



Systematic Review

# **Brain Tumor Characterization Using Radiogenomics in Artificial Intelligence Framework**

Biswajit Jena <sup>1</sup>, Sanjay Saxena <sup>1</sup>, Gopal Krishna Nayak <sup>1</sup>, Antonella Balestrieri <sup>2</sup>, Neha Gupta <sup>3</sup>, Narinder N. Khanna <sup>4</sup>, John R. Laird <sup>5</sup>, Manudeep K. Kalra <sup>6</sup>, Mostafa M. Fouda <sup>7</sup>, Luca Saba <sup>2</sup> and Jasjit S. Suri <sup>8</sup>,\*

- Department of CSE, International Institute of Information Technology, Bhubaneswar 751003, India
- Department of Radiology, AOU, University of Cagliari, 09124 Cagliari, Italy
- Department of IT, Bharati Vidyapeeth's College of Engineering, New Delhi 110056, India
- Department of Cardiology, Indraprastha APOLLO Hospitals, New Delhi 110076, India
- <sup>5</sup> Heart and Vascular Institute, Adventist Health St. Helena, St. Helena, CA 94574, USA
- Department of Radiology, Massachusetts General Hospital, 55 Fruit Street, Boston, MA 02114, USA
- Department of Electrical and Computer Engineering, Idaho State University, Pocatello, ID 83209, USA
- <sup>8</sup> Stroke Diagnosis and Monitoring Division, AtheroPoint™, Roseville, CA 95661, USA
- Correspondence: jasjit.suri@atheropoint.com; Tel.: +1-(916)-749-5628

**Simple Summary:** Radiogenomics is a relatively new advancement in the understanding of the biology and behaviour of cancer in response to conventional treatments. One of the most terrible types of cancer, brain cancer, must be targeted in light of the current advancements in therapies. Even though several recent studies on brain cancer have shown promising outcomes when employing the radiogenomics concept, a cutting-edge review of research has been required. In this research review, we provide a 360-degree aspect of brain tumor diagnosis and prognosis employing the new era technology of radiogenomics. The review provides information to the reader about the various aspects that should be considered as accomplishments, opportunities, and limitations in the current therapeutic procedures.

Abstract: Brain tumor characterization (BTC) is the process of knowing the underlying cause of brain tumors and their characteristics through various approaches such as tumor segmentation, classification, detection, and risk analysis. The substantial brain tumor characterization includes the identification of the molecular signature of various useful genomes whose alteration causes the brain tumor. The radiomics approach uses the radiological image for disease characterization by extracting quantitative radiomics features in the artificial intelligence (AI) environment. However, when considering a higher level of disease characteristics such as genetic information and mutation status, the combined study of "radiomics and genomics" has been considered under the umbrella of "radiogenomics". Furthermore, AI in a radiogenomics' environment offers benefits/advantages such as the finalized outcome of personalized treatment and individualized medicine. The proposed study summarizes the brain tumor's characterization in the prospect of an emerging field of research, i.e., radiomics and radiogenomics in an AI environment, with the help of statistical observation and risk-of-bias (RoB) analysis. The PRISMA search approach was used to find 121 relevant studies for the proposed review using IEEE, Google Scholar, PubMed, MDPI, and Scopus. Our findings indicate that both radiomics and radiogenomics have been successfully applied aggressively to several oncology applications with numerous advantages. Furthermore, under the AI paradigm, both the conventional and deep radiomics features have made an impact on the favorable outcomes of the radiogenomics approach of BTC. Furthermore, risk-of-bias (RoB) analysis offers a better understanding of the architectures with stronger benefits of AI by providing the bias involved in them.

**Keywords:** brain tumor; brain tumor characterization; genomics; radiomics; radiogenomics; segmentation; classification; risk-of-bias



Citation: Jena, B.; Saxena, S.; Nayak, G.K.; Balestrieri, A.; Gupta, N.; Khanna, N.N.; Laird, J.R.; Kalra, M.K.; Fouda, M.M.; Saba, L.; et al. Brain Tumor Characterization Using Radiogenomics in Artificial Intelligence Framework. *Cancers* 2022, 14, 4052. https://doi.org/10.3390/cancers14164052

Academic Editor: Sam Payabvash

Received: 15 July 2022 Accepted: 19 August 2022 Published: 22 August 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

Cancers 2022, 14, 4052 2 of 37

#### 1. Introduction

A brain tumor is a fatal disease, and its threat to human life makes it more challenging with the escalation in the number of deaths globally [1]. Brain and other nervous system cancer is a prevalent type affecting adult men, women, and children and is the tenth *primary cause* of death [2]. According to the 2018 Global Cancer Registry results, there were nearly 18 million registered cancer cases in both sexes, of which about 30,000 were related to brain cancer [3]. The brain cancer burden continues to rise globally, producing enormous physical, sentimental, and monetary burdens on individuals, families, communities, and healthcare systems. Countries with mediocre and poor healthcare support systems do not have the exposure to on-time standard diagnosis and treatment for many patients [4].

The traditional diagnosis of brain tumors includes chemotherapy, radiation therapy, biopsy, and surgery based on the tumor size, type, location, and grade [5]. To provide an add-on to these traditional treatment methodologies, medical imaging techniques have been providing phenomenal results for the treatment. The various medical image modalities that have been used for neurological disorders' and brain tumors' detection include magnetic resonance imaging (MRI) [6], computed tomography (CT) [7], cranial ultrasound imaging [8], and positron emission tomography (PET) [9–11], of which MRI is considered to be the most desirable option because of its soft tissue characteristics, and radiation-free nature [12]. However, the radiologist's manual inspection of the medical image to detect tumor features with disease characteristics is tiresome, and the performance varies based on the radiologist's experience.

The advancement of artificial intelligence (AI) technologies, such as machine learning (ML) and deep learning (DL), have shown benefits in disease detection and classification [5,13]. Several ML methods have been adapted in cancer such as brain [14], liver [15], thyroid [16–18], tumor vascularity in breast cancer [19,20], ovarian [21,22], skin [23,24], diabetes [25,26], heart disease [27], and coronary artery disease diagnosis [28–30]. These ML techniques have performed well; however, the feature extraction techniques are ad hoc, causing variabilities in ML results [5].

More recently, deep learning (DL) methods came into existence [31,32] and have shown several medical imaging applications such as in brain cancer [33], carotid wall segmentation [34,35], COVID lesion detection, lung segmentation [36,37], and coronary/carotid plaque classification [38,39]. These DL techniques are certainly better than ML, but they are pretty challenging due to the cost of training time. New techniques such as transfer learning (TL) [40–42] and hybrid deep learning (HDL) [37,43] have improved the training time for the diagnosis process of brain tumor classification, detection, and segmentation with a greater level of automation. These AI technologies have been able to find the tissue-based characteristics of the disease from the medical image using various feature extraction methods and with the subsequent segmentation and classification of disease characteristics [44]. Hence, the amalgamation of AI with radiography takes the brain cancer diagnosis to the next level.

For early detection, quality treatment, and survivorship, the AI paradigm provides classification, detection, and segmentation for brain tumors and has proven successful so far [14,33]. However, the performance of these classification and segmentation methods needs to be further improved for a better diagnosis rate [45]. Detecting genomics information during brain tumor classification and segmentation can further enhance the diagnosis process as the genetic mutation is the prime cause of brain cancer [46]. Due to the exposure of healthy cells to external substances, there is an alteration in the genes present in the cell; thereby, the cells turn to being cancerous cells [47]. Hence, genetic mutation is the prime cause of brain tumors that should be identified during diagnosis. The genomic assessment and status of genetic mutation on various genes and cell proteins, which are the molecular characteristics of the disease, can be detected from the radiological medical imagery using the AI paradigms [48–51].

In the proposed review for brain tumor characterization using radiomics and genomics features, we start with the biology of brain tumors to understand the underlying process of

Cancers 2022. 14, 4052 3 of 37

brain tumors and the mutation process of genomics in the glia brain cells. The essential genetics that have a vital role in brain tumors are discussed with their mutation process and the status of the gene. The classification and grading system of various brain tumors based on genetic mutations have also been tabularized to provide broad information about the brain tumor's characterization [52].

Radiomics and radiogenomics are the two emerging research directions for cancer diagnosis that "finds the disease" and for "genetic" characteristics. Radiomics deals only with the imaging characteristics or phenotype of the disease, while "radiogenomics" links both imaging (phenotype) and genetic (genotype) characteristics of the diseases. The AI technology such as ML, DL, and TL has been helpful in both radiomics and radiogenomics studies in finding out the features of brain tumors accurately and successfully. We thus hypothesize that "AI combined with radiogenomics" can be a powerful paradigm for the diagnosis of brain tumors.

The remaining layout of the article for brain lesion characterization is as follows. Section 2 discusses the PRISMA model search strategy and AI attributes' statistical distribution in radiogenomics studies. Section 3 presents the biology and mutation process of brain cancer. Additionally, classification and grading information of various brain tumors are presented. Section 4 narrates the essential genetics that cause brain cancer and their summary. The radiomics approach for brain lesion characterization using imaging modality is discussed in Section 5. Section 6 displays the radiogenomics method and some critical discussion on various aspects of brain tumor characterization in Section 7. Finally, the study concludes in the last section.

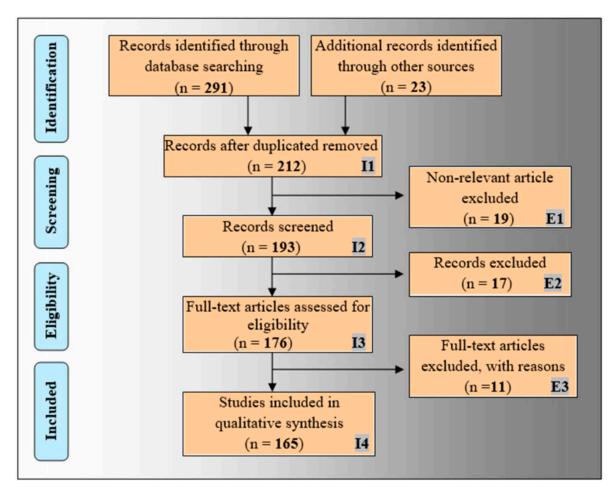
#### 2. PRISMA Model and Search Strategy

This section discusses the radiomics and radiogenomics studies' search approach using the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) model, statistical distribution, and analysis of several important and impacting AI features.

# 2.1. The PRISMA Model

The proposed review included the bias analysis in radiogenomics. The PRISMA prototype has been employed for the selection process of relevant publications as shown in Figure 1. The top academic search databases such as PubMed, IEEE Xplore, Google Scholar, MDPI, and Scopus have been used for this purpose. The admissible keywords used for the search are: "brain tumor", "radiogenomics for brain tumor", "radiomics", "genomics", "radiogenomics", radiogenomics using AI", "deep learning for radiomics", "machine learning for radiomics", "radiogenomics using deep learning", and "radiogenomics using machine learning". A total of 314 records were collected from search databases and other sources. The duplicate records were excluded from this cluster using the "Find Duplicates" feature in the EndNote X9 citation software tool by Clarivate Analytics, which resulted in 212 articles. Again, E1, E2, and E3, as shown in Figure 1, are marked as the three exclusion criteria removing 19, 17, and 11 publications, respectively, under the genre of (i) non-relevant articles, (ii) studies not related to AI, and (iii) articles with insufficient data, resulting in 165 publications. Finally, 20 selected articles [50,53–71] out of these 165 were identified for the bias analysis of radiogenomics studies for risk-of-bias analysis as discussed in Section 7.3.

