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Association of polygenic score and the involvement of cholinergic and glutamatergic pathways with lithium treatment response in patients with bipolar disorder

Azmeraw T. Amare ¹ et al.

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Lithium is regarded as the first-line treatment for bipolar disorder (BD), a severe and disabling mental health disorder that affects about 1% of the population worldwide. Nevertheless, lithium is not consistently effective, with only 30% of patients showing a favorable response to treatment. To provide personalized treatment options for bipolar patients, it is essential to identify prediction biomarkers such as polygenic scores. In this study, we developed a polygenic score for lithium treatment response (Li^+_{PGS}) in patients with BD. To gain further insights into lithium's possible molecular mechanism of action, we performed a genome-wide gene-based analysis. Using polygenic score modeling, via methods incorporating Bayesian regression and continuous shrinkage priors, Li^+_{PGS} was developed in the International Consortium of Lithium Genetics cohort (ConLi⁺Gen: $N = 2367$) and replicated in the combined PsyCourse ($N = 89$) and BipoLife ($N = 102$) studies. The associations of Li^+_{PGS} and lithium treatment response — defined in a continuous ALDA scale and a categorical outcome (good response vs. poor response) were tested using regression models, each adjusted for the covariates: age, sex, and the first four genetic principal components. Statistical significance was determined at $P < 0.05$. Li^+_{PGS} was positively associated with lithium treatment response in the ConLi⁺Gen cohort, in both the categorical ($P = 9.8 \times 10^{-12}$, $R^2 = 1.9\%$) and continuous ($P = 6.4 \times 10^{-9}$, $R^2 = 2.6\%$) outcomes. Compared to bipolar patients in the 1st decile of the risk distribution, individuals in the 10th decile had 3.47-fold (95%CI: 2.22–5.47) higher odds of responding favorably to lithium. The results were replicated in the independent cohorts for the categorical treatment outcome ($P = 3.9 \times 10^{-4}$, $R^2 = 0.9\%$), but not for the continuous outcome ($P = 0.13$). Gene-based analyses revealed 36 candidate genes that are enriched in biological pathways controlled by glutamate and acetylcholine. Li^+_{PGS} may be useful in the development of pharmacogenomic testing strategies by enabling a classification of bipolar patients according to their response to treatment.

Molecular Psychiatry (2023) 28:5251–5261; <https://doi.org/10.1038/s41380-023-02149-1>

INTRODUCTION

Bipolar disorder (BD) is a severe and often disabling mental health disorder that affects more than 1% of the population worldwide and is characterized by recurrent episodes of depression and mania [1]. BD accounted for 9.3 million disability-adjusted life years (DALYs) in 2017, and imposes a significant social and economic burden on society and healthcare systems [2, 3]. BD is associated with a significant somatic and psychiatric comorbidity [1] and an increased risk of suicide [4].

Since the discovery of lithium's mood-stabilizing property in 1949 [5], it has been widely used as a first-line therapy for patients with BD [6, 7]. Lithium is effective in treating acute episodes of illness and reduces the risk of future recurrences of mania and depression [8]. It has also been shown to reduce the risk of suicide [9]. Despite these merits, the efficacy of lithium is highly variable, with about 30% of treated patients showing a favorable response while more than 30% of them have no clinical response at all [8, 10]. Thus far, the causes and predictors of such heterogeneity in treatment response are insufficiently understood.

Genetic factors are thought to contribute, at least in part, to the large interindividual differences in response to lithium [10–15]. So far,

only a few genetic studies have identified specific single nucleotide polymorphisms (SNPs) and candidate genes associated with patients' response to lithium or treatment-related side effects [10, 11, 13–16]. Each employing a genome-wide association study (GWAS) approach, the Taiwan Bipolar Consortium found SNPs in the introns of *GADL1* associated with lithium treatment response [17], whereas the International Consortium on Lithium Genetics (ConLi⁺Gen) identified a locus on chromosome 21 [10], and a follow-up analysis uncovered additional variants within the human leukocyte antigen (HLA) region [14, 16]. Gene expression analysis of ConLi⁺Gen data also showed overexpression of genes involved in mitochondrial functioning in lithium responder patients, highlighting the electron transport chain as a potential target of lithium [18].

In our recent work, we applied a polygenic score (PGS) modeling approach and demonstrated associations between a poor response to lithium and a high genetic loading for schizophrenia (SCZ) [14], major depression (MD) [13], or a meta-PGS combining both SCZ and MD [15]. Machine-learning models that combined clinical variables with the PGS of SCZ and MD has further improved the prediction of lithium treatment response, explaining 13.7% of the variance [19].

A full list of authors and their affiliations appears at the end of the paper. email: azmeraw.amare@adelaide.edu.au

Received: 12 February 2023 Revised: 31 May 2023 Accepted: 16 June 2023

Published online: 11 July 2023

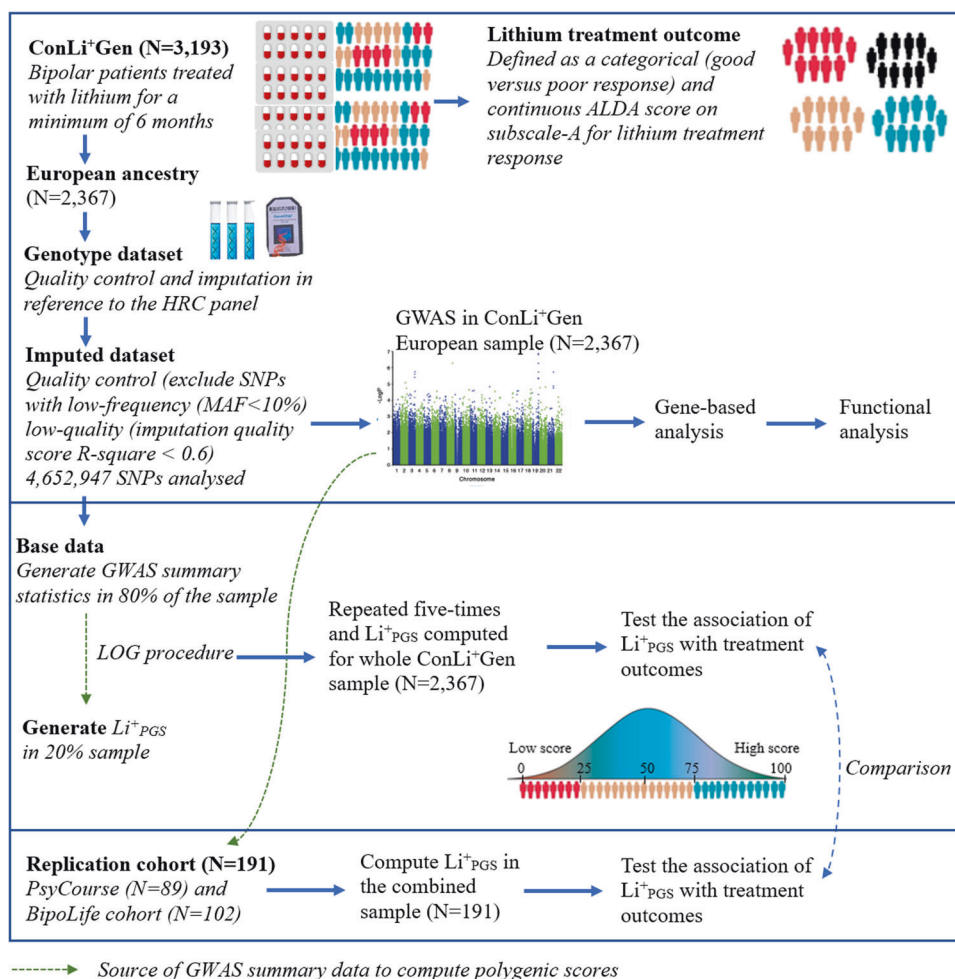


Fig. 1 Overview of input datasets and steps of data analyses. ConLi+Gen = The International Consortium on Lithium Genetics, ALDA = Retrospective Criteria of Long-Term Treatment Response in Research Subjects with Bipolar Disorder scale, HRC = Haplotype Reference Consortium, SNPs = Single Nucleotide Polymorphisms, MAF=Minor Allele Frequency, GWAS = Genome Wide Association analysis, Li+PGS = Polygenic score for lithium treatment response, LOG = Leave-one-group out procedure; PsyCourse = Pathomechanisms and Signature in the Longitudinal Course of Psychosis study and BipoLife = German research consortium for the study of bipolar disorder.

Based on these previous results, translation of PGS testing into clinical practice requires the consideration of three important learnings. First, the PGS of a single phenotype (e.g., SCZ or MD) explains only a small proportion (<2%) of the variability to treatment response in patients with BD [13, 14], providing insufficient power for clinical use. Second, a meta-PGS from multiple related phenotypes has better predictive power than a PGS from a single phenotype [15], suggesting the need to explore additional biological markers, including additional PGSs, that can either independently or together with existing PGSs better predict lithium treatment response. Third, developing polygenic markers with *direct* pharmacogenomic implications is essential, for example, a PGS for lithium treatment response (Li+PGS), which is perhaps biologically more related to lithium's pharmacological actions than PGSs built for other clinical phenotypes (i.e., SCZ or MD; that may indirectly influence treatment response or symptom severity, but do not index pharmacogenetic signatures per se).

