arrhizus* biomass.
467 *Appl. Environ. Microbiol.* 47, 821-824.

468 Turnau, K., Dexheimer, J., 1995. Acid phosphatase activity in *Pisolithus arrhizus* mycelium treated
469 with cadmium dust. *Mycorrhiza.* 5(3), 205-211.

470 Vacca, A., Vacca, S., 2001. Soil degradation in Sardinia – Historical causes and current processes
471 due to anthropogenic pressure. *Petermanns Geographische Mitteilungen* 145, 68-78.

472 Vigliotta G., Matrella S., Cicatelli A., Guarino F., Castiglione S.. 2016. Effects of heavy metals and
473 chelants on phytoremediation capacity and on rhizobacterial communities of maize. *J. Environ.*

474 Manage., vol. 179, p. 93-102, ISSN: 0301-4797, doi: 10.1016/j.jenvman.2016.04.055
475 Vu, D., Groenewald, M., De Vries, M., Gehrman, T., Stielow, B., Eberhardt, U., Al-Hatmi A.,
476 Groenewald, J.Z., Cardinali, G., Houbraken, J., Boekhout, T., 2019. Large-scale generation and
477 analysis of filamentous fungal DNA barcodes boosts coverage for kingdom fungi and reveals
478 thresholds for fungal species and higher *taxon* delimitation. Stud. Mycol. 92, 135-154.
479 10.1016/j.simyco.2018.05.001.