Cancers 2022, 14, 4052 4 of 37



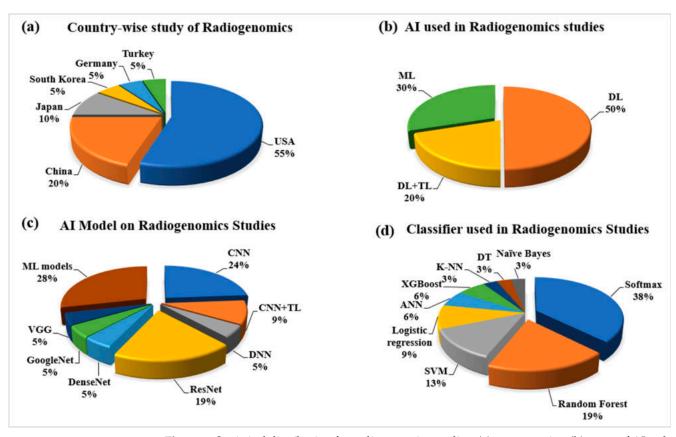
**Figure 1.** The PRISMA framework for the flow diagram of the selection process.

2.2. Statistical Distributions of AI Attributes for Radiogenomics Studies

# 2.2.1. Statistical Distribution of Country-Wise Study of Radiogenomics Using AI

With the worldwide evolution of radiogenomics studies using the cutting-edge technology of AI, it is essential to understand the origin and research of different nations. It has been observed that more than 50% of the research on this new technology is conducted in the USA [53,54,59–61,65,68,71]. The other major contributor is the Republic of China, with 20% of the total studies conducted throughout the review. However, Japan, South Korea, and Turkey have also been actively involved in this radiogenomics study with contributions of 10%, 5%, and 5%, respectively. Such a nation-wide distribution is shown in Figure 2a. It is observed that other parts of the world are not much involved, which shows some biased nature of the radiogenomics study. It should be expected from the remaining part of the globe to be actively involved in this quite innovative research to take it to a different level.

Cancers 2022, 14, 4052 5 of 37



**Figure 2.** Statistical distribution for radiogenomics studies: (a) country-wise; (b) types of AI technology; (c) types of AI models; and (d) AI-based classifiers used in radiogenomics. Notes: ML: machine learning, DL: deep learning, TL: transfer learning, CNN: convolutional neural network, DNN: deep neural network, VGG: visual geometric group, SVM: support vector machine, ANN: artificial neural network, K-NN: K-nearest neighbor, DT: decision tree.

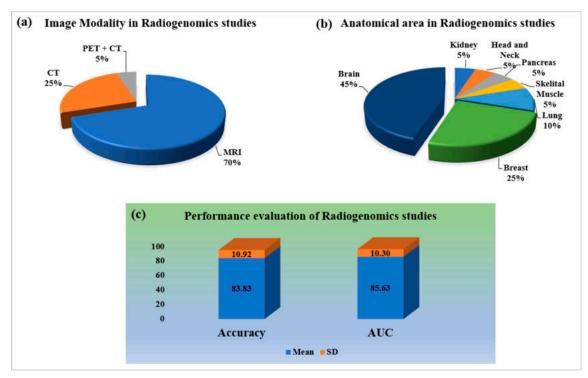
#### 2.2.2. Statistical Distribution of AI Used in the Radiogenomics' System

Since the year of inception in 2002 by Andreassen et al. [72], the study of radiogenomics has been developed in many dimensions. The introduction of AI makes the performance of radiogenomics applications better. The AI technologies such as ML, DL, and transfer learning (TL) have been performing the task of automatic radiomics feature extraction and genomics classification with much ease. Due to its recent success, DL technology has been predominately used in computer vision problems, and it covers 50% of the study considered under this review, as depicted in Figure 2b. Under the DL technology, both phenotype and genotype information of the image can be detected automatically [60,61,66,67]. With the advantage of transfer learning, the small dataset on radiogenomics has been able to perform better. In the case of ML, the hand-crafted radiomics features also provide quite better level performances [63,64]. Figure 2c shows the types of AI models for radiogenomics studies. Among the DL-based model, the convolutional neural network (CNN) and ResNet models are the most popular choices among researchers [60,61,65]. However, other DLbased models used are GoogleNet [71], DenseNet [67], VGG [71], and DNN [53,66]. The ML-based models showed a 28% share. The classifier has always played an essential role in an AI classification model. Figure 2d shows the distribution of different types of classifiers. In radiogenomics study for genotype status prediction, various AI-based classifiers successfully used are the softmax type with 38%, random forest (RF) with 19%, and SVM with 13%. A smaller component is taken by the classifiers such as logistic regression, ANN, XGBoost, K-NN, decision tree (DT), and naïve Bayes, which are all less than 10%.

Cancers 2022, 14, 4052 6 of 37

# 2.2.3. Statistical Distribution by Image Modality and Anatomical Area of Radiogenomics' System

Imaging modality always plays an important part in the medical imaging system. Figure 3a shows the kinds of medical imaging modalities employed. Magnetic resonance imaging (MRI) and computer tomography (CT) are prominent imaging modalities for all kinds of oncology, with 70% and 30% of studies, respectively. Imaging modality directly relates to the anatomical area considered for imaging. Figure 3b shows the distribution of radiogenomics studies for different kinds of organs. Brain images are primarily compatible with MRI; however, CT and PET are also considered. It can be observed from the proposed review that researchers are more oriented toward the brain parts of oncology study with 45% as compared to other anatomical features of the human body. The remaining important anatomical section is breast cancer, with 25% of the studies. The other critical anatomical sections of the body for oncology study are the lung, pancreas, kidney, and skeletal muscle.



**Figure 3.** Statistical distribution for radiogenomics studies: (a) imaging modality; (b) anatomical area; (c) performance evaluation. Notes: CT: computer tomography, PET: positron emission tomography, MRI: magnetic resonance imaging, AUC: area under curve, SD: standard deviation.

# 2.2.4. Statistical Distribution of Performance Evaluation on Radiogenomics System

An AI-based diagnosis system's final and most crucial component is performance evaluation. The better the AI system, the higher the values of the performance evaluation parameters. Accuracy and area under the receiver-operating characteristic curve (AUC) have been chosen as the most appropriate performance evaluation measures in the majority of radiogenomics investigations. However, statistical evaluations such as accuracy, sensitivity, and specificity have been applied to some extent. The mean and standard deviation (SD) of the radiogenomics studies has been found to be 83.83  $\pm$  10.92 and 85  $\pm$  10.30, respectively, as shown in Figure 3c.

# 2.2.5. Statistical Distribution by Dataset Size Covering all the Objectives and Modalities

In this review, the dataset size appraised for the radiogenomics research incorporates the number of patients evaluated for the corresponding research. This covers all of the study's goals and its methodology. The dataset is not easily accessible by the general public and is not volumetric because radiogenomics is a relatively young area of study.

Cancers 2022. 14, 4052 7 of 37

The datasets used in various studies under this review are basically systematic electronic radiological data with open access such as The Cancer Imaging Archives (TCIA) [60,61], The Cancer Genome Atlas (TCGA) [73], The Quantitative Imaging Networks (QIN) [74], and brain tumor segmentation (BraTS) [67], and some have been collected from various hospitals and consortiums [53,63,64,66]. Moreover, we further analyze some other important brain tumor cohorts with a large collection of genotype features. The Repository of Molecular Brain Neoplasia Data, termed *REMBRANDT* [75–77], provides a facility for establishing a connection between genetic data and clinical information. It has data of 874 patients suffering from a glioma containing gene expression arrays. Furthermore, it contains MR scans of 130 REMBRANDT patients. Overall, this dataset offers an opportunity to explore brain tumors in current clinical practice in the era of precision medicine. Moreover, *VASARI* (Visually Accessible REMBRANDT Images) [78,79] was used for defining quantitative features from the MR scans of brain tumor (glioma) patients using the REMBRANDT project. This set of features has 24 observations for neuroradiologists that describes the brain tumor.

Dealing with all kinds of medical research data, including cancer, has been an important issue for both researchers and the healthcare industry. To develop a robust AI system for cancer research, open access to resources such as gene expression and molecular characteristics and radiomics data must be available to the researchers [80,81]. Genomic and molecular information obtained through multiple tissue samples provides detailed spatial information of tumors at different locations [82]. This information could be used for the heterogeneity analysis of tumors. The research band in cancer is narrowing because of the limited availability of open and public databases. To access important cancer research data available under various super specialist cancer hospitals has become even more restrictive. There are several reasons for such restrictions, such as many patients not wanting to participate, and sometimes hospitals show their restrictive behavior, patients leave their follow-ups, etc. To produce an efficient AI model for brain cancer studies, a large cohort with follow-up information is highly recommended. In our opinion, there could be a solution for such restrictions and an increase in the cohort size in the future by collaborating with multiple institutes, in which institutes will have to share the trained AI model (based on certain standard guidelines) instead of sharing the patient's information. Furthermore, based on these trained models, further analysis (test) will be conducted. Moreover, the restrictions imposed will be scarcer due to the incorporation of privacy-preserving methods. Even more controlled-access initiatives have been taken to protect the privacy of research participants [83,84]. It is noticed that all the studies incorporated have datasets within 1000 subjects, and a few studies [50,54,59] have been limited to below 100 subjects, as depicted in Figure 4. To minimize data imbalance and overfitting circumstances, a larger amount of data is anticipated for a more accurate assessment of the radiogenomics' AI system.

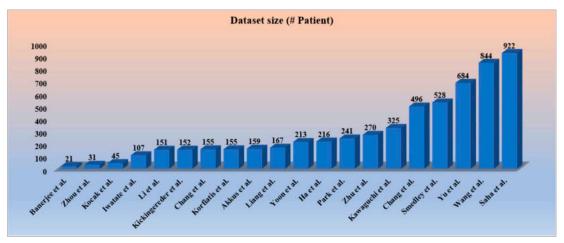


Figure 4. The distribution of increasing dataset size in various radiogenomics studies [53–71].

Cancers 2022, 14, 4052 8 of 37

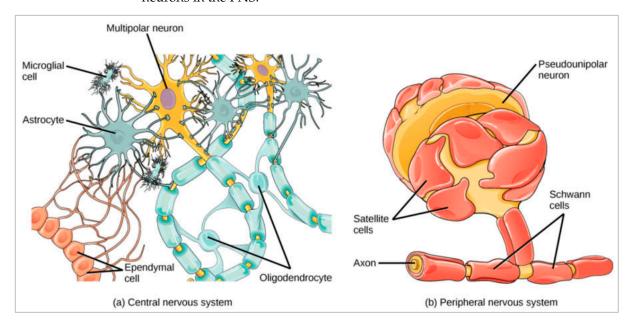
#### 3. Biology of Brain Tumor

The biology or pathophysiology of brain tumors is discussed in this section. This includes the cells, the brain cell (glia and neuron), deoxyribonucleic acid (DNA), mutation, mutation process, and uncontrol growth of the mutated cell.

# 3.1. Brain Glia Cell and Deoxyribonucleic Acid

The cell is the smallest unit of an organism with a nucleus and the molecular fragments of the human anatomy that determine the purpose of each organ within the body. The main job of the cell includes blood flow, oxygen flow, and metabolic management in the human body. However, the most regular brain cells are neurons and non-neuron cells (called *glia* cells) [85]. These neurons have an essential role in the proper functioning of the brain and passing signals to other parts of the body. The glia cells are the supporting cast of the nervous system that fuel the function of the neurons. The neuroglia cells are the non-neural cells in the central nervous system (CNS) and peripheral nervous system (PNS) that do not produce neurological electrical signals [86]. The average adult human brain has about 100 billion neurons, with glia cells accounting for ten times that number.