Here, we developed a novel Li+PGS for lithium treatment response and applied gene-based pathway analyses to identify molecular mechanisms impacted by genetic variation in response phenotypes. Findings may assist in optimizing and personalizing the selection of mood stabilizers in patients with BD, and may point to novel molecular targets for future drug development.

METHODS AND MATERIALS

Study samples

For this study, we obtained genetic and clinical data from the International Consortium on Lithium Genetics (ConLi+Gen: $N = 2367$), Pathomechanisms and Signature in the Longitudinal Course of Psychosis study (PsyCourse: $N = 89$), and BipoLife cohort ($N = 102$). Figure 1 shows the detailed steps of data analysis.

Discovery cohort

ConLi+Gen is a global collaboration of scientists established to study the pharmacogenomics of lithium treatment in patients with BD [10]. In the current study, we analyzed the genome-wide genotype and clinical data of 2367 lithium-treated bipolar patients of European ancestry collected by 22 participating sites in 13 countries, including Australia ($n = 122$), Austria ($n = 43$), Czech Republic ($n = 45$), France ($n = 210$), Germany ($n = 218$), Italy ($n = 255$), Poland ($n = 97$), Romania ($n = 152$), Spain ($n = 74$), Sweden ($n = 304$), Switzerland ($n = 57$), Canada ($n = 353$) and the USA ($n = 437$) [10, 20].

Replication cohort

To replicate Li+PGS associations found in the discovery ConLi+Gen sample, we utilized datasets from PsyCourse and BipoLife where the study participants were of European ancestry. PsyCourse is a longitudinal multicenter study conducted from 2012 to 2019 in Germany and Austria, with up to four assessments at 6 monthly intervals. The study comprises 1320 patients from psychotic-to-affective spectrum, of which, datasets

from 89 patients with BD who received lithium treatment were obtained for this study [21]. *BipoLife* is a multicenter cohort study, established to investigate the biological basis of BD and patients' response to treatment and being conducted across ten university hospitals in Germany (Berlin, Bochum, Dresden, Frankfurt, Göttingen, Hamburg, Heidelberg, Marburg, Munich and Tübingen) and the medical informatics section of the University of Göttingen [22].

Target outcome

In both discovery and replication cohorts, patient's treatment response was assessed using the "Retrospective Criteria of Long-Term Treatment Response in Research Subjects with Bipolar Disorder" scale, also called the ALDA scale [10]. The target outcome "lithium treatment response" was defined in categorical and continuous scales among patients who had received lithium for a minimum of 6 months [10]. In both the discovery ConLi⁺Gen cohort and the replication cohorts (PsyCourse and BipoLife), a minimum of 6 months of lithium treatment follow-up was implemented as an inclusion criterion. This duration was chosen based on previous analyses of clinical trials, which established that a 6-month follow-up period is appropriate for assessing the minimum efficacy of lithium in patients with bipolar disorder [23]. Furthermore, clinical guidelines highly recommended to regularly monitor lithium levels during the initial six months of treatment, as this period is characterized by potential variability in lithium concentrations and an increased likelihood of side effects. After the six-month mark, stable lithium concentrations are typically achieved, allowing for an evaluation of the risk of toxicity and patients' adherence to treatment. These factors ultimately influence the effectiveness of the treatment [24–26]. The detailed procedures of ALDA scale measurement and its validity are described elsewhere [13, 14, 20]. Briefly, the ALDA scale consists of two subscales: the A scale and the B scale. The A scale measures the response to lithium treatment on a continuum ranging from 0 to 10. Assessors evaluate the change in illness activity while the patient is receiving lithium, and the response is rated accordingly. The anchor points for the A scale range from no change or worsening (score = 0) to complete response, which includes no recurrences during adequate treatment, no residual symptoms, and full functional recovery (score = 10). On the other hand, the B scale describes five factors that could potentially confound the response to lithium treatment or the interpretation of its magnitude. These factors are the number and frequency of episodes before starting lithium (B1 and B2, respectively), the duration of lithium treatment (B3), adherence to the prescribed lithium regimen (B4), and the use of additional medications (B5). Each item on the B scale is rated on a scale of 0 to 2, with a higher B score indicating a lower level of confidence that any observed clinical improvement is solely due to lithium [27]. Once we calculated the total score as 'A-score minus B-score and setting negative scores to zero', the categorical (good versus poor) lithium treatment response was defined at a cut-off score of 7, where patients with a total score of 7 or higher were considered as "responders" [10]. The continuous outcome for lithium treatment response was defined on subscale-A, but patients with a total B score greater than 4 or who had missing data on the totals of ALDA subscale-A or B were excluded [10].

Genotyping, quality control and imputation. We obtained the genotype data assayed with different types of commercial SNP arrays across multiple cohorts [10, 21, 22] and applied a series of quality control (QC) procedures before and after imputation using PLINK [28]. First, SNPs that had a poor genotyping rate (<95%), strand ambiguity (A/T and C/G SNPs), a minor allele frequency (MAF) less than 1% or showed deviation from Hardy-Weinberg Equilibrium ($P < 10^{-6}$) were removed. Then, individuals with low genotype rates (<95%), who had sex inconsistencies (between the documented and genotype-derived sex), and who were genetically related were excluded.

Imputation

The genotype data passing QC were imputed on the Michigan server [24, 29] (<https://imputationserver.sph.umich.edu>) separately for each genotyping platform, using the Haplotype Reference Consortium (HRC) reference panel that consists of the largest available set (64,976 human haplotypes) of broadly European haplotypes at 39,235,157 SNPs [30]. For each cohort, imputation quality procedures were implemented to exclude SNPs of low-frequency (MAF < 10%) and low-quality (imputation quality score R-square < 0.6). From the imputed dosage score, genotype calls for the filtered SNPs were derived and common sets of 4,652,947 SNPs across the cohorts were merged using PLINK [28].

Statistical analysis

We implemented polygenic score modeling, genome-wide SNP association, gene-based and functional analysis as described below.

Genome-wide SNP association analysis. Genome-wide SNP association analyses were performed on the binary lithium treatment response and continuous ALDA total score using logistic and linear regression models as implemented in PLINK software [28], respectively. Each analysis was adjusted for the covariates: age, sex, chip type and the first four genetic principal components (PCs). After careful examination of the Multi-dimensional (MD) plot, we observed that the first four PCs successfully captured and delineated any underlying population structure that could potentially influence the genetic association analyses. Consequently, these four PCs were incorporated as covariates in all association analyses. This approach aligns with the methodology employed by previous researchers who utilized the same dataset [10].

Polygenic score development. Using a polygenic score model constructed via Bayesian regression framework and continuous shrinkage (CS) prior on SNP effect sizes implemented in the PRS-CS software [31], we built Li⁺_{PGS} for individuals of European descent who participated in the ConLi⁺Gen study and replicated the findings in the combined PsyCourse and BipoLife datasets. Polygenic scores were computed using PRS-CS to infer posterior SNP effect sizes under continuous shrinkage (CS) using GWAS summary statistics and an external linkage disequilibrium (LD) reference panel. For the current analysis, the precomputed LD pattern of the 1000 Genomes European reference panel [32] and the discovery GWAS summary statistics were used to calculate PGS scores.

For the ConLi⁺Gen study, Li⁺_{PGS} was derived only for the European ancestry individuals ($n = 2367$) using a five-fold leave-one-group out (LOG) procedure [33] to remove discovery-target circularity. In each fold, 80% of the sample ($n = 1893$) was used to generate GWAS summary statistics that were used as discovery for PGS calculation in the 20% left-out target sample ($n = 474$). The procedure was repeated five times by selecting a non-overlapping set of 20% left-out samples to calculate PGS for the entire cohort. Finally, Li⁺_{PGS} was computed for the PsyCourse and BipoLife participants using ConLi⁺Gen's GWAS summary statistics (discovery sample) generated from the full European cohort ($n = 2367$).

Polygenic score association analysis. To assess the association of Li⁺_{PGS} with lithium treatment response, a binary logistic regression model was applied for the binary outcome (good versus poor response to lithium treatment), and a Tobit analysis model (censored regression) was used for the continuous outcome (ALDA total) [34]. In addition, we divided the ConLi⁺Gen sample into deciles, ranging from the lowest polygenic load (1st decile, reference group) to the highest polygenic load (10th decile). Then, we compared BD patients in the higher polygenic load deciles (2nd–10th deciles) with patients in the lowest polygenic load decile (1st decile). In both the binary and continuous outcomes, the proportion of phenotypic variance explained by Li⁺_{PGS} was computed as the difference in R² of the model fit with Li⁺_{PGS} plus covariates, compared to the model fit with only covariates. Each modeling analysis was adjusted for the covariates: age, sex, and the first four genetic PCs, and statistical significance was set at $p < 0.05$.