There are six types of glia cells having different functions, such as (a) astrocytes, (b) oligodendrocytes, (c) microglia, (d) ependymal cells, (e) satellite cells, and (f) Schwann cells. The first four glia cells belong to the CNS and the last two to the PNS, as shown in Figure 5b. Astrocytes are star-shaped cells that keep a neuron's operating environment in a good condition. They provide protection and structural support to neurons, exchanging nutrients and other important chemicals with neurons. Oligodendrocytes help neurons in the central nervous system, especially those traveling vast distances within the brain, by supporting their axons. They make a fatty material called myelin, which forms a protective covering surrounding the axon. Microglia scavenge and destroy dead cells while protecting the brain from microbes. Ependymal cells line the spinal cord and ventricles of the brain. They help make cerebrospinal fluid (CSF), which serves as a brain cushion and transfers fluid between the spinal cord and the brain. It is also a component of the choroid plexus. The Satellite glia [87] supplies nutrition and structural support for neurons in the PNS. Schwann cells of the PNS are similar to oligodendrocytes in the CNS, which myelinate neurons in the PNS.



**Figure 5.** Glia cells of the neuron and their environment of the central nervous system and peripheral nervous system [88].

Cancers 2022, 14, 4052 9 of 37

> The nucleus, which contains 23 pairs of chromosomes and millions of genes, is the core control system of each glia cell. [33]. Deoxyribonucleic acid (DNA), which acts as a blueprint for genes and specifies their function, contains the instructions for these genes.

#### 3.2. Mutation and Its Process

A mutation causes a change in the DNA sequence, which is the fundamental cause of gene malfunction. DNA mutations are influenced by a variety of factors, including the environment, lifestyle, and eating habits. Three types of cancer-causing genes have been identified such as (a) tumor suppressors, which control the cell death cycle (apoptosis), (b) genes that are accountable for the restoration of the DNA, and (c) proto-oncogenes, which work in opposition to tumor suppressor genes and are responsible for the synthesis of a protein that promotes cell proliferation while blocking standard cell death [33].

#### 3.3. Central Nervous System and Brain Tumor Types' Classification and Grading

The knowledge of classification of brain tumor types and grading provides many advantages in treating the various types of tumors. The World Health Organization (WHO), the apex organization of world health, has defined the latest, i.e., the fifth issue of classification of tumor types in 2021 following the initial publications from 1979, 1993, 2000, 2007, and 2016. The new edition of tumor types' classification basically focuses on molecular information with histology that helps more accurate diagnoses, prognoses, and treatment plans, and to predict therapeutic responses for patients. The recent advances relate to classification approaches according to the WHO: more than 120 brain and central nervous system (CNS) tumors exist for both adult and pediatric types. The various types and subtypes of CNS and brain tumor classification systems have been illustrated by Louis et al. [52] in Table A1 in Appendix A. Apart from the classification system of brain tumors, the WHO also provided the grading system of CNS and brain tumors based on the cancer type's severity. Table 1 lists the four grading systems and some selected brain tumor types associated with each grade.

| Grading | Some Selected Types of CNS and Brain Tumor  |  |  |
|---------|---|--|--|
| Grade 1 | Meningioma, solitary fibrous tumor, diffuse astrocytoma (MYB- or MYBL1-altered), polymorphous low-grade neuroepithelial tumor of the young, multi-nodular and vacuolating neuronal tumor.   |  |  |
| Grade 2 | Meningioma, solitary fibrous tumor, oligodendroglioma, astrocytoma (IDH-mutant), IDH-mutant, and 1p/19q-co-deleted, myxopapillary ependymoma, pleomorphic xanthoastrocytoma, supratentorial ependymoma, posterior fossa ependymoma. |  |  |
| Grade 3 | Meningioma, solitary fibrous tumor, oligodendroglioma, Astrocytoma (IDH-mutant), IDH-mutant, and 1p/19q-co-deleted, pleomorphic xanthoastrocytoma, supratentorial ependymoma posterior fossa ependymoma.                            |  |  |
| Grade 4 | Glioblastoma (IDH-wildtype), astrocytoma (IDH-mutant), diffuse hemispheric  |  |  |

Table 1. WHO grading of CNS and brain tumors.

# 4. Genetics of Brain Tumor

#### 4.1. Genomics Types' Information of Brain Tumor

The genes and enzymes of the human body play an essential role in human growth and nourishment. Due to the various external parameters based on alteration, these genomes and enzymes may cause brain cancer [89]. Here, we will discuss prominent genotypes that are accountable for the formation of brain tumors and discuss their impact and severity, such as glioblastoma multiforme.

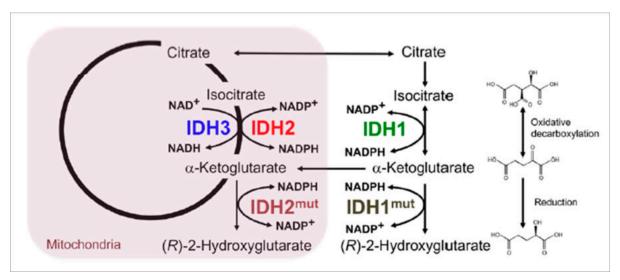
glioma (H3 G34-mutant).

Isocitrate Dehydrogenase (IDH): IDH1 and IDH2 are two families of a gene that generate enzymes called isocitrate dehydrogenase (IDH), which are helpful for the breakdown of nutrients and produce energy for cells [33,89,90]. The enzymes produced from these genes are involved in the amino acid and citric acid formation cycle. The mutation of these

Cancers 2022, 14, 4052 10 of 37

genes is strongly associated with glioma and other types of cancer. Figure 6 shows the chemical reaction and metabolic pathways present in a brain tumor cell, with emphasis on enzymatic effectors—IDH1 and IDH2 mutations. IDH1 gene mutation is found in 5% of patients having a primary glioblastoma; however, 70% to 80% of patients develop a glioma of grade II to III and secondary glioblastoma [90]. IDH2 mutations are primarily seen in oligodendroglia tumors. Another IDH variant is the IDH-wildtype gene, which has a more rigorous form of IDH mutation, with distinctive genetic and clinical characteristics than IDH1 and IDH2. Patients with the IDH-wildtype gene present have 90% chance of a glioblastoma developing and carry a worse prognosis than other IDH mutants.

The systematic review and meta-analysis conducted by Suh et al. [91] for IDH (isocitrate dehydrogenase) mutation prediction in glioma patients with the help of imaging radiomics data summarizes: Compared to IDH-wildtype glioma, IDH-mutant glioma consistently displayed less aggressive imaging characteristics. MRI demonstrated the potential to non-invasively predict IDH mutation in patients with glioma despite the wide range of different MRI techniques used. The pooled sensitivity of 2-hydroxyglutarate MRS is higher than that of other imaging modalities.



**Figure 6.** Chemical reaction and metabolic pathways present in a brain tumor cell, with emphasis on enzymatic effectors—IDH1 and IDH2 mutations [92].

Yan et al. [89] presented the IDH1 and IDH2 mutation in glioma, which is the most frequently occurring brain tumor in the adult brain and central nervous system. The conclusion of the study reveals that in the majority of the various forms of malignant gliomas, mutations of the NADP + dependent isocitrate dehydrogenases encoded by IDH1 and IDH2 are found as shown in Figure 6.

1p and 19q Co-deletion: The mutation of 1p and 19q co-deletion implies the compound loss of both the short arm of chromosome 1 (1p) and the long arm of chromosome 19 (19q) is the molecular genetic signature of oligodendrogliomas, a subtype of primary brain tumors accounting for approximately ten to fifteen percent of all diffuse gliomas in adults [93]. The 1p and 19q co-deletion chromosomes help in the glioma's prognosis, diagnostic, and predictive biomarker. Hence the mutation effect of the gene results in a poor prognosis of the brain tumor [33]. Up to 80% of oligodendrogliomas, 60% of anaplastic oligodendrogliomas, 30–50% of oligoastrocytomas, and 20–30% of anaplastic oligoastrocytomas have the mutation impact of this gene [90].

A systematic review and meta-analysis by Hu et al. [94] for the role of chromosomal 1p/19q co-deletion in connection with personalized treatments for patients suffering from oligodendrogliomas demonstrate that the pooled findings of the study show the prognosis of grade II/III oligodendrogliomas is significantly improved by chromosomal 1p/19q co-deletion.

Cancers 2022, 14, 4052 11 of 37

The research study conducted by Han et al. [95] for the development and validation of an MRI-based radiomics signature for the non-invasive genotype prediction of chromosome 1p/19q co-deletion in lower-grade gliomas summarizes that an MRI-based radiomics signature may successfully detect the 1p/19q co-deletion in lower-grade gliomas (WHO grade II and III gliomas) with histological diagnosis, potentially enabling the non-invasive genetic subtype prediction of gliomas.

O6-methylguanine DNA methyltransferase (MGMT): The MGMT is a DNA rebuilding enzyme that shields the tumor cells from the alkylating agent (induced damage during chemotherapy) by eliminating alkyl groups from the O<sup>6</sup> position of the guanine [82]. The MGMT promoter methylation has been established as an important clinical biomarker in neuro-oncology. The presence of the promoter has a better prognosis and longer survival with glioblastomas who receive alkylating agents. It is useful for identifying newly diagnosed glioblastomas and also helps in a patient's survival prediction [33,90]. MGMT promoter methylation has also been reported to have high rates in oligodendrogliomas and astrocytomas of lower grade, which variably correlate with 1p19q co-deletion and IDH mutations. Methylation of the MGMT promoter is detected in 35–75 percent of glioblastomas. [96].

The review research by Cabrini et al. [97] on the most common genomics biomarkers of brain cancer, i.e., MGMT for the regulation of expression of MGMT and treatment of glioblastoma, reviews various aspects such as the role of MGMT in cancer, nuclear transcription factors that control MGMT gene expression, the effect of histone modifications, the MGMT promoter, regulation of MGMT expression by microRNAs, MGMT as a prognostic biomarker, and, finally, the manipulation of MGMT expression to enhance first-line glioblastoma treatment.

Silber et al. [98] in their review of the promise and problems of O6-methylguanine DNA methyltransferase in glioma therapy, suggest the improvement in the utility of MGMT methylation status in planning optimal therapies tailored to individual patients.

B-Raf proto-oncogene (BRAF): The BRAF is a proto-oncogene that encodes the protein B-Raf of the RAF family of serine/threonine protein kinases. This protein influences cell division, differentiation, and secretion via modulation of the MAP kinase/ERK signaling pathway [99]. This gene's mutations, most notably the V600E mutation, are the most commonly detected cancer-causing mutations in melanoma and have also been found in various other malignancies. [100].

Werner et al. [101] in their review research on isocitrate dehydrogenase and B-Raf proto-oncogene-wildtype epithelioid glioblastoma treatment response viable on MRI radiological imaging features, suggest that this subtype of genomic biomarkers is frequently linked to a poor clinical outcome and overall survival. Moreover, epithelioid glioblastoma without an isocitrate dehydrogenase or B-Raf proto-oncogene mutation was found to be a very aggressive molecular subtype by neuropathology.

The systematic review by González-González et al. [99] on B-Raf proto-oncogene serine and threonine kinase V600E mutation for ameloblastoma-targeted patients' treatments summarizes that when surgical treatments for ameloblastomas are not an option, the discovery of these genomic biomarkers has revolutionized alternative therapies.

Tumor Protein 53 (TP53 or p53): The TP53 gene has a vital role in the formation of a protein called tumor protein (p53) and also in DNA repair [33]. The TP53 also acts as a tumor suppressor, a transcription factor that controls the cell proliferation from an uncontrolled growing and dividing of cells and regulates the cell cycle. The alteration and abnormalities of TP53 lead to genetic instability, reduced apoptosis, and angiogenesis [90]. The degree of mutation is more relevant to high-grade glioma and 80% of the tumor. It is otherwise called the guardian of the genome as it repairs or self-destructs (apoptosis) the DNA when there is damage by external agents such as radiation or ultraviolet (UV) rays from sunlight and toxic chemicals.