Gene-based and functional analysis. The gene-based analysis was based on summary statistics generated through genome-wide SNP association analysis of the full European ConLi⁺Gen sample ($n = 2367$) and employed MAGMA (Multi-marker Analysis of GenoMic Annotation) [35], a tool that uses a multiple regression approach to incorporate LD between markers and to detect multi-marker effects.

To explore the biological context of the genes discovered from the gene-based analysis, a pathway analysis was implemented using PANTHER (Protein ANalysis THrough Evolutionary Relationships; <http://pantherdb.org/>) classification system. PANTHER is designed to classify proteins (and their genes) into biological pathways [36]. To prepare the input genes for PANTHER, we selected genes that showed gene-level association with lithium treatment response (either with the categorical or continuous outcome) at MAGMA adjusted p -value < 0.001. This list of genes was entered into PANTHER version-17 which compares the proportion of input genes mapping to a biological pathway to the reference gene list from its databases. Molecular relationships previously experimentally observed in *Homo sapiens* (human) were included. The significance of the overrepresented PANTHER pathways was determined using Fisher's exact test and later adjusted for multiple testing using the

Bonferroni correction method. Significant associations were defined at p -value < 0.05.

RESULTS

Sample characteristics

The discovery analysis consisted of ConLi⁺Gen data obtained from 2,367 bipolar patients of European ancestry who had undergone lithium treatment for at least six months. The mean (sd) age of the patients was 47.5(13.9) years and 1,369 (57.8%) were female. In all, 660 (27.9%) of patients had a good response to lithium treatment (ALDA score ≥ 7). The mean (sd) ALDA score for ConLi⁺Gen participants was 4.1 (3.1). Among 2362 patients who underwent assessment for the type of bipolar diagnosis, the majority (80.0%) were diagnosed with type I bipolar disorder. These patients also presented with comorbid conditions such as psychosis, alcohol dependence, panic disorder, and obsessive-compulsive disorder. Of the 438 patients assessed for possible side effects related to lithium treatment, 153(34.9%) of them reported experiencing at least one of the following: nausea, vertigo, polyuria, diarrhea, hypothyroidism, loss of libido, EEG abnormalities, increased thirst, dermal problems, weight gain, and strangury. The replication analysis was based on a combination of the PsyCourse and BipoLife datasets ($N = 191$), whose mean (sd) age was 49.1(13.0) years. Of the 191 patients with BD, 48(25.1%) had a good response to lithium. This replication cohort exhibits similar characteristics to the discovery sample in terms of the type of bipolar disorder, comorbidities, and patients' reports of lithium treatment side effects (Table 1).

Associations of Li⁺PGS with lithium treatment response in bipolar patients

Using ConLi⁺Gen data, we found statistically significant associations between Li⁺PGS and lithium treatment response — both in the categorical ($P = 9.8 \times 10^{-12}$, $R^2 = 1.9\%$) and continuous ($P = 6.4 \times 10^{-9}$, $R^2 = 2.6\%$) outcomes. Li⁺PGS was positively associated with response to lithium treatment, with an adjusted odds ratio (OR) [95%CI] of **1.39** [1.26, 1.54]. In other words, BD patients who carry a higher genetic loading for lithium responsive genetic variants, measured using the Li⁺PGS, have higher odds of favorable lithium treatment response, compared to patients carrying a low Li⁺PGS load. Table 2 shows the association results of Li⁺PGS and lithium treatment response in categorical and continuous outcomes. The odds of a favorable treatment response increased as the Li⁺PGS increased, ranging from 1.59 fold [95%CI: 1.02–2.49] at the 2nd decile to 3.47 fold [95%CI: 2.22–5.47] at 10th decile, compared to the reference Li⁺PGS at the 1st decile (Table 2). While there was an increasing trend in the odds of lithium treatment response across the deciles, the most significant prediction contrast was found at the 'extremes' (1st and 10th decile) which comprised of ~20% of the total cohort (Fig. 2). A replication PGS analysis in the combined PsyCourse and BipoLife samples found a statistically significant association of Li⁺PGS with the categorical lithium treatment response ($P = 3.9 \times 10^{-4}$, $R^2 = 0.9\%$), but not with the continuous outcome ($P = 0.13$).

Genome-wide association, gene-based and functional analysis

After re-imputing the ConLi⁺Gen data in reference to the latest HRC genomes, we conducted GWASs on lithium response, both in categorical and continuous outcomes. This GWAS analysis identified a single locus with lead SNP rs9396756 located near the stathmin domain containing 1 (*STMND1*) gene that reached genome-wide significance for association with the categorical outcome ($P = 2.7 \times 10^{-8}$) and showed a suggestive association with the continuous ALDA score ($P = 7.6 \times 10^{-8}$) (Fig. 3). A follow-up gene-based analysis of the newly derived ConLi⁺Gen GWAS summary statistics found 36 candidate genes likely associated with lithium treatment response — assessed in either continuous or categorical outcomes ($P < 0.001$). In silico functional analysis of the 36 genes

Table 1. The characteristics of patients with BD and lithium treatment outcomes.

| Characteristics BD patients | ConLi ⁺ Gen | PsyCourse and BipoLife combined |
|---|-------------------------------------|-------------------------------------|
| $N = 2558$ | $N = 2,367$ | $N = 191$ |
| Good responders to lithium defined as ALDA total score ≥ 7 , N (%) | 660 (27.9%) | 48 (25.1%) |
| Mean (se) total ALDA score | 4.12 (3.15) | 4.3 (2.9) |
| Country of origin | N (%) | N (%) |
| Australia | 122 (5.2) | |
| Austria | 43 (1.8) | |
| Canada | 353 (14.9) | |
| Czech Republic | 45 (1.9) | |
| France | 210 (8.9) | |
| Germany | 218 (9.2) | 191 (100%) |
| Italy | 255 (10.8) | |
| Poland | 97 (4.1) | |
| Romania | 152 (6.4) | |
| Spain | 74 (3.1) | |
| Sweden | 304 (12.8) | |
| Switzerland | 57 (2.4) | |
| USA | 437 (18.5) | |
| Age at interview, mean (sd) | 47.5 (13.9) | 49.1 (13.0) |
| Sex, Female, N (%) | 1369 (57.8) | 84 (44.0%) |
| Type of bipolar diagnosis, N (%) | 2362 (99.8) | 89 (46.6) |
| Bipolar type I | 1890(80.0) | 75(84.3) |
| Bipolar type II | 440(18.6) | 14(15.7) |
| Bipolar type III | 7(0.3) | |
| Bipolar not specified | 7(0.3) | |
| Schizoaffective bipolar disorder | 18(0.8) | |
| Comorbidity | $N^{\#}$ (% with) | $N^{\#}$ (% with) |
| Psychosis | 2096 (53.2) | 103 (3.9) |
| Alcohol dependence | 933 (18.0) | 102 (5.9) |
| Panic disorder | 926 (13.6) | 102 (8.8) |
| Obsessive-compulsive disorder | 923 (5.2) | 103 (2.9) |
| Suicidal ideation | - | 98 (66.3) |
| Lithium side effects | 438 (34.9) | 102 (83.3) |

BD Bipolar disorder, N Number of individuals in each group, sd Standard deviation, se Standard error.

$N^{\#}$ refers to the number of individuals assessed for comorbidities, suicidal ideation or lithium side effects.

revealed enriched biological pathways including the muscarinic acetylcholine receptors 1 and 3 (P -value corrected for multiple testing = 0.026) and metabotropic glutamate receptor group III pathway ($P = 0.043$). These genes and pathways may have an impact on clinical response to lithium treatment and be potential molecular targets for lithium (Supplementary Figure 1 and Table 1).

DISCUSSION

This study presents findings from a comprehensive analysis of genetic and clinical data on lithium treatment response that involved the development of a polygenic score for lithium

Table 2. The association of PGS for lithium variants and treatment response to lithium in patients with BD at different sample splits.

| Sample split | N | Categorical outcome, OR (95%CI) | | Continuous outcome: ALDA total score, OR (95%CI) | |
|--|------------------|---------------------------------|-------------------------------|--|-----------------------|
| | | unadjusted | adjusted | unadjusted | adjusted [¥] |
| ConLi⁺Gen | 2367 | | | | |
| 80%/20% | 2083/284 | 1.31(1.19,1.43) | 1.39(1.26, 1.54) [¥] | 1.15(1.11, 1.20) | 1.17(1.13, 1.22) |
| Li ⁺ _{PGS} by decile | [§] R/N | | | | |
| First (lowest score) | 44/236 | 1[Reference] | 1[Reference] [¥] | 1[Reference] | 1[Reference] |
| Second | 60/237 | 1.48(0.96, 2.30) | 1.59(1.02, 2.49) | 0.94(0.79,1.12) | 0.96(0.81,1.15) |
| Third | 54/237 | 1.29(0.82, 2.02) | 1.32(0.84, 2.08) | 1.07(0.90,1.28) | 1.14(0.95,1.35) |
| Fourth | 70/237 | 1.83(1.19, 2.83) | 1.87(1.21, 2.91) | 1.09(0.92,1.31) | 1.14(0.96,1.36) |
| Fifth | 59/236 | 1.45(0.94, 2.27) | 1.50(0.96, 2.35) | 1.12(0.93,1.34) | 1.17(0.98,1.40) |
| Sixth | 62/237 | 1.55(1.00, 2.40) | 1.83(1.17, 2.87) | 1.22(1.02,1.46) | 1.31(1.09,1.55) |
| Seventh | 76/237 | 2.06(1.35, 3.17) | 2.27(1.48, 3.53) | 1.15(0.96,1.38) | 1.23(1.04,1.48) |
| Eighth | 68/237 | 1.76(1.14, 2.72) | 1.91(1.23, 2.99) | 1.12(0.93,1.34) | 1.17(0.98,1.39) |
| Nineth | 78/237 | 2.14(1.41, 3.29) | 2.33(1.51, 3.64) | 1.45(1.21,1.72) | 1.55(1.31,1.86) |
| Tenth (highest score) | 89/236 | 2.64(1.74, 4.05) | 3.47(2.22, 5.47) | 1.52(1.27,1.82) | 1.67(1.39,1.99) |

The reference decile (1st decile) is the PGS category with the lowest polygenic load for lithium variants. OR (95%CI) for the continuous outcome: ALDA total score is calculated as the exponent of beta coefficient from the linear regression model.