Zhang et al. [102] in their review research on "The p53 Pathway in Glioblastoma" summarize the following statements. One of the most often dysregulated genes in cancer

Cancers 2022. 14, 4052 12 of 37

is TP53. In 84 percent of glioblastoma (GBM) patients and 94 percent of GBM cell lines, the p53-ARF-MDM2 pathway is dysregulated. Components of the p53 pathway that are dysregulated have been linked to GBM cell invasion, migration, proliferation, apoptosis evasion, and cancer cell stemness. Numerous microRNAs and long non-coding RNAs are also involved in the regulation of these pathway constituents. The majority of TP53 mutations in GBM are point mutations that increase the expression of the p53 protein's gain of function (GOF) oncogenic variants. These GOF p53 mutants, which have received relatively little research, may cause GBM malignancy by acting as transcription factors on a different set of genes from those controlled by wildtype p53. Their expression is associated with a worse outcome, suggesting that they may be important indicators and treatment targets for GBM.

Retinoblastoma1 (RB1): The RB1 gene helps in the formation of protein in a cell called P<sup>RB</sup>. This RB1-generated protein works as a tumor suppressor that regulates the growth and uncontrolled division of cells. The mutation of this genomics hence blocks cell cycle progression and unchecked cell cycle progression. The tumor suppressor pathway is commonly seen in the case of high-grade gliomas such as glioblastoma multiforme (GBM). Additionally, the RB1-generated protein influences cell survival and cell self-destruction (apoptosis) [33,90].

Goldhoff et al. [103] in their proposed research, focus on the clinical classification of glioblastoma based on retinoblastoma tumor suppressor protein (RB1) mutations and link with the proneural subtype. The result analysis suggests that clinical GBM specimens using IHC to determine RB1 status, and they hypothesize that RB1 changes may be more prevalent in specific GBM subtypes.

Epidermal Growth Factor Receptor (EGFR): The EGFR is a transmembrane protein that is a receptor for members of the epidermal growth factor family such as phospha tase and tensin homolog (PTEN), receptor tyrosine kinase (RTK), and phosphatidylinositol 3-ki nase (PI3K) cell proliferation pathway. The mutation of this protein increases the cell proliferation rate. Upregulation or amplification of the EGFR gene has been linked to various malignancies, including 50 percent of primary glioblastoma cases [33,90].

The review research on "Epidermal Growth Factor Receptor (EGFR) in Glioblastoma" by Xu et al. [104] focuses on various aspects such as EGFR mutation in glioblastoma, EGFR therapies in glioblastoma, and glioblastoma's dysregulated EGFR signaling networks such as PI3K, MAPK.

Phosphatase and Tensin Homolog (PTEN): Phosphatase and tensin homolog (PTEN) is a phosphatase in humans that is encoded by the PTEN gene. Mutations in this gene cause a variety of malignancies, including glioblastoma, lung cancer, breast cancer, and prostate cancer. It is also a tumor suppressor gene. The mutation effect of this gene has increased cell proliferation and reduced cell death in 80% of glioblastomas [33,90].

Karsy et al. [105] in the practical review on prognostic correlations of various molecular biomarkers in glioblastoma including the phosphatase and tensin homolog (PTEN) discuss the clinical trials on these biomarker indicators and their applicability to clinical practice, the distinction between prognostic and predictive biomarkers, and other related information. Furthermore, the survey finds that despite PTEN's significance in the development of gliomas, there is still no evidence linking it to survival.

#### 4.2. Brain Tumor Types and Their Associated Genes

Brain tumors are associated with several familial cancer predisposition syndromes. This portion of the article depicts the key genomes and proteins whose germline mutation forms the different types of brain tumors, as provided in Table 2.

Cancers 2022, 14, 4052 13 of 37

Table 2. Brain tumor types and their underlying genetic and epigenetic alterations [33,52,89,90,106–108].

| SN. | <b>Brain and CNS Tumor Types</b>               | Key Genes and Protein Alterations for Tumor   |
|-----|--|---|
| 1   | Astrocytoma Grade I: Pilocytic Astrocytoma     | BRAF, NF1, KIAA1549-BRAF  |
| 2   | Astrocytoma Grade II: Low-grade Astrocytoma    | EGFR1, BRAF   |
| 3   | Astrocytoma Grade III: Anaplastic Astrocytoma  | IDH1/2, TP53, ATRX, CDKN2A/B  |
| 4   | Astrocytoma Grade IV: Glioblastoma (GBM)       | IDH1/2, TERT, chromosomes 7/10, EGFR  |
| 5   | Oligodendroglioma                              | IDH1/2, TERT promoter, 1p/19q, NOTCH1, FUBP1, CIC   |
| 6   | Angiocentric glioma                            | MYB   |
| 7   | Diffuse astrocytoma                            | MYB, MYBL1  |
| 8   | Medulloblastoma                                | TP53, CTNNB1, PTCH1, APC, SUFU, GLI2, SMO, MYC, MYCN, PRDM6, KDM6A.   |
| 9   | Meningiomas                                    | NF2, TRAF7, AKT1, PIK3CA; SMO, SMARCE1, KLF4, BAP1 in subtypes; H3K27me3; TERT, CDKN2A/B in CNS WHO grade 3 |
| 10  | Retinoblastoma                                 | Retinoblastoma (Rb) protein   |
| 11  | Ependymomas                                    | ZFTA, YAP1, RELA, MAML2, H3 K27me3, NF1, NF2, EZHIP, MYCN,<br>KMT2D, RELA, FANCE, and EP300                 |
| 12  | Primitive neuroectodermal tumors               | Isochrome (17q)   |
| 13  | Astroblastoma                                  | MN1   |
| 14  | Chordoid glioma                                | PRKCA   |
| 15  | Ganglion cell tumors                           | BRAF  |
| 16  | Polymorphous low-grade neuroepithelial tumor   | BRAF, FGFR family   |
| 17  | Diffuse midline glioma, H3 K27-altered         | TP53, H3 K27, PDGFRA, EGFR, ACVR1, EZHIP  |
| 18  | Diffuse hemispheric glioma, H3 G34-mutant      | TP53, H3 G34, ATRX  |
| 19  | Diffuse pediatric-type high-grade glioma       | IDH-wildtype, H3-wildtype, MYCN, PDGFRA, EGFR   |
| 20  | Infant-type hemispheric glioma                 | NTRK family, ROS, ALK, MET  |
| 21  | High-grade astrocytoma with piloid features    | ATRX, BRAF, CDKN2A/B (methylome), NF1   |
| 22  | Pleomorphic xanthoastrocytoma                  | CDKN2A/B, BRAF  |
| 23  | Subependymal giant cell astrocytoma            | TSC1, TSC2  |
| 24  | Solitary fibrous tumor                         | NAB2-STAT6  |
| 25  | Meningeal melanocytic tumors                   | NRAS (diffuse), GNA11, GNAQ, CYSLTR2, PLCB4   |
| 26  | Atypical teratoid/rhabdoid tumor               | SMARCA4, SMARCB1  |
| 27  | Embryonal tumor with multi-layered rosettes    | C19MC, DICER1   |
| 28  | Glioneuronal tumor                             | NF1, PDFGRA, PRKCA, FGFR1, PIK3CA, KIAA1549-BRAF fusion,<br>1p, Chromosome 14                               |
| 29  | Dysplastic cerebellar gangliocytoma            | PTEN  |
| 30  | Extraventricular neurocytoma                   | IDH-wildtype, FGFR (FGFR1-TACC1 fusion)   |
| 31  | Multi-nodular and vacuolating neuronal tumor   | MAPK pathway  |
| 32  | Dysembryoplastic neuroepithelial tumor         | FGFR1   |
| 33  | CNS neuroblastoma                              | FOXR2, BCOR   |
| 34  | Desmoplastic myxoid tumor of the pineal region | SMARCB1   |

BRAF: proto-oncogene B-Raf, NF1: neurofibromin 1, FGER: fibroblast growth factor receptors, EGFR: epidermal growth factor receptor, IDH: isocitrate dehydrogenase, ATRX: alpha thalassemia X-linked mental retardation, TP53: tumor protein53, CDKN2A/B: cyclin-dependent kinase inhibitor 2A/B, TERT: telomerase reverse transcriptase, CIC: capicua transcriptional repressor, FUBP1: far upstream element binding protein 1, NOTCH1: notch homolog 1, MYB: myeloblastosis, MYBL1: MYB like1, CTNNB1: catenin beta-1, APC: adenomatous polyposis coli, PTCH1: protein patched homolog 1, SUFU: suppressor of fused protein, SMO: smoothened, AKT1: threonine kinase 1, TRAF7: TNF receptor associated factor 7, ZFTA: zinc finger translocation associated, YAP1: yes-associated protein 1, MAML2: mastermind-like transcriptional coactivator 2, H3: histone3, EZHIP: EZH inhibitory protein, KMT2D: lysine methyltransferase 2D, FANCE: FA complementation group E, MN1: meningioma, PRKCA: protein kinase C alpha, ACVR1: activin A receptor type 1, PDGFRA: platelet-derived growth factor receptor alpha, NTRK: neurotrophic tyrosine receptor kinase, ALK: anaplastic lymphoma kinase, MET: mesenchymal epithelial transition, TSC: tuberous sclerosis, STAT6: signal transducer and activator of transcription 6, NRAS: neuroblastoma RAS viral oncogene homolog, GNAQ: G protein subunit alpha Q, GNA11: G protein subunit alpha 11, PLCB4: phospholipase C beta 4, CYSLTR2: cysteinyl leukotriene receptor 2, SMARCB: SWI/SNF related, matrix associated, actin dependent regulator Of chromatin, subfamily B C19MC: chromosome 19 miRNA cluster, DICER1: dicer 1, ribonuclease III, PRKCA: protein kinase C alpha, PTEN: phosphatase tensin homologue, MAPK: mitogen-activated protein kinase, FOXR2: forkhead Box R2, BCOR: BCL6 corepressor.

Cancers 2022. 14, 4052 14 of 37

#### 5. A Radiomics Approach to Tumor Characterization

This section is based on radiological imaging or radiomics to characterize the brain tumor with segmentation and classification as part of the radiomics process. To proceed into the central paradigm of brain tumor characterization, we enlighten the understanding of the pre-requisite concepts such as imaging modalities and the functionality of the modality.

Magnetic resonance imaging (MRI) is a non-invasive, highly sophisticated, highly detailed, three-dimensional imaging diagnostic method of the interior structures of the human body [14,109]. In neurology and neurosurgery, MRI is one of the most often used tests. MRI has an advantage over CT in detecting flowing blood and cryptic vascular malformations. It can also detect demyelinating disease and has no beam-hardening artifacts such as the damaging ionizing radiation of x-rays that can be seen with CT. Due to the non-use of ionizing radiation in X-rays, this MRI imaging modality is used when there is a need for frequent imaging for diagnosis and therapy of the brain. The brain, spinal cord, and nerves are more easily visualized on MRI than on CT [110]. The MRI is the preferred imaging diagnostic method for head and neck injuries, cancers, and hemorrhages. Table 3 provides the comparison of prominent imaging modalities based on various factors.

|             | 0 0              |                  |                  |             |
|-------------|------------------|------------------|------------------|-------------|
| Factor      | MRI              | CT               | X-Ray            | Ultrasound  |
| Duration    | 30–45 min        | 3–7 min          | 2–3 min          | 5–10 min    |
| Cost        | High             | Moderate         | Low              | Low         |
| Soft tissue | Excellent detail | Poor detail      | Poor detail      | Poor detail |
| Bone        | Poor detail      | Excellent detail | Excellent detail | Poor detail |
| Dimension   | 3                | 3                | 2                | 2           |
| Radiation   | No               | 10 mSv           | 0.15 mSv         | No          |

Table 3. Comparison among the image modalities.