[§]R/N: number of lithium responders versus total in that decile; [¥] adjusted for age, sex and 4-genetic principal components, OR: odds ratio.

treatment response (Li⁺_{PGS}), genome-wide SNP association and gene-based and functional analyses.

Since the publication of the first GWAS report by the ConLi⁺Gen team [10], two landmark studies that independently showed the negative association of PGSs for SCZ and MD with lithium treatment response have been published [13–15]. The first study found that 10% of bipolar patients with the lowest polygenic load for SCZ were 3.46 times more responsive to lithium compared to 10% of patients with the highest genetic load for SCZ [14, 15]. Similarly, in the second study, 10% of patients who had the lowest genetic loading for MD were 1.54 times more responsive to lithium than 10% of patients with the highest genetic loading for MD [13, 15]. Nevertheless, each of these PGSs accounts for <2% of the total variance to lithium treatment response [13], suggesting the need to explore additional biological traits that can either independently, or in concert with existing PGSs better predict lithium response. Moreover, the previous PGSs for SCZ and MD are difficult to interpret in a pharmacogenomic context, making the development of a specific lithium response PGS necessary, which is assumed to be more likely to be associated with lithium treatment response and perhaps is biologically more related to lithium's pharmacological actions.

In this novel study, we constructed a PGS for lithium response-Li⁺_{PGS}, a biological marker of direct pharmacogenomic relevance, and showed a positive relationship between a high genetic loading for lithium treatment response variants and long-term therapeutic response to lithium in patients with BD. We demonstrated that bipolar patients at the extreme tail end of the distribution have the strongest association, i.e. 10% of patients who carry high genetic loading for lithium responsive variants (10th decile) were 3.47 times more likely to respond to lithium compared to 10% of those with the lowest Li⁺_{PGS} (1st decile). These results indicated that Li⁺_{PGS} has the potential to help stratify bipolar patients according to predicted lithium response.

In a GWAS of lithium treatment response, we identified a locus near the *STMND1* gene, which encodes for proteins known to be involved in neuron projection development, and active in neuron junctions and cytoplasm. Previous analysis that employed the 1000 Genomes Project reference panel for imputation reported a

suggestive association between genetic variants within the *STMND1* gene and lithium treatment response [10].

Using our newly derived ConLi⁺Gen GWASs summary statistics as an input, we then carried out a gene-based analysis where several genetic variations were examined together for their association with lithium treatment response [35]. This approach found 36 potential target genes for lithium treatment that are enriched in the muscarinic acetylcholine receptors (mAChRs) 1 and 3 and the metabotropic glutamate receptor group III signaling pathways — well characterized biological pathways modulated by the most abundant neurotransmitters in the brain (glutamate and acetylcholine).

Acetylcholine is the central regulator of the mAChRs signaling pathways, which are subfamily of G protein-coupled receptor complexes located in the cell membranes of neurons and other cells that regulate fundamental functions of the central and peripheral nervous system including acting as the main end-receptor stimulated by acetylcholine released from postganglionic fibers in the parasympathetic nervous system [37]. The muscarinic antagonist scopolamine has antidepressant activity, while physostigmine, a cholinesterase inhibitor induces depressive symptoms, suggesting muscarinic receptors may play a role, not only in the pathogenesis of mood disorders, but also as therapeutic targets [38]. M1 and 3 receptors are localized in the cortex, hippocampus and substantia nigra and are known to activate protein kinase C (PKC), causing post-synaptic excitation. PKC is thought to be central in the molecular pathogenesis of BD.

Glutamate, the primary excitatory neurotransmitter in the central nervous system (CNS), exerts neuromodulatory actions via the activation of metabotropic glutamate (mGlu), a type of glutamate receptor that modulates synaptic transmission and neuronal excitability throughout the central nervous system [39]. Group III metabotropic glutamate receptors are largely presynaptically localized and downregulate neurotransmitter release from presynaptic terminals directly or indirectly. These receptors have a prominent expression in the brain, especially in the region of the hippocampus, and can lead to the inhibition of the cAMP cascade which is critical for the maintenance of long-term synaptic plasticity [40]. Growing evidence indicates that abnormalities in the glutamatergic system are implicated in the pathogenesis and

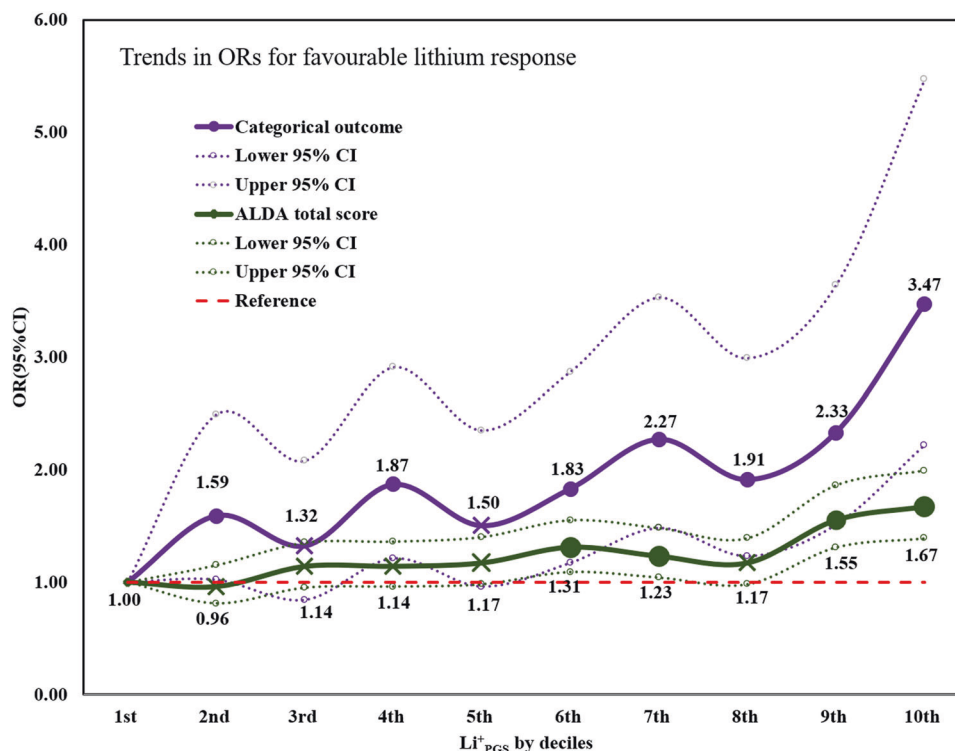


Fig. 2 Trends in the odds ratios (ORs) for favourable treatment response to lithium for patients with bipolar disorder in the higher genetic loading for lithium responsive variants, (2nd to 10th deciles) compared with patients in the lowest (1st decile) of genetic loading for lithium response ($n = 2367$). The X mark on the line plot indicates that the association is not statistically significant at that decile. OR Odds ratio, CI Confidence interval, Li⁺PGS Polygenic score for lithium treatment response.

treatment of mental health disorders [41] including BD [42, 43], SCZ [44], neurodevelopmental disorders [45], Huntington's disease [46] and Alzheimer's disease [47]. Studies have reported SNPs of the mGluRs system associated with BD [48], and in animal studies, lithium was found to alter intracellular calcium by modulating the activity of the metabotropic glutamatergic receptor system [49]. To summarise, findings from the genome-wide SNP association, gene-based and functional analysis highlight the possibility that mechanisms involving glutamate and acetylcholine signaling pathways might influence the therapeutic effects of lithium in patients with BD. Modulation of these pathways through genetic variants may disrupt or enhance lithium's clinical effectiveness.