Note: mSv: millisievert.

The MRI has several limitations [90,111]. A magnetic resonance imaging (MRI) scan is a painless and safe procedure that gives crisper images of the body and its tissues from any angle. This is very helpful for spotting soft tissue cancers all over the body. While an MRI scan is a reasonably safe process that does not expose you to harmful radiation, some dangers are still associated. However, MRI safety has recently become a significant emphasis in hospital and outpatient settings because of the possible attraction to ferromagnetic materials and devices. In comparison to conventional X-ray and CT scan procedures, an MRI is an exceedingly high-cost and time-consuming inquiry. The bone, bone injury, and acute brain hemorrhage are better examined using CT and X-ray techniques. It also generates motion artifacts. Some illness processes may be challenging to distinguish with an MRI. Because of the longer time it consumes and all other instruments must be vacated from the room while the MRI device runs, it is not the right investigation for emergencies. This restricts its application in trauma and emergencies, where CT scanning is frequently favored. The other details regarding the MRI and other imaging modalities used in brain tumor analysis have been provided in Appendix B.

#### 5.1. Radiomics of Brain Tumor Characterization Using AI Framework

Radiological scans have proven to be an effective non-invasive tool in the screening and diagnosing of early-stage brain cancer, treatment strategy assistance, prognosis evaluation, and follow-up for advanced-stage brain cancer [112]. Recently, radiological features have evolved from semantic to radiomic hand-crafted and deep features [48,113]. Semantic features are the qualitative features commonly extracted by the experienced radiologist directly by analyzing and looking at the clinical image to describe the lesion [114,115]. However, radiomic features entail extracting and analyzing quantitative features from medical images using mathematical algorithms, machine learning, and deep learning methods to explore possible ties with biology and clinical outcomes [116]. The anatomical

Cancers 2022. 14, 4052 15 of 37

and functional information of the lesions can be better viewed by the radiomic features extracted from structural and functional images [117]. Structural imaging is a term that describes ways of visualizing and analyzing anatomical data from the human body, such as the brain structure, tumor location, traumas, and other brain problems. Functional imaging is a term that describes methods for studying tumor physiology and molecular processes such as metabolic alterations, finer-scale lesions, and viewing brain functions [113,118,119].

# 5.1.1. Traditional Radiomics

In the traditional radiomics approach, the hand-crafted features of radiological images play an essential role. Using the AI framework, the conventional radiomics workflow involves many steps such as image acquisition, pre-processing, region of interest segmentation, feature extraction, feature reduction, model selection, evaluation, and validation in clinical implementation [120]. The vital steps of all the workflows are the radiomics feature extraction and selection from the delineated region of interest. The hand-crafted radiomics features have been generated by software implementing mathematical algorithms. The input image of manifold modalities such as MRI, CT, PET, etc., with the region of interest, is provided to a mathematical model to obtain the feature values [121,122]. The mathematical formulations of the features generated are independent of the imaging modality [123].

The wide range of hand-crafted radiomics features derived from medical images includes intensity, texture, geometric, dynamics curve, morphological, metabolic, angiogenesis, spatial, histogram, and statistical features [42,115]. These features' traits, known as radiomic features, may reveal tumoral patterns and characteristics that are not visible to the human eye. The texture and intensity feature provides the pixel and voxel label representation of a tumor and their corresponding intensity and texture representation. The radiomics texture features can serve as potential biomarkers for determining *BRAF* mutation status and as predictors [124]. The radiomics geometrics feature describes the tumor's shape, size, volume, diameter, surface, and compactness. The statistical features of the clinical images such as the mean, median, mode, maximum, minimum, variance, standard deviation, percentiles, energy, entropy, skewness, uniformity, kurtosis, bias, histogram, and dynamics curve demonstrate the statistical distribution of pixel or voxel intensities of the tumor region.

These radiomics features have provided inputs to the various supervised, semisupervised, and unsupervised machine learning models such as support vector machine (SVM), K-nearest neighbor (K-NN), decision tree (DT), random forest (RF), naïve Bayes, ensemble, artificial neural network (ANN), logistic regression, clustering, and auto-encoder for classification of a brain tumor's characterization. Finally, data analysis with the outcome of radiomics includes survival prediction, personalized treatment planning, and precision medicine.

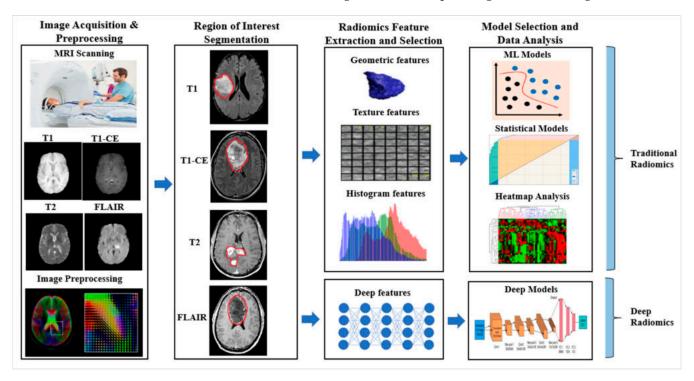
#### 5.1.2. Deep Radiomics

The deep radiomics study belongs to the DL technology, a subset of ML. The well-known DL models such as convolutional neural network (CNN) and recurrent neural network (RNN) have been successfully used in computer vision studies. The pre-trained model of CNN, such as AlexNet, ResNet, GoogleNet, etc., training on the voluminous ImageNet dataset, showed quite good performance of image features of the multi-set datasets [125]. The transfer learning (TL) also adds dimensionality to the deep feature extraction and successful computation with a suitable classifier. However, the deep radiomics study with deep feature extraction and subsequent classification is marginally better than the conventional hand-crafted radiomics study. The automatic feature extraction methods under deep radiomics select the necessary and proper features from the radiological medical image. The deeper the deep learning network, the more powerful features are extracted, and they are more abstract and meaningful and passed to the next layer of the network. The single deep learning model can perform radiomics feature extraction, selection, and

Cancers 2022, 14, 4052 16 of 37

final data analysis with the classification of brain tumors. In contrast, multiple models are involved in the case of traditional radiomics.

Image pre-processing is essential when dealing with clinical images, both for traditional and deep radiomics. Due to the varying intensity of the medical image, the presence of salt, noise, and blur, and the position of the scanners, this step is of utmost importance and enhances the performance of the model. The various pre-processing processes involved in brain tumor imaging are denoising [126], normalization, bias correction (N4ITK), image registration [29,127,128], skull stripping, and histogram matching. After the pre-processing, the region of interest (ROI) is detected, from where radiomics features are extracted. As discussed earlier, the radiomics features may be hand-crafted or deep. The final step of the radiomics process is the model selection, and data analysis from the radiomics features for better clinical decision making and treatment planning, as shown in Figure 7.



**Figure 7.** Radiomics workflow for brain lesion characterization. Notes: T1: T1-weighted MRI, T2: T2-weighted MRI, T1-CE: T1-contrast-enhanced, FLAIR: fluid-attenuated inversion recovery.

#### 6. AI Modelling Buffered with Genetics for BTC: A Radiogenomics Approach

Brain tumor classification and segmentation have been the most desired choices of the radiologist for the diagnosis process and the detection of the brain lesion. The intervention of AI for the process of brain tumor segmentation and classification has made automation in the diagnosis process and helped the radiologist to a greater extent for the detection of the lesion and to classify the tumor accurately. As discussed, genomes and genetic mutation have been the primary causes of brain tumors. Hence, detecting the status of various genomes in genetic mutation in the brain tumor can further help the diagnosis and prognosis process in a personalized treatment plan. Using ML and DL technologies, AI can detect the genetic status from radiography [129,130].

Table 4 summarizes some helpful research articles and their findings that use AI, radiomics, genomics features, and performance. The MRI is the most suitable scan for any brain disease and brain cancer because of the soft tissue level imaging. The multiparametric (T1, T1CE, T2, FLAIR, SWI, DWI, etc.) nature of MRI further improves and analyses the cancer tissues and provides their exact shape, size, location, and texture. The contrast is enhanced with external agents for MRI modalities such as T1-CE, SWI, and DWI and is more able to predict the genomic status from the better cancer features

Cancers 2022, 14, 4052 17 of 37

available in them [53,63,64,131]. From the observation, deep radiomics is the preferred radiomics feature over conventional hand-crafted radiomics features because of its deep, automatic feature extraction, and selection process. This further indicates the use of various DL models over ML models to classify and detect genomics status with better performance [53,60,61,66–68,131]. Among the various deep model signatures, the ResNet and its variants models have been considered the most desired because of the residual block to handle vanishing gradient features [60,61,65,67]. Out of all molecular signaling responsible for brain tumors, IDH and MGMT are the most common genes whose mutation causes brain cancer; hence, the prediction of the mutation status of these genes is more important for better prognosis [60,61,63,131]. The accuracy, which is the very commonly used performance metric of AI models and AUC is preferable for classification problems with probabilities to analyze the prediction more profoundly and has been used here for genetic status prediction.

| Author, Year and Reference       | Image Modality  | Radiomics Feature | Genomics Feature  | AI Model Used | Result                  |
|----------------------------------|---|-------------------|---|---------------|-------------------------|
| Akkus et al. [53] (2016)         | MRI: T1-CE, T2  | Deep radiomics    | 1p19q deletion of LGG   | DL (CNN)      | Acc.: 87.7              |
| Kickingereder et al. [64] (2016) | MRI: T1, T1-CE, FLAIR,<br>DWI, DSWCEI, PSWI             | Hand-crafted      | EGFR, PTEN, PDGFRA,<br>MDM4, CDK4<br>CDKN2A, NF1, and RB1         | ML            | Acc.: 63<br>AUC: 69     |
| Chang et al. [60] (2017)         | MRI: T1, T1-CE,<br>T2, FLAIR                            | Deep radiomics    | IDH1 prediction for LGG   | DL (ResNet)   | Acc.: 89.1<br>AUC: 95   |
| Li et al. [66]<br>(2017)         | MRI: T1, T2   | Deep radiomics    | IDH1 prediction for LGG   | DL (CNN)      | Acc.: 92.4<br>AUC: 95   |
| Liang et al. [67] (2017)         | MRI: T1, T1-CE, T2,<br>FLAIR                            | Deep radiomics    | IDH1 prediction for Glioma  | DL (DenseNet) | Acc.: 91.4<br>AUC: 94.8 |
| Korfiatis et al. [65] (2017)     | MRI: T2   | Deep radiomics    | MGMT status   | DL (ResNet50) | Acc.: 94.9              |
| Chang et al. [61] (2018)         | MRI: T1, FLAIR  | Deep radiomics    | IDH1, 1p/19q co-deletion, MGMT                                    | DL (ResNet)   | Acc.: 94<br>AUC: 91     |
| Smedley et al. [68] (2018)       | MRI: T1-CE, T2, FLAIR                                   | Deep radiomics    | Tumor<br>morphology   | DL (AE)       | MAE: 0.0114             |
| Calabrese et al. [131] (2020)    | MRI: T1, T1-CE, T2,<br>FLAIR, SWI, DWI,<br>ASLPI, HARDI | Deep radiomics    | ATRX, IDH, 7/10aneuploidy, CDKN2,<br>EGFR, TERT, PTEN, TP53, MGMT | TL (CNN+ RF)  | AUC: 97                 |
| Kawaguchi et al. [63] (2021)     | MRI: T1, T1-CE, T2,<br>FLAIR                            | Hand-crafted      | IDH, MGMT, TERT, 1p19q  | ML            | AUC: 90                 |

**Table 4.** AI model for radiogenomics study on the brain tumor.

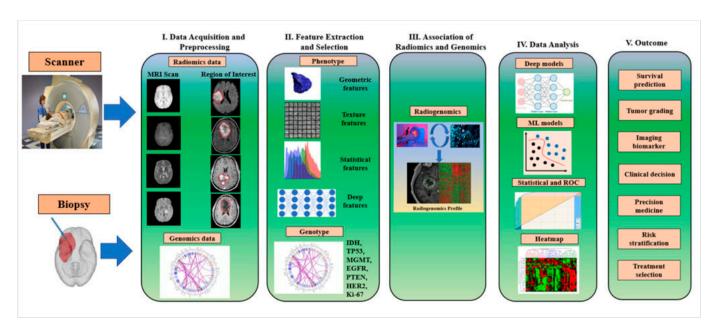
Abbreviation: DWI: diffusion-weighted image, SWI: susceptibility-weighted image, DSWCEI: dynamic susceptibility-weighted contrast-enhanced imaging, PSWI: pre-contrast susceptibility-weighted imaging, ASLPI: arterial spin labeling perfusion images, HARDI: high angular resolution diffusion imaging, Acc: accuracy, AUC: area under ROC curve, MAE: mean absolute error, AE: auto-encoder, RF: random forest.

# 6.1. Radiogenomics Pipeline for Brain Tumor Classification Using AI Model

The radiogenomics workflow for brain tumor genomics and disease characteristics can be divided into five steps, as shown in Figure 8.

Step 1. Data acquisition and pre-processing: At this stage, both radiomics and genomics data have been acquired and fed into the AI model for further processing. For the brain radiomics data, MRI scans of various modalities are preferred, and these images need pre-processing. It is wise to pre-process medical image modalities because of the various artifacts present. Additionally, medical images are captured under various scanners and consider the situation with the severity of patients. Intensity normalization, skull stripping, bias correction, denoising, and histogram matching are the major steps for pre-processing brain tumor medical images [132]. However, data augmentation is also essential while training under DL- and ML-based AI models [41,42,133,134]. Another necessary procedure is finding the region of interest (ROI) to focus on the disease characteristics and genomics features. For acquiring genomics data, a biopsy has been considered a suitable method to obtain the genomics information on the brain tumor [51].

Cancers 2022, 14, 4052 18 of 37



**Figure 8.** The workflow of radiogenomics for brain tumor genomics and disease characterization. Notes: IDH: isocitrate dehydrogenase, TP53: tumor protein53, MGMT: O6-methylguanine DNA methyltransferase, EGFR: epidermal growth factor receptor, PTEN: phosphatase and tensin homolog, HER2: human epidermal growth factor receptor 2.

Step 2. Feature extraction and selection: The radiomics features may be hand-crafted, leading to a conventional radiomics approach with subsequent ML models for lesion detection and classification. They may be deep radiomics features, where a deep learning model extracts deep, automatic, and desirable features, and the same DL model is used for subsequent segmentation, classification, and further analysis. Similarly, under the study of radiogenomics, obtaining various genomics data or genotypes features such as MGMT, IDH, EGFR, TP53, HER2, Ki-67, PTEN, etc., corresponding to the radiomics features of the patient is needed for the mapping of phenotype and genotype information and then the subsequent analysis [51,135].

Step 3. Association of radiomics and genomics: The genotype–phenotype mapping is a conceptual model in genetic architecture in the context of evolutionary biology and was first coined by P. Alberch in 1991 [136]. This amalgamates radiomics features (phenotypes) with the corresponding genomics features (genotypes) to generate the radiogenomics profile for a particular patient. In this mapping, the biological information is stored and passed on in the form of genotypes and expressed as phenotypes. The structural characteristics of mapping show that there may be phenotypic redundancy, where many genotypes map to the same phenotypes, a highly non-uniform distribution of the number of genotypes per phenotype, high phenotypic robustness, and the capacity to reach a large number of novel phenotypes in a limited number of mutational steps [137,138]. Finally, the genotype and phenotype information mapping are then passed to the AI model for the final analysis.

Step 4. Data analysis: The resulting stage of the whole radiogenomics process is the data analysis stage with appropriate model selection for genomics status prediction from the radiomics features. The classifier at this stage may be a supervised ML-based classifier such as a support vector machine (SVM), random forest (RF), K-nearest neighbor (K-NN), decision tree (DT), naïve Bayes, artificial neural network (ANN), and ensemble classifier. The various DL models involved at this stage use the softmax classifier. Apart from these, statistical models are used for statistical analysis, and performance analysis metrics such as confusion matrix and area under the ROC curve (AUC) are also used for the quantitative measurement of performance.

Step 5. Outcome: Radiogenomics holds the promise of improving diagnostic accuracy, prognosis assessment, and treatment response prediction, giving useful information for

Cancers 2022, 14, 4052 19 of 37

patient care throughout the course of the disease, given that this information is easily obtainable in imaging [139]. As shown in the workflow model, the outcome of radiogenomics involves survival prediction, tumor grading, clinical decision making, precision medicine, and personalized treatment.

# 6.2. Challenges of Radiogenomics

Recently, radiogenomics has evolved as a translational field of research with new trends in the diagnosis and prognosis of cancers and also being applicable to other severe diseases. It provides the best understanding of cancer's biology and behavior in response to standard therapy. Radiogenomics is the combination of "radiomics" and "genomics," which has significantly drawn the attention of researchers to determine imaging surrogates for genomic or molecular signatures and to advance biomarkers leveraging the numerous data types used to characterize cancer. However, the new era of technology has been confronting many issues so far from its development.

In comparison to conventional brain imaging, radiomics offers meaningful, highdimensional, quantitative data of clinical images connected to useful biologic properties. However, the acquisition, availability, accessing, and pre-processing of these radiological imaging databases is an overhead. Firstly, this kind of imaging requires a high-end scanner for its data acquisition, which is more expensive when compared to the standard scanner. Secondly, the open and public availability of these radiomics imaging databases is very limited because they are sophisticated and costlier to deal with. Thirdly, as a consequence of their availability, accessing this imaging is again more complicated due to several restrictions. To develop a robust radiomics imaging database system for cancer research, open access to resources such as gene expression and molecular characteristics and radiomics data must be available to the researchers [80,81]. Genomic and molecular information obtained through multiple tissue samples provides detailed spatial information of tumors at different locations [82]. This information could be used for heterogeneity analysis of the tumor. The research band in cancer is narrowing because of the limited availability of open and public databases. To access important cancer research data available under various super specialist cancer hospitals is even more restrictive. There are several reasons for such restrictions, such as many patients who do not want to participate, and sometimes hospitals show their restrictive behavior, patients leave their follow-ups, etc. Finally, the pre-processing of radiomics datasets is mandatory and requires high-end complex algorithmics. The pre-processing task should be so sophisticated, as it preserves the molecular information of the disease and there should be no leakage of genomics or molecular information. Again, as these radiological images are collected from different scanners of different resolutions and specifications, intensity normalization is the minimum requirement as a pre-processing step. Ultimately, the radiogenomics approach for cancer studies requires a mandatory delicate pre-processing job for suitable feature extraction and selection and subsequently better performance accomplishment.

The other challenges include the region of interest (ROI) generation, challenging model selection, critical analyzing tools, and, finally, the clinical validations of the radiogenomics outcomes. After the radiomics pre-processing steps, accurate region of interest (ROI) and segmentation of the lesion regions is the most difficult phase of radiogenomics that has a significant impact on the outcome and robustness of the system. Again, due to tumors' heterogeneity in the shape, size, location, texture, and polymorphic nature of cancer, manual ROI generation has been a tedious type of task. Hence, for the volumetric radiological data, deep learning-based segmentation methods such as U-net [140], U-Net++, V-net [141], etc., have been the first choice for ROI generation. Due to large quantitative radiomics features being present in the radiological images, in most cases, the deep learning-based imaging signatures automatically extract and select the desired and meaningful deep features rather than hand-crafted traditional machine learning algorithms for conventional radiomics features [142–145]. Again, to handle the volumetric features and outcome results generated by the top-selected model, heavy computational and data

Cancers 2022, 14, 4052 20 of 37

visualization models are required for the data analysis and statistical pattern generation. Hence, mostly, the deep learning model, hybrid deep leaning model, machine learning algorithmic models, statistical tools, heatmaps, and grad-cam models are necessary for this step. Finally, the clinical validations of the radiogenomics outcomes state that the current standards, particularly for retrospective data, lack outcomes validation, have insufficient reports of results, and have unidentified confounding variables in the source database.

#### 7. Critical Discussion

# 7.1. Principal Findings

Brain tumor characterization is an essential aspect of the diagnosis and prognosis of brain tumor disease. It helps in analyzing underlying disease characteristics, such as the genetic mutation status due to the type of brain tumor that occurs. The proposed review emphasizes the genetic behavior of the brain tumor. For that purpose, we present a comprehensive list of the various kinds of brain and CNS tumors and map the key genomics associated with the kinds of tumors in Table 2. The disease characteristics of cancer can be best analyzed from the radiology features (phenotypes or radiomics) and from the genetic features or data (genotypes or genomics). Hence, the emerging technologies for cancer study are radiomics and radiogenomics approaches. In the case of the radiomics approach, only the radiological image is considered, from which the radiomics features of diseases are extracted and analyzed. However, under the study of radiogenomics, both radiomics and genomics features of the disease are considered to analyze the status of the mutation in brain tumors. Radiomics and radiogenomics, the emerging fields, deal with all kinds of cancer diagnoses and prognosis processes for personalized treatment. However, our statistical analysis suggests extensive research has been conducted on brain tumors (Figure 3b). Therefore, magnetic resonance imaging (MRI) is a suitable imaging modality for brain scanning as it provides soft tissue level characteristics in images without any harmless ionizing radiation, unlike in the case of CT and X-ray imaging (Figure 3a).

The AI has performed supremely in brain cancer detection and diagnosis in the recent past. Hence, it makes the use of radiogenomics approaches for brain tumor detection, diagnosis, and even for personalized treatment and medicine more successful. Out of the ML, DL, and TL models of AI, the DL and DL with TL models are the most suitable and adaptable for radiogenomics as they deal with automatic deep radiomics features extraction and selection for the analysis of brain tumor mutation status (Figure 2b). The various successful DL models involved in the radiogenomics study for brain tumors are convolutional neural network (CNN), VGG, ResNet, DenseNet, and GoogleNet (Figure 2c). This signifies the use of a deep learning-based softmax classifier for predicting the status of genomics mutation in brain tumors (Figure 2d). Therefore, it proves that AI paradigms are advantageous in brain tumor diagnosis and prognosis with radiogenomics. Recently, emerging technologies have helped to a great extent in cancer diagnosis research, with most of the research carried out in the United States of America (Figure 2a).

#### 7.2. Benchmarking

Tumor characterization by the emerging technology of radiomics and radiogenomics has been an active research area among researchers in the recent past. Very little research has been conducted using the AI framework to predict genetic mutation status in various cancer. Additionally, there are a few reviews regarding tumor characterization in the radiomics and radiogenomics paradigms that have been published [49,51,113,114,135,139,146–148]. However, only a few are dedicated to brain tumors [49,135,148]. Even though these three studies [49,135,148] dedicatedly discuss the genetic characteristics of brain tumors, they have not covered several aspects for review altogether. Overall, the review articles on cancer characterization are summarized in Table 5. Some did not discuss the AI framework on the prospect of radiomics and radiogenomics. However, some of them discussed only radiomics and only radiogenomics parts separately and may miss the AI framework. Hence, they do not consider an all-round review of these emerging technologies. The proposed review on

Cancers 2022, 14, 4052 21 of 37

specialized cancer (brain) characterization, while considering radiomics and radiogenomics individually in the AI framework, our specialization also includes the statistical portrayal of radiogenomics studies of cancer characterization along with risk-of-bias (RoB) analysis.