Our study has some limitations. First, while our findings were replicated in an independent small size sample, the fact that it was replicated in the binary outcome, but not in the continuous outcome indicates the need for a larger replication cohort. Second, because Li⁺PGS was developed and evaluated in European-ancestry populations, the findings should be replicated in a multi-ethnic population to gauge generalizability. Furthermore, the risks and benefits of predictive models consisting of Li⁺PGS should be evaluated in prospective studies. Third, Li⁺PGS only explains about 2% of response variance in our cohort, and as such is comparable to PGSs from other phenotypes (SCZ, MDD) that have shown an association with treatment outcomes. On their own, these PGSs are not suited to clinical pharmacogenomic testing as they would not predict treatment response prospectively in individual patients. Prediction models combining Li⁺PGS with other PGSs [13, 14] and clinical characteristics [19] may improve the clinical utility of PGSs. Such models would then need to be tested in prospective studies and clinical trials. Fourth, studies have shown that approaches to phenotyping of lithium treatment response can be improved using advanced methods such as machine learning [19]. Employing a more precise phenotype definition may result in the identification of novel candidate genes

implicated in lithium treatment response and ultimately the development of more informative Li⁺PGS. Fifth, the current analysis did not include important covariates such as medication dose, information on lithium blood levels, side effects, and the use of concomitant medications (such as Angiotensin-converting enzyme (ACE) inhibitors, diuretics, Non-steroidal anti-inflammatory drugs (NSAIDs)), which can potentially influence lithium clearance and treatment response [50]. Moreover, maintaining therapeutic blood levels is crucial to achieving treatment response with limited side effects in lithium therapy [50]. Lithium possesses a narrow therapeutic index, meaning that there is a relatively small margin between an effective dose and a potentially toxic one. Typically, lithium levels are initially monitored more frequently (weekly or biweekly) during the initiation or adjustment phase of medication, and then less frequently (every 3 to 6 months) once stable therapeutic levels are achieved. While the duration of lithium treatment and the use of certain psychiatric medications (antidepressants, anti-psychotics, mood stabilizers) were assessed as part of the B scale measure of ALDA score, information on the specific dosage, medication blood level and the use of concomitant medications were not available in the ConLi⁺Gen dataset, and thus, they were not considered in our analyses. The inclusion of these pharmacogenomic covariates could provide stronger evidence and should be considered in future research.

In conclusion, we developed a unique lithium treatment response polygenic score (Li⁺PGS) that showed a positive association with better lithium treatment response in patients with BD. Our gene-based and functional analyses build upon the findings from existing molecular studies by linking lithium treatment response with muscarinic acetylcholine receptor signaling and metabotropic glutamate receptor pathways. Further pharmacological evaluation of these pathways in the context of BD and mood stabilizing treatments may prove fruitful.

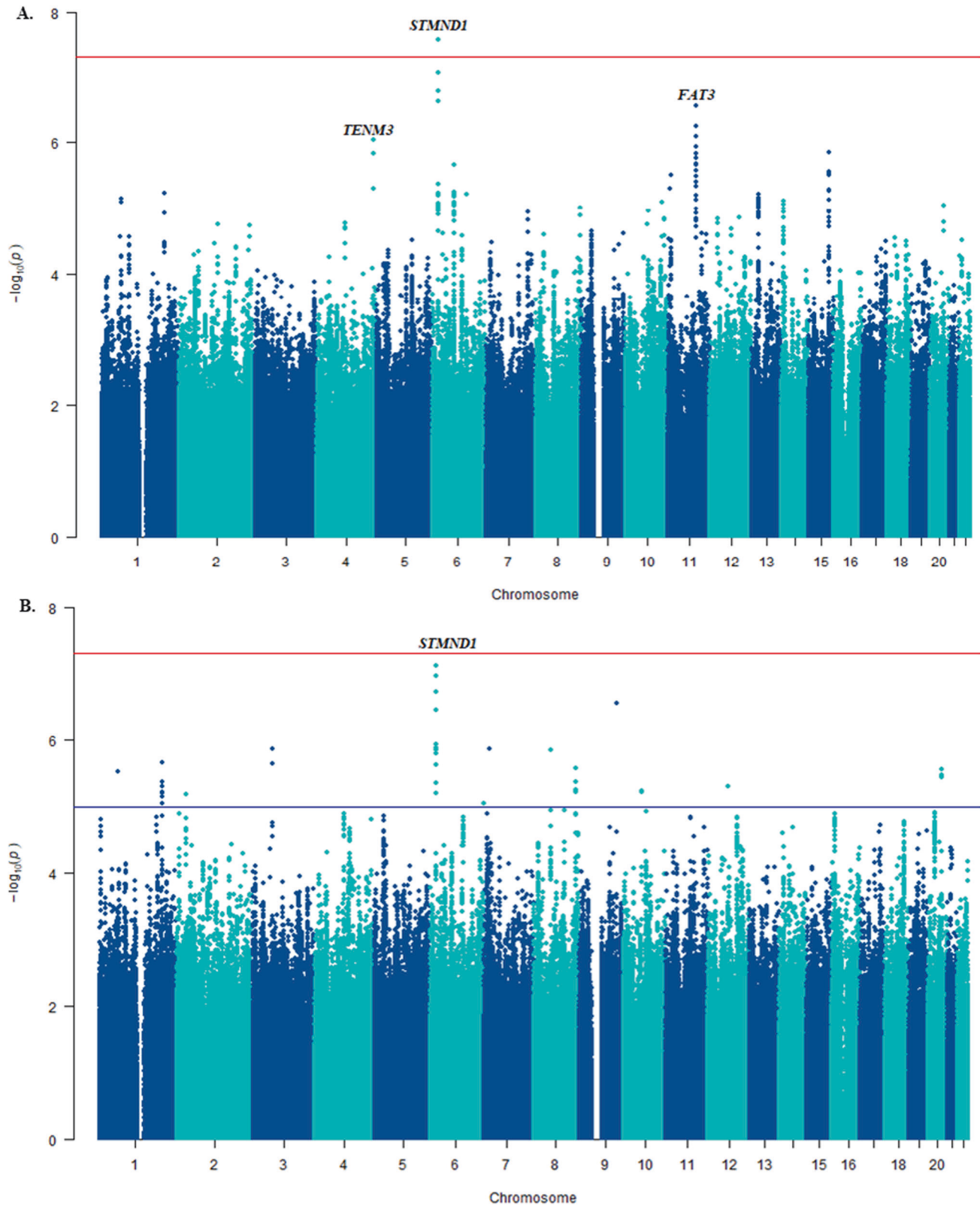


Fig. 3 Manhattan plots showing the SNP-based GWAS results of lithium treatment response in patients with bipolar disorder. **A** In the categorical outcome and **(B)** continuous scale, highlighting the loci that showed genome-wide significance (orange). The $-\log_{10}(p)$ -value is plotted against the physical position of each SNP on each chromosome. The threshold for genome-wide significance (p -value $< 5 \times 10^{-8}$) is indicated by the red dotted horizontal line.

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ACKNOWLEDGEMENTS

The authors are grateful to all patients who participated in the study and we appreciate the contributions of clinicians, scientists, research assistants and study staff who helped in the patient recruitment, data collection and biological sample preparation of the studies. We are also indebted to the members of the ConLi⁺Gen Scientific Advisory Board (<http://www.conligen.org/>) for critical input over the course of the project. The analysis of this study was carried out using the high-performance computational (HPC) capabilities of the University of Adelaide's Phoenix Super-computer <https://www.adelaide.edu.au/phoenix/>.

AUTHOR CONTRIBUTIONS

AT Amare conceived and designed the study hypothesis, as well as secured a fellowship to lead the study. AT Amare and A Thalamuthu conducted the statistical analysis and interpreted the findings. AT Amare, A Thalamuthu, and KO Schubert drafted the manuscript. BT Baune and SR Clark provided supervision for the study. All authors contributed genetic and clinical data, and critically revised the manuscript for important intellectual content.