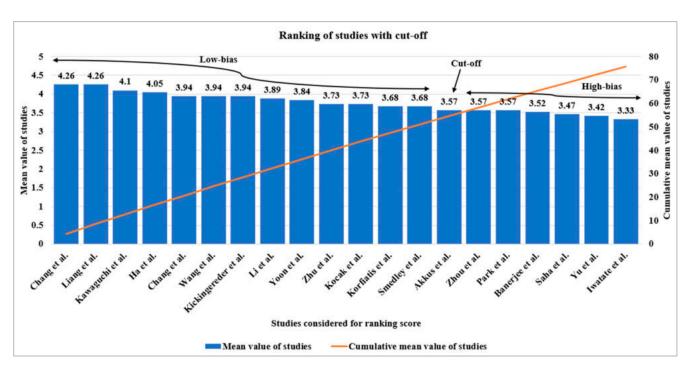
| <b>Table</b> | 5. | Benchm | arking | table. |
|--------------|----|--------|--------|--------|
|--------------|----|--------|--------|--------|

| Author, Year and<br>Reference | Radiomics | Radio-<br>Genomics | AI Framework | Anatomical Cancer<br>Discussed | Statistics and<br>Risk-of-Bias<br>(RoB) Analysis |
|-------------------------------|-----------|--------------------|--------------|--------------------------------|--|
| Rizzo et al. (2018) [114]     | ✓         | ×                  | ×            | Generalized                    | ×  |
| Kazerooni et al. (2019) [135] | ×         | ✓                  | ×            | Brain (Glioblastoma)           | ×  |
| Bodalal et al. (2019) [146]   | ×         | ✓                  | ✓            | All Cancer                     | ×  |
| Trivizakis et al. (2020) [51] | ×         | ✓                  | ✓            | All Cancer                     | ×  |
| Gullo et al. (2020) [139]     | ×         | ✓                  | ×            | All Cancer                     | Statistical analysis only                        |
| Shui et al. (2021) [147]      | ×         | ✓                  | ✓            | All Cancer                     | ×  |
| Singh et al. (2021) [148]     | ✓         | ✓                  | ×            | Brain (Glioma)                 | ×  |
| Wu et al. (2021) [113]        | ✓         | ×                  | ✓            | Lung                           | ×  |
| Habib et al. (2021) [49]      | ✓         | ✓                  | ×            | Brain                          | ×  |
| Jena et al. (2022) (Proposed) | ✓         | ✓                  | ✓            | Brain                          | <b>√</b>   |

#### 7.3. Risk-of-Bias Analysis of Radiogenomics Studies

We have considered 20 radiogenomics studies using AI such as DL, ML, and TL to check the bias in these studies to show that AI is good for radiogenomics studies for oncology care. Each study has 19 AI-based attributes such as phenotype, genotype, AI model used number of AI model used, anatomical area of the tumor, image modality, subimage modality, image pre-processing, dataset size, demographic information of dataset, feature extraction type, feature selection type, data augmentation, classifier information, statistical analysis performed, number of performance evaluation parameters, accuracy, area under ROC curve, and impact factor of the journal. These radiogenomics attributes using AI features are initially qualitative and then quantified by assigning a number between 1 and 5 based on the consensus of the AI scientist's experience. The value of AI-based attributes has been assigned based on the strength of the attribute that ranges from 1 to 5 of five different classes such as low, moderate, high-moderate, low-of-a-high, and high-of-a-high. Then each individual study's aggregate score is the sum of all attribute values for that selected study. The mean of each study was then calculated by dividing by the number of AI attributes considered (i.e., 19 in our case). Using this principle, all the studies (i.e., 20) are ranked based on their mean score, which ranges from 4.26 to 3.33, and plotted in decreasing order as shown in Figure 9. The raw cut-off of 3.57 was determined based on the intersection of the "cumulative plot of the mean score" and "the frequency plot curve of the studies". This raw cut-off mark estimates the whole number of studies into low-bias and high-bias categories. The higher the mean value, the lower is the risk-of-bias; hence, studies above the cut-off mark belong to the low-bias category, while 35% of studies [53–59] belong to a high-bias category. The highly biased studies have not considered all AI attributes while evaluating the radiogenomics' system and may have the lower proportioned values for the attributes considered.

Cancers 2022, 14, 4052 22 of 37



**Figure 9.** The ranking score technique shows the frequency distribution of radiogenomics studies in descending order succeeded by the cumulative plot, showing the raw cut-off mark for bias analysis [51–71].

#### 7.4. Advanced Features of Artificial Intelligence

There have been two significant developments in the field of AI that cannot be ignored, namely, (a) pruning of AI (PAI) models and (b) explainable AI (XAI). The concept of pruning AI is motivated by the amount of storage needed during the training process of the AI models [149]. In contrast, the concept of explainable AI is inspired by the process of knowing how the AI black box performs [150,151]. The origination of evidence for scientific validation is also nowadays coined under the umbrella of explainable AI because XAI is used for satisfying the evidence for scientific validation [15,123,128,152–154]. Recently, explainable artificial intelligence (XAI) has attracted much interest in medicine. To reach a level of explainable medicine we need causability, where causability encompasses measurements for the quality of explanations, in the same way, that usability encompasses measurements for the quality of use [155–157].

# 7.5. Strength, Weakness, and Extension

Radiomics and radiogenomics are the next generations of methodologies for non-invasive cancer treatments, which consider the radiological features and molecular characteristics of the disease. The quantitative imaging feature (phenotype or radiomics) describes the disease characteristics in the case of radiomics analysis. Similarly, both the phenotype and genotype of the cancer are useful for radiogenomics analysis. However, artificial intelligence frameworks such as ML, DL, and TL have provided an edge over traditional analysis of the disease. They have many positive outcomes such as survival prediction, tumor grading, imaging biomarker, clinical decision making, treatment planning, risk stratification, personalized treatment, and precision medicine. As it provides an entirely non-invasive diagnosis with an effective outcome, it is the most considered and most rated treatment choice over invasive treatment methods such as biopsy, surgery, chemotherapy, and radiation therapy.

Over the past few years, numerous research analyses have been carried out on this emerging technology; however, systematic computation is still a matter of concern due to several factors. The scarcity and the lack of availability of standard and volumetrics radiogenomics public datasets may lead to poor training of the AI models of ML and DL.

Cancers 2022. 14, 4052 23 of 37

The training can be improved by breeding, fusion, or stochastic imaging methods with AI models for superior segmentation paradigms. These models are data-centric and provide better accuracy on the test dataset when trained with volumetric datasets. Additionally, multi-regional, inter-, and intra-institutional heterogeneity datasets are needed as the genetic behaviors of the different demographic areas vary. Gene expression and signaling pathways are exceptionally complex, therefore it is tiresome work to generate a huge quantity of data from whole-genome sequencing with imaging data. As we observed from the statistical analysis, most of the current research on radiogenomics has been conducted in the USA; hence, other parts of the globe are still unable to share its view. Hence, there should be a need for the involvement of multi-center studies. Examples of multi-center studies can be seen here for other applications [34,38,158]. Another extension would be to use extensive data analysis for the radiogenomics [159].

Furthermore, one could study the brain connectivity in brain tumor patients [160,161]. Association studies can be incorporated, which study the relationships between brain tumor radiomics and genomics for survival analysis [129,162]. Radiogenomics is inevitable and needs further high-level, multi-center, multi-intuitional, multi-geographical research. The standardized workflow of radiogenomics studies needs a computational guide to provide a different level of performance. Lastly, there is the opportunity of studying the effect of COVID-19 on brain tumor patients [163].

#### 8. Conclusions

A brain tumor is a deadly disease where several biological assessments and biomedical research studies have been conducted to understand the tumorigenesis over the decade. Brain tumor characterization is the process of learning the molecular signature of the underlying nature of the disease in terms of lesion segmentation, classification, detection, and analysis. Radiomics and radiogenomics are relatively new fields of research on cancer that provide hope for the most acceptable assessment methods. Radiogenomics is the association between established MR imaging features and the molecular characteristics of brain tumors to improve disease diagnosis and characterization with greater precision. Radiomics and radiogenomics are promising fields in cancer research treatment providing personalized treatment planning, precision medicine, and imaging biomarkers during different stages of treatment of the patients. These technologies are not intended to replace the radiologist completely; however, they need to accept this new technology and make the necessary adjustments to accommodate the changes it will bring to the clinical process.

**Author Contributions:** Conceptualization, B.J., S.S., G.K.N., J.R.L. and J.S.S.; resources, S.S., B.J., M.M.F., N.N.K., G.K.N. and J.S.S.; writing—original draft preparation, S.S., B.J., N.G., N.N.K. and J.R.L., writing—review and editing, B.J., N.G., A.B., M.K.K., L.S. and J.S.S.; supervision, S.S., M.K.K., M.M.F., L.S. and J.S.S. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

**Conflicts of Interest:** The authors declare no conflict of interest.

#### Appendix A

**Table A1.** WHO's CNS and brain tumor classification system of 2021 [52].

- 1. Gliomas, glioneuronal tumors, and neuronal tumors
- 1.1 Adult-type diffuse gliomas
- 1.1.1 Astrocytoma, IDH-mutant
- 1.1.2 Glioblastoma, IDH-wildtype
- $1.1.3\ Oligodendrog lioma, IDH-mutant, and\ 1p/19q-co-deleted$

Cancers 2022, 14, 4052 24 of 37

# Table A1. Cont. 1.2 Pediatric-type diffuse low-grade gliomas 1.2.1 Angiocentric glioma 1.2.2 Diffuse astrocytoma, MYB- or MYBL1-altered 1.2.3 Diffuse low-grade glioma, MAPK pathway-altered 1.2.4 Polymorphous low-grade neuroepithelial tumor of the young 1.3 Pediatric-type diffuse high-grade gliomas 1.3.1 Infant-type hemispheric glioma 1.3.2 Diffuse midline glioma, H3 K27-altered 1.3.3 Diffuse pediatric-type high-grade glioma, H3-wildtype and IDH-wildtype 1.3.4 Diffuse hemispheric glioma, H3 G34-mutant 1.4 Circumscribed astrocytic gliomas 1.4.1 Chordoid glioma 1.4.2 Pilocytic astrocytoma 1.4.3 Pleomorphic xanthoastrocytoma 1.4.4 High-grade astrocytoma with piloid features 1.4.5 Astroblastoma, MN1-altered 1.4.6 Subependymal giant cell astrocytoma 1.5 Glioneuronal and neuronal tumors 1.5.1 Ganglioglioma 1.5.2 Gangliocytoma 1.5.3 Central neurocytoma 1.5.4 Dysembryoplastic neuroepithelial tumor 1.5.5 Desmoplastic infantile ganglioglioma/desmoplastic infantile astrocytoma 1.5.6 Papillary glioneuronal tumor 1.5.7 Diffuse glioneuronal tumor with oligodendroglioma-like features and nuclear clusters 1.5.8 Myxoid glioneuronal tumor 1.5.9 Rosette-forming glioneuronal tumor 1.5.10 Multi-nodular and vacuolating neuronal tumor 1.5.11 Diffuse leptomeningeal glioneuronal tumor 1.5.12 Dysplastic cerebellar gangliocytoma (Lhermitte—Duclos disease) 1.5.13 Cerebellar liponeurocytoma 1.5.14 Extraventricular neurocytoma 1.6 Ependymal tumors 1.6.1 Posterior fossa ependymoma 1.6.2 Posterior fossa ependymoma, group PFA 1.6.3 Posterior fossa ependymoma, group PFB 1.6.4 Supratentorial ependymoma 1.6.5 Supratentorial ependymoma, ZFTA fusion-positive 1.6.6 Supratentorial ependymoma, YAP1 fusion-positive 1.6.7 Spinal ependymoma, MYCN-amplified 1.6.8 Spinal ependymoma 1.6.9 Subependymoma 1.6.10 Myxopapillary ependymoma 2. Choroid plexus tumors 2.1 Choroid plexus papilloma 2.2 Choroid plexus carcinoma 2.3 Atypical choroid plexus papilloma 3. Embryonal tumors 3.1 Medulloblastoma 3.1.1 Medulloblastomas, molecularly defined 3.1.1.1 Medulloblastoma, WNT-activated 3.1.1.2 Medulloblastoma, SHH-activated and TP53-mutant