FUNDING

AT Amare received the 2019–2021 National Alliance for Research on Schizophrenia and Depression (NARSAD) Young Investigator Grant from the Brain & Behaviour Research Foundation (BBRF) and is currently supported by National Health and Medical Research Council (NHMRC) Emerging Leadership (EL1) Investigator Grant (APP2008000). The primary sources of funding were the Deutsche Forschungsgemeinschaft (DFG; grant no. RI 908/7-1; grant FOR2107, RI 908/11-1 to Marcella Rietschel, NO 246/10-1 to Markus M. Nöthen) and the Intramural Research Program of the National Institute of Mental Health (ZIA-MH00284311; ClinicalTrials.gov identifier: NCT00001174). The genotyping was in part funded by the German Federal Ministry of Education and Research (BMBF) through the Integrated Network IntegraMent (Integrated Understanding of Causes and Mechanisms in Mental Disorders), under the auspices of the e:Med Programme (grants awarded to Thomas G. Schulze, Marcella Rietschel, and Markus M. Nöthen). Improving Recognition and Care in Critical Areas of Bipolar Disorders (BipoLife) study was funded by Bundesministerium für Bildung und Forschung (BMBF): Pls – Felix Bempohl, Philipp Ritter, Michael Bauer, Andreas Reif, Sarah Kittel-Schneider, Thomas G. Schulze, Jens Wiltfang, Georg Juckel, Andreas Fallgatter and Martin Lambert. Urs Heilbronner was supported by European Union's Horizon 2020 Research and Innovation Programme (PSY-PGx, grant agreement No 945151). Some data and biomaterials were collected as part of eleven projects (Study 40) that participated in the National Institute of Mental Health (NIMH) Bipolar Disorder Genetics Initiative. From 2003–2007, the Principal Investigators and Co-Investigators were: Indiana University, Indianapolis, IN, R01 MH59545, John Nurnberger, M.D., Ph.D., Marvin J. Miller, M.D., Elizabeth S. Bowman, M.D., N. Leela Rau, M.D., P. Ryan Moe, M.D., Nalini Samavedy, M.D., Rif El-Mallakh, M.D. (at University of Louisville), Hussein Manji, M.D. (at Johnson and Johnson), Debra A. Glitz, M.D. (at Wayne State University), Eric T. Meyer, Ph.D., M.S. (at Oxford University, UK), Carrie Smiley, R.N., Tatiana Foroud, Ph.D., Leah Flury, M.S., Danielle M. Dick, Ph.D. (at Virginia Commonwealth University), Howard Edenberg, Ph.D.; Washington University, St. Louis, MO, R01 MH059534, John Rice, Ph.D., Theodore Reich, M.D., Allison Goate, Ph.D., Laura Bierut, M.D. K02 DA21237; Johns Hopkins University, Baltimore, M.D., R01 MH59533, Melvin McInnis, M.D., J. Raymond DePaulo, Jr., M.D., Dean F. MacKinnon, M.D., Francis M. Mondimore, M.D., James B. Potash, M.D., Peter P. Zandi, Ph.D., Dimitrios Avramopoulos, and Jennifer Payne; University of Pennsylvania, PA, R01 MH59553, Wade Berrettini, M.D., Ph.D.; University of California at San Francisco, CA, R01 MH60068, William Byerley, M.D., and Sophia Vinogradov, M.D.; University of Iowa, IA, R01 MH059548, William Coryell, M.D., and Raymond Crowe, M.D.; University of Chicago, IL, R01 MH59535, Elliot Gershon, M.D., Judith Badner, Ph.D., Francis McMahon, M.D., Chunyu Liu, Ph.D., Alan Sanders, M.D., Maria Caserta, Steven Dinwiddie, M.D., Tu Nguyen, Donna Harakal; University of California at San Diego, CA, R01 MH59567, John Kelsoe, M.D., Rebecca McKinney, B.A.; Rush University, IL, R01 MH059556, William Scheftner, M.D., Howard M. Kravitz, D.O., M.P.H., Diana Marta, B.S., Annette Vaughn-Brown, M.S.N., R.N., and Laurie Bederow, M.A.; NIMH Intramural Research Program, Bethesda, MD, 1Z01MH002810-01, Francis J. McMahon, M.D., Layla Kassem, Psy.D., Sevilla Detera-Wadleigh, Ph.D., Lisa Austin, Ph.D., Dennis L. Murphy, M.D.; Howard University, William B. Lawson, M.D., Ph.D., Evarista Nwulia, M.D., and Maria Hipolito, M.D. This work was supported by the NIH grants P50CA89392 from the National Cancer Institute and 5K02DA021237 from the

National Institute of Drug Abuse. The Canadian part of the study was supported by the Canadian Institutes of Health Research grant (#166098), as well as Genome Canada and Research Nova Scotia grants to MA. Collection and phenotyping of the Australian UNSW sample, by Philip B. Mitchell, Peter R. Schofield, Janice M. Fullerton and Adam Wright, was funded by an Australian NHMRC Program Grant (No.1037196). The collection of the Barcelona sample was supported by the Centro de Investigación en Red de Salud Mental (CIBERSAM), IDIBAPS, and the CERCA Programme / Generalitat de Catalunya (grant numbers PI080247, PI1200906, PI12/00018, 2014SGR1636, and 2014SGR398). The Swedish Research Council, the Stockholm County Council, Karolinska Institutet and the Söderström-Königskga Foundation supported this research through grants awarded to Lena Backlund, Louise Frise'n, Catharina Lavebratt and Martin Schalling. The collection of the Geneva sample was supported by the Swiss National Foundation (grants Synapsy 51NF40-158776 and 32003B-125469). The collection of the Romanian sample was supported by U.E.F.I.S.C.D.I., Romania, grant awarded to Maria Grigoriou-Serbanescu. Open Access funding enabled and organized by CAUL and its Member Institutions.

COMPETING INTERESTS

Eduard Vieta has received grants and served as consultant, advisor or CME speaker for the following entities: AB-Biotics, Allergan, Angelini, AstraZeneca, Bristol-Myers Squibb, Dainippon Sumitomo Pharma, Farindustria, Ferrer, Forest Research Institute, Gedeon Richter, GlaxoSmith-Kline, Janssen, Lundbeck, Otsuka, Pfizer, Roche, Sanofi-Aventis, Servier, Shire, Sunovion, Takeda, the Brain and Behaviour Foundation, the Spanish Ministry of Science and Innovation (CIBERSAM), and the Stanley Medical Research Institute. Michael Bauer has received grants from the Deutsche Forschungsgemeinschaft (DFG), and Bundesministerium für Bildung und Forschung (BMBF), and served as consultant, advisor or CME speaker for the following entities: Allergan, Aristo, Janssen, Lilly, Lundbeck, neuraxpharm, Otsuka, Sandoz, Servier and Sunovion outside the submitted work. Sarah Kittel-Schneider has received grants and served as consultant, advisor or speaker for the following entities: Medice Arzneimittel Pütter GmbH and Shire/Takeda. Bernhard Baune has received grants and served as consultant, advisor or CME speaker for the following entities: AstraZeneca, Bristol-Myers Squibb, Janssen, Lundbeck, Otsuka, Servier, the National Health and Medical Research Council, the Fay Fuller Foundation, the James and Diana Ramsay Foundation. Scott Clark has received grants and served as consultant, advisor or CME speaker for the following entities: Otsuka Australia, Lundbeck Australia, Janssen-Cilag Australia, Servier Australia. Tadafumi Kato received honoraria for lectures, manuscripts, and/or consultancy, from Kyowa Hakko Kirin Co, Ltd, Eli Lilly Japan K.K., Otsuka Pharmaceutical Co, Ltd, GlaxoSmithKline K.K., Taisho Toyama Pharmaceutical Co, Ltd, Dainippon Sumitomo Pharma Co, Ltd, Meiji Seika Pharma Co, Ltd, Pfizer Japan Inc., Mochida Pharmaceutical Co, Ltd, Shionogi & Co, Ltd, Janssen Pharmaceutical K.K., Janssen Asia Pacific, Yoshitomiya Kuhn, Astellas Pharma Inc, Wako Pure Chemical Industries, Ltd, Wiley Publishing Japan, Nippon Boehringer Ingelheim Co Ltd, Kanae Foundation for the Promotion of Medical Science, MSD K.K., Kyowa Pharmaceutical Industry Co, Ltd and Takeda Pharmaceutical Co, Ltd. Tadafumi Kato also received a research grant from Takeda Pharmaceutical Co, Ltd. Peter Falkai has received grants and served as consultant, advisor or CME speaker for the following entities Abbott, GlaxoSmithKline, Janssen, Essex, Lundbeck, Otsuka, Gedeon Richter, Servier and Takeda as well as the German Ministry of Science and the German Ministry of Health. Eva Reininghaus has received grants and served as consultant, advisor or CME speaker for the following entities: Janssen and Institut Allergosan. Mikael Landén declares that, over the past 36 months, he has received lecture honoraria from Lundbeck and served as a scientific consultant for EPID Research Oy; no other equity ownership, profit-sharing agreements, royalties or patent. Kazufumi Akiyama has received consulting honoraria from Taisho Toyama Pharmaceutical Co, Ltd. In 2021, Jörg Zimmermann served as an advisor for Biogen concerning Aducanumab (Alzheimer's Disease). The other authors have no other conflict of interest to disclose.

ADDITIONAL INFORMATION

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41380-023-02149-1>.

Correspondence and requests for materials should be addressed to Azmeraw T. Amare.

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Azmeraw T. Amare¹✉, Anbupalam Thalamuthu², Klaus Oliver Schubert^{1,3}, Janice M. Fullerton^{4,5}, Muktar Ahmed¹, Simon Hartmann¹, Sergi Papiol^{6,7}, Urs Heilbronner⁶, Franziska Degenhardt^{8,9}, Fasil Tekola-Ayele¹⁰, Liping Hou¹¹, Yi-Hsiang Hsu^{12,13}, Tatyana Shekhtman¹⁴, Mazda Adli¹⁵, Nirmala Akula¹¹, Kazufumi Akiyama¹⁶, Raffaella Ardaù¹⁷, Bárbara Arias¹⁸, Jean-Michel Aubry¹⁹, Roland Hasler¹⁹, Hélène Richard-Lepouriel¹⁹, Nader Perroud¹⁹, Lena Backlund^{20,21}, Abesh Kumar Bhattacharjee¹⁴, Frank Bellivier²², Antonio Benabarre²³, Susanne Bengesser²⁴, Joanna M. Biernacka^{25,26}, Armin Birner²⁴, Cynthia Marie-Claire^{22,27}, Pablo Cervantes²⁸, Hsi-Chung Chen²⁹, Caterina Chillotti¹⁷, Sven Cichon^{30,31}, Cristiana Cruceanu³², Piotr M. Czerski³³, Nina Dalkner²⁴, Maria Del Zompo³⁴, J. Raymond DePaulo³⁵, Bruno Étain²², Stéphane Jamain³⁶, Peter Falkai^{7,37}, Andreas J. Forstner^{8,31}, Louise Frisen^{20,21}, Mark A. Frye²⁶, Sébastien Gard³⁸, Julie S. Garnham³⁹, Fernando S. Goes³⁵, Maria Grigoriou-Serbanescu⁴⁰, Andreas J. Fallgatter⁴¹, Sophia Stegmaier⁴², Thomas Ethofer^{42,43}, Silvia Biere⁴⁴, Kristiyana Petrova⁴⁴, Ceylan Schuster⁴⁴, Kristina Adorjan^{6,7}, Monika Budde⁶, Maria Heilbronner⁶, Janos L. Kalman^{6,7}, Mojtaba Oraki Kohshour^{6,45}, Daniela Reich-Erkelenz⁶, Sabrina K. Schaupp⁶, Eva C. Schulte^{6,7}, Fanny Senner^{6,7}, Thomas Vogl⁶, Ion-George Anghelescu⁴⁶, Volker Arolt⁴⁷, Udo Dannlowski⁴⁷, Detlef Dietrich^{48,49}, Christian Figge⁵⁰, Markus Jäger⁵¹, Fabian U. Lang⁵¹, Georg Juckel⁵², Carsten Konrad⁵³, Jens Reimer^{54,55}, Max Schmauß⁵⁶, Andrea Schmitt^{7,57}, Carsten Spitzer⁵⁸, Martin von Hagen⁵⁹, Jens Wiltfang^{60,61}, Jörg Zimmermann⁶², Till F. M. Andlauer⁶³, Andre Fischer⁶¹, Felix BERPpohl¹⁵, Philipp Ritter⁶⁴, Silke Matura⁴⁴, Anna Gryaznova⁶, Irina Falkenberg⁶⁵, Cüneyt Yildiz⁶⁵, Tilo Kircher⁶⁵, Julia Schmidt⁶⁶, Marius Koch⁶⁶, Kathrin Gade⁶⁰, Sarah Trost⁶⁰, Ida S. Haussleiter⁶², Martin Lambert⁶⁴, Anja C. Rohenkohl⁶⁴, Vivien Kraft⁶⁴, Paul Grof⁶⁷, Ryota Hashimoto⁶⁸, Joanna Hauser⁶³, Stefan Herms^{8,30}, Per Hoffmann^{8,30}, Esther Jiménez²³, Jean-Pierre Kahn⁶⁹, Layla Kassem¹¹, Po-Hsiu Kuo⁷⁰, Tadafumi Kato⁷¹, John Kelsoe¹⁴, Sarah Kittel-Schneider^{72,73}, Ewa Ferensztajn-Rochowiak⁷⁴, Barbara König⁷⁵, Ichiro Kusumi⁷⁶, Gonzalo Laje¹¹, Mikael Landén^{77,78}, Catharina Lavebratt^{20,21}, Marion Leboyer⁷⁹, Susan G. Leckband⁸⁰, Alfonso Tortorella⁸¹, Mirko Manchia^{82,83}, Lina Martinsson⁸⁴, Michael J. McCarthy^{14,85}, Susan McElroy⁸⁶, Francesc Colom^{87,88}, Vincent Millischer^{20,21,89}, Marina Mitjans^{88,90,91}, Francis M. Mondimore³⁵, Palmiero Monteleone^{92,93}, Caroline M. Nievergelt¹⁴, Markus M. Nöthen⁸, Tomas Novák⁹⁴, Claire O'Donovan³⁹, Norio Ozaki⁹⁵, Andrea Pfennig⁶⁴, Claudia Pisanu³⁴, James B. Potash³⁵, Andreas Reif⁷², Eva Reininghaus²⁴, Guy A. Rouleau⁹⁶, Janusz K. Rybakowski⁷⁴, Martin Schalling^{20,21}, Peter R. Schofield^{4,5}, Barbara W. Schweizer³⁵, Giovanni Severino³⁴, Paul D. Shilling¹⁴, Katutaka Shimoda⁹⁷, Christian Simhandl⁹⁸, Claire M. Slaney³⁹, Alessio Squassina³⁴, Thomas Stamm^{15,99}, Pavla Stopkova¹⁰⁴, Mario Maj⁹³, Gustavo Turecki³², Eduard Vieta²³, Julia Veeh⁷², Stephanie H. Witt¹⁰⁰, Adam Wright¹⁰¹, Peter P. Zandi¹⁰², Philip B. Mitchell¹⁰¹, Michael Bauer⁶⁴, Martin Alda^{39,94}, Marcella Rietschel¹⁰⁰, Francis J. McMahon¹¹, Thomas G. Schulze^{6,35,103}, Scott R. Clark¹ and Bernhard T. Baune^{104,105,106}

¹Discipline of Psychiatry, School of Medicine, University of Adelaide, Adelaide, SA, Australia. ²Centre for Healthy Brain Ageing (CHeBA), Discipline of Psychiatry and Mental Health, UNSW Medicine & Health, University of New South Wales, Sydney, Australia. ³Northern Adelaide Local Health Network, Mental Health Services, Adelaide, SA, Australia. ⁴Neuroscience Research Australia, Sydney, NSW, Australia. ⁵School of Medical Sciences, University of New South Wales, Sydney, NSW, Australia. ⁶Institute of Psychiatric Phenomics and Genomics (IPPG), University Hospital, LMU Munich, Munich, Germany. ⁷Department of Psychiatry and Psychotherapy, University Hospital, Ludwig-Maximilians-University Munich, Munich, Germany. ⁸Institute of Human Genetics, University of Bonn, School of Medicine & University Hospital Bonn, Bonn, Germany. ⁹Department of Child and Adolescent Psychiatry, Psychosomatics and Psychotherapy, LVR Klinikum Essen, University of Duisburg-Essen, Rheinische Kliniken, Essen, Germany. ¹⁰Epidemiology Branch, Division of Population Health Research, Division of Intramural Research, Eunice Kennedy Shriver National Institute of Child Health and Human Development, National Institutes of Health, Bethesda, MD, USA. ¹¹Intramural Research Program, National Institute of Mental Health, National Institutes of Health, US Department of Health & Human Services, Bethesda, MD, USA. ¹²HSL Institute for Aging Research, Harvard Medical School, Boston, MA, USA. ¹³Program for Quantitative Genomics, Harvard School of Public Health, Boston, MA, USA. ¹⁴Department of Psychiatry, University of California San Diego, San Diego, CA, USA. ¹⁵Department of Psychiatry and Psychotherapy, Charité - Universitätsmedizin Berlin, Campus Charité Mitte, Berlin, Germany. ¹⁶Department of Biological Psychiatry and Neuroscience, Dokkyo Medical University School of Medicine, Mibu, Tochigi, Japan. ¹⁷Unit of Clinical Pharmacology, Hospital University Agency of Cagliari, Cagliari, Italy. ¹⁸Unitat de Zoologia i Antropologia Biològica (Dpt. Biologia Evolutiva, Ecologia i Ciències Ambientals), Facultat de Biologia and Institut de Biomedicina (IBUB), University of Barcelona, CIBERSAM, Barcelona, Spain. ¹⁹Department of Psychiatry, Mood Disorders Unit, HUG - Geneva University Hospitals, Geneva, Switzerland. ²⁰Department of Molecular Medicine and Surgery, Karolinska Institute, Stockholm, Sweden. ²¹Center for Molecular Medicine, Karolinska University Hospital, Stockholm, Sweden. ²²INSERM UMR-S 1144, Université Paris Cité, Département de Psychiatrie et de Médecine Addictologique, AP-HP, Groupe Hospitalier Saint-Louis-Lariboisière-F.Widal, Paris, France. ²³Bipolar and Depressive Disorders Program, Institute of Neuroscience, Hospital Clinic, University of Barcelona, IDIBAPS, CIBERSAM, Barcelona, Catalonia, Spain. ²⁴Department of Psychiatry and Psychotherapeutic Medicine, Research Unit for bipolar affective disorder, Medical University of Graz, Graz, Austria. ²⁵Department of Quantitative Health Sciences, Mayo Clinic, Rochester, MN, USA. ²⁶Department of Psychiatry and Psychology, Mayo Clinic, Rochester, MN, USA. ²⁷Université Paris Cité, Inserm, Optimisation Thérapeutique en Neuropsychopharmacologie, F-75006 Paris, France. ²⁸The Neuromodulation Unit, McGill University Health Centre, Montreal, Canada. ²⁹Department of Psychiatry & Center of Sleep Disorders, National Taiwan University Hospital, Taipei, Taiwan. ³⁰Department of Biomedicine, University Hospital Basel, Basel, Switzerland. ³¹Institute of Neuroscience and Medicine (INM-1), Research Center Jülich, Jülich, Germany. ³²Douglas Mental Health University Institute, McGill University, Montreal, Canada. ³³Psychiatric Genetic Unit, Poznan University of Medical Sciences, Poznan, Poland. ³⁴Department of Biomedical Sciences, University of Cagliari, Cagliari, Italy. ³⁵Department of Psychiatry and Behavioral Sciences, Johns Hopkins University, Baltimore, MD, USA. ³⁶Inserm U955, Translational Psychiatry laboratory, Fondation FondaMental, Créteil, France. ³⁷Max Planck Institute of Psychiatry, Munich, Germany. ³⁸Pôle de Psychiatrie Générale Universitaire, Hôpital Charles Perrens, Bordeaux, France. ³⁹Department of Psychiatry, Dalhousie University, Halifax, Nova Scotia, Canada. ⁴⁰Biometric Psychiatric Genetics Research Unit, Alexandru Obregia Clinical Psychiatric Hospital, Bucharest, Romania. ⁴¹University Department of Psychiatry and Psychotherapy Tuebingen, University of Tübingen, Tuebingen, Germany. ⁴²Department of General Psychiatry, University of Tuebingen, Tuebingen, Germany. ⁴³Department of Biomedical Resonance, University of Tuebingen, Tuebingen, Germany. ⁴⁴Department of Psychiatry, Psychotherapy and Psychosomatics, University Hospital of Frankfurt, Goethe University, Frankfurt, Germany. ⁴⁵Department of Immunology, Faculty of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran. ⁴⁶Department of Psychiatry and Psychotherapy, Mental Health Institute Berlin, Berlin, Germany. ⁴⁷Institute for Translational Psychiatry, University of Münster, Münster, Germany. ⁴⁸AMEOS Clinical Center Hildesheim, Hildesheim, Germany. ⁴⁹Center for Systems Neuroscience (ZSN), Hannover, Germany. ⁵⁰Karl-Jaspers Clinic, European Medical School Oldenburg-Groningen, Oldenburg 26160, Germany. ⁵¹Department of Psychiatry II, Ulm University, Bezirkskrankenhaus Günzburg, Günzburg, Germany. ⁵²Department of

Psychiatry, Ruhr University Bochum, LWL University Hospital, Bochum, Germany. ⁵³Department of Psychiatry and Psychotherapy, Agaplesion Diakonieklinikum, Rotenburg, Germany. ⁵⁴Department of Psychiatry and Psychotherapy, University Medical Center Hamburg-Eppendorf, Hamburg, Germany. ⁵⁵Department of Psychiatry, Health North Hospital Group, Bremen, Germany. ⁵⁶Department of Psychiatry and Psychotherapy, Bezirkskrankenhaus Augsburg, Augsburg, Germany. ⁵⁷Laboratory of Neuroscience (LIM27), Institute of Psychiatry, University of Sao Paulo, São Paulo, Brazil. ⁵⁸Department of Psychosomatic Medicine and Psychotherapy, University Medical Center Rostock, Rostock, Germany. ⁵⁹Clinic for Psychiatry and Psychotherapy, Clinical Center Werra-Meißner, Eschwege, Germany. ⁶⁰Department of Psychiatry and Psychotherapy, University Medical Center Göttingen, Göttingen, Germany. ⁶¹German Center for Neurodegenerative Diseases (DZNE), Göttingen, Germany. ⁶²Psychiatrieverbund Oldenburger Land gGmbH, Karl-Jaspers-Klinik, Bad Zwischenahn, Germany. ⁶³Department of Neurology, University Hospital rechts der Isar, School of Medicine, Technical University of Munich, Munich, Germany. ⁶⁴Department of Psychiatry and Psychotherapy, University Hospital Carl Gustav Carus, Medical Faculty, Technische Universität Dresden, Dresden, Germany. ⁶⁵Department of Psychiatry and Psychotherapy, Philipps-University Marburg, Marburg, Germany. ⁶⁶Institute for Medical Informatics, University Medical Center Göttingen, Göttingen, Germany. ⁶⁷Mood Disorders Center of Ottawa, Ontario, Canada. ⁶⁸Department of Pathology of Mental Diseases, National Institute of Mental Health, National Center of Neurology and Psychiatry, 4-1-1 Ogawahigashi, Kodaira, Tokyo 187-8553, Japan. ⁶⁹Service de Psychiatrie et Psychologie Clinique, Centre Psychothérapique de Nancy - Université de Lorraine, Nancy, France. ⁷⁰Department of Public Health & Institute of Epidemiology and Preventive Medicine, College of Public Health, National Taiwan University, Taipei, Taiwan. ⁷¹Laboratory for Molecular Dynamics of Mental Disorders, RIKEN Brain Science Institute, Saitama, Japan. ⁷²Department of Psychiatry, Psychosomatic Medicine and Psychotherapy, University Hospital Frankfurt, Frankfurt, Germany. ⁷³Department of Psychiatry, Psychotherapy and Psychosomatic Medicine, University Hospital of Würzburg, Würzburg, Germany. ⁷⁴Department of Adult Psychiatry, Poznan University of Medical Sciences, Poznan, Poland. ⁷⁵Department of Psychiatry and Psychotherapeutic Medicine, Landeskrankenhaus Neunkirchen, Neunkirchen, Austria. ⁷⁶Department of Psychiatry, Hokkaido University Graduate School of Medicine, Sapporo, Japan. ⁷⁷Institute of Neuroscience and Physiology, the Sahlgrenska Academy at the Gothenburg University, Gothenburg, Sweden. ⁷⁸Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, Stockholm, Sweden. ⁷⁹Inserm U955, Translational Psychiatry laboratory, Université Paris-Est-Créteil, Department of Psychiatry and Addictology of Mondor University Hospital, AP-HP, Fondation FondaMental, Créteil, France. ⁸⁰Office of Mental Health, VA San Diego Healthcare System, San Diego, CA, USA. ⁸¹Department of Psychiatry, University of Perugia, Perugia, Italy. ⁸²Section of Psychiatry, Department of Medical Sciences and Public Health, University of Cagliari, Cagliari, Italy. ⁸³Department of Pharmacology, Dalhousie University, Halifax, NS, Canada. ⁸⁴Department of Clinical Neurosciences, Karolinska Institutet, Stockholm, Sweden. ⁸⁵Department of Psychiatry, VA San Diego Healthcare System, San Diego, CA, USA. ⁸⁶Department of Psychiatry, Lindner Center of Hope / University of Cincinnati, Mason, OH, USA. ⁸⁷Mental Health Research Group, IMIM-Hospital del Mar, Barcelona, Catalonia, Spain. ⁸⁸Department of Genetics, Microbiology and Statistics, Faculty of Biology, University of Barcelona, Barcelona, Spain. ⁸⁹Department of Psychiatry and Psychotherapy, Medical University of Vienna, Vienna, Austria. ⁹⁰Institut de Biomedicina de la Universitat de Barcelona (IBUB), Barcelona, Spain. ⁹¹Centro de Investigación Biomédica en Salud Mental (CIBERSAM), Instituto de Salud Carlos III, Madrid, Spain. ⁹²Neurosciences Section, Department of Medicine, Surgery and Dentistry "Scuola Medica Salernitana", University of Salerno, Salerno, Italy. ⁹³Department of Psychiatry, University of Campania "Luigi Vanvitelli", Naples, Italy. ⁹⁴National Institute of Mental Health, Klecany, Czech Republic. ⁹⁵Department of Psychiatry & Department of Child and Adolescent Psychiatry, Nagoya University Graduate School of Medicine, Nagoya, Japan. ⁹⁶Montreal Neurological Institute and Hospital, McGill University, Montreal, Canada. ⁹⁷Department of Psychiatry, Dokkyo Medical University School of Medicine, Mibu, Tochigi, Japan. ⁹⁸Bipolar Center Wiener Neustadt, Sigmund Freud University, Medical Faculty, Vienna, Austria. ⁹⁹Department of Clinical Psychiatry and Psychotherapy, Brandenburg Medical School, Brandenburg, Germany. ¹⁰⁰Department of Genetic Epidemiology in Psychiatry, Central Institute of Mental Health, Medical Faculty Mannheim, University of Heidelberg, Mannheim, Germany. ¹⁰¹School of Psychiatry, University of New South Wales, and Black Dog Institute, Sydney, Australia. ¹⁰²Department of Mental Health, Johns Hopkins Bloomberg School of Public Health, Baltimore, MD, USA. ¹⁰³Department of Psychiatry and Behavioral Sciences, SUNY Upstate Medical University, Norton College of Medicine, Syracuse, NY, USA. ¹⁰⁴Department of Psychiatry and Psychotherapy, University of Münster, Münster, Germany. ¹⁰⁵Department of Psychiatry, Melbourne Medical School, University of Melbourne, Parkville, VIC, Australia. ¹⁰⁶The Florey Institute of Neuroscience and Mental Health, The University of Melbourne, Parkville, VIC, Australia.