3.1.1.3 Medulloblastoma, SHH-activated and TP53-wildtype

3.1.1.4 Medulloblastoma, non-WNT/non-SHH3.1.2 Medulloblastomas, histologically defined

Cancers **2022**, 14, 4052 25 of 37

## Table A1. Cont.

|   | 3.2 Other CNS embryonal tumors   |
|---|--|
|   | 3.2.1 Cribriform neuroepithelial tumor 3.2.2 Atypical teratoid/rhabdoid tumor 3.2.3 Embryonal tumor with multi-layered rosettes 3.2.4 CNS tumor with BCOR internal tandem duplication 3.2.5 CNS neuroblastoma, FOXR2-activated 3.2.6 CNS embryonal tumor |
|   | 4. Pineal tumors   |
| - | 4.1 Pineocytoma 4.2 Pineoblastoma 4.3 Pineal parenchymal tumor of intermediate differentiation 4.4 Desmoplastic myxoid tumor of the pineal region, SMARCB1-mutant 4.5 Papillary tumor of the pineal region   |
|   | 5. Cranial and paraspinal nerve tumors   |
| _ | 5.1 Schwannoma 5.2 Perineurioma 5.3 Neurofibroma 5.4 Malignant melanotic nerve sheath tumor 5.5 Malignant peripheral nerve sheath tumor 5.6 Hybrid nerve sheath tumor 5.7 Paraganglioma  |
|   | 6. Meningiomas   |
|   | 7. Mesenchymal, non-meningothelial tumors  |
|   | 7.1 Soft tissue tumors   |
|   | 7.1.1 Fibroblastic and myofibroblastic tumors  |
|   | 7.1.1.1 Solitary fibrous tumor   |
|   | 7.2 Vascular tumors  |
|   | 7.2.1 Hemangioblastoma 7.2.2 Hemangiomas and vascular malformations  |
|   | 7.3 Skeletal muscle tumors   |
|   | 7.3.1 Rhabdomyosarcoma   |
|   | 7.4 Uncertain differentiation  |
|   | 7.4.1 Primary intracranial sarcoma, DICER1-mutant 7.4.2 Intracranial mesenchymal tumor, FET-CREB fusion-positive 7.4.3 CIC-rearranged sarcoma 7.4.4 Ewing sarcoma  |
|   | 7.5 Chondro-osseous tumors   |
|   | 7.5.1 Chondrogenic tumors  |
|   | 7.5.1.1 Mesenchymal chondrosarcoma<br>7.5.1.2 Chondrosarcoma   |
|   | 7.5.2 Notochordal tumors   |
|   | 7.5.2.1 Chordoma (including poorly differentiated chordoma)  |
|   | 8. Melanocytic tumors  |
|   | 8.1 Diffuse meningeal melanocytic neoplasms  |
| _ | 8.1.1 Meningeal melanocytosis and meningeal melanomatosis  |
|   | 8.2 Circumscribed meningeal melanocytic neoplasms  |
|   | 9. Hematolymphoid tumors   |
|   | 9.1 Lymphomas  |
|   | 9.1.1 CNS lymphomas  |
|   | 9.1.1.1 Lymphomatoid granulomatosis 9.1.1.2 Primary diffuse large B-cell lymphoma of the CNS 9.1.1.3 Intravascular large B-cell lymphoma 9.1.1.4 Immunodeficiency-associated CNS lymphoma  |
|   |  |

Cancers 2022. 14, 4052 26 of 37

#### Table A1. Cont.

| 9.1.2 Miscellaneous rare lymphomas in the CNS  |
|--|
| 9.1.2.1 Other low-grade B-cell lymphomas of the CNS 9.1.2.2 MALT lymphoma of the dura 9.1.2.3 T-cell and NK/T-cell lymphomas 9.1.2.4 Anaplastic large cell lymphoma ( $ALK+/ALK-$ )                                    |
| 9.2 Histiocytic tumors   |
| 9.2.1 Rosai–Dorfman disease 9.2.2 Erdheim–Chester disease 9.2.3 Juvenile xanthogranuloma 9.2.4 Histiocytic sarcoma 9.2.5 Langerhans cell histiocytosis   |
| 10. Germ cell tumors   |
| 10.1 Germinoma 10.2 Immature teratoma 10.3 Choriocarcinoma 10.4 Mature teratoma 10.5 Embryonal carcinoma 10.6 Mixed germ cell tumor 10.7 Teratoma with somatic-type malignancy 10.8 Yolk sac tumor                     |
| 11. Tumors of the sellar region  |
| 11.1 Pituitary blastoma 11.2 Pituitary adenoma/PitNET 11.3 Papillary craniopharyngioma 11.4 Adamantinomatous craniopharyngioma 11.5 Pituicytoma, granular cell tumor of the sellar region, and spindle cell oncocytoma |
| 12. Metastases to the CNS  |
| 12.1 Metastases to the meninges 12.2 Metastases to the brain and spinal cord parenchyma  |

# Appendix B

Appendix B.1. Magnetic Resonance Imaging (MRI)

The MRI is a non-invasive, highly sophisticated, highly detailed, three-dimensional imaging diagnostic method of the interior structures of the human body. The MRI scanner machine is a big cylindrical-shaped tube surrounded by a magnet as shown in Figure A1a. MRI creates images by using radio waves rather than X-rays, and it provides superb detail of the body's soft tissues. It is very useful for detecting aberrant tissues in soft tissues such as the brain, spinal cord, muscles, and ligaments; hence, it is often used in the detection of tumors, strokes, and hemorrhage. The life-saving medical imaging technique is nuclear magnetic resonance (NMR) imaging, which uses the property of NMR to image nuclei of atoms within the body. In September 1971, Paul C. Lauterbur established a way to encode spatial information into an NMR signal using magnetic field gradients, according to the history of MR imaging [111].

Cancers 2022, 14, 4052 27 of 37

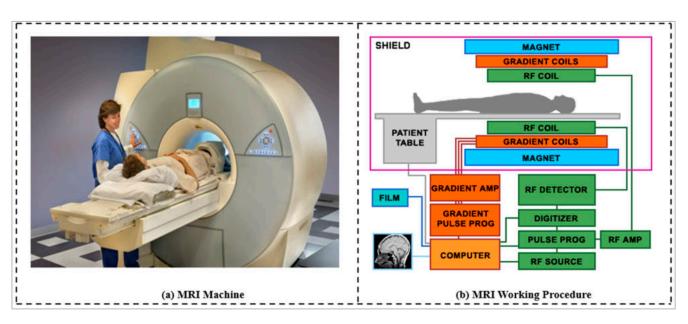


Figure A1. (a) MRI scanner machine. (b) Working procedure of MRI.

# Appendix B.1.1. Image View or MRI Planes

MRI scans, as with CT scans, can have three different anatomical views such as axial, sagittal, and coronal. These basic orientation MRIs of the body decide the image view or planes. From the top-down would be an axial or transverse plane; from the side would be a sagittal plane; from the front to back would be a coronal plane, as shown in Figure A2 (top).

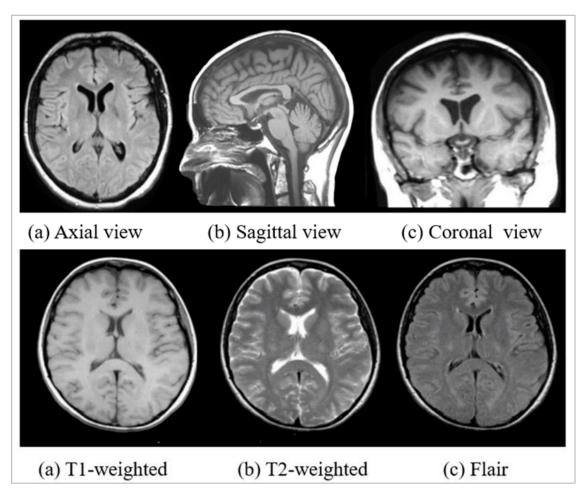
# Appendix B.1.2. Image Weight or Sequence

The weight of the image or MRI sequences is determined by the magnetic field produced by the scanner and various time factors. T1-weighted and T2-weighted scans are the most common MRI sequences. The Fluid-Attenuated Inversion Recovery (FLAIR) sequence is the third most widely utilized sequence. The comparison among all the primary imaging sequences has been depicted in Figure A2 (bottom) and Table A2.

| Table A2. | Comparison | of all basic | weighted | images of MRI. |
|-----------|------------|--------------|----------|----------------|
|-----------|------------|--------------|----------|----------------|

| T1-Weighted | T2-Weighted                 | Flair  |
|-------------|-----------------------------|--|
| Dark        | Bright                      | Dark   |
| Light       | Dark Gray                   | Dark Gray  |
| Gray        | Light Gray                  | Light Gray   |
| Bright      | Light                       | Light  |
| Dark        | Bright                      | Bright   |
| Dark        | Dark                        | Light  |
|             | Dark Light Gray Bright Dark | Dark Bright Light Dark Gray Gray Light Gray Bright Light Dark Bright |

Cancers 2022, 14, 4052 28 of 37

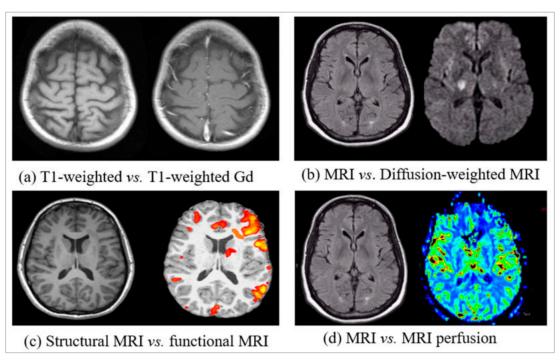


**Figure A2.** The three prominent views of MRI scanning: axial, sagittal, coronal (Top), and brain MRI slices of T1-weighted vs. T2-weighted and flair (Bottom).

# Appendix B.1.3. Functional and Perfusion, Diffusion MRI

The ubiquitous and important sequence of brain images is T1-CE (T1-contrast-enhanced or post-contrast T1), which the clinician prefers most in general practice. This type of weighted image is generated when a non-toxic Gadolinium (Gad), a non-toxic paramagnetic contrast enhancement agent, is injected during the T1-weighted scan. Gad-enhanced signal intensities or bright signals of blood vessels in images are useful for looking at vascular structures and breakdown in the blood–brain barrier. However, the other sequences of MRI also exist apart from these three having their own importance based on the requirement. They include (a) functional MRI (fMRI) [164], (b) MR spectroscopy (MRS) [165], (c) diffusion-weighted imaging (DWI) [164], (d) proton density-weighted imaging (PD-WI) [164], (e) MR perfusion [166], and (f) tractography [167]. Figure A3 depicts the differences between some of these above types of MRI and the standard modality of MRI. However, the other imaging modalities apart from MRI, such as CT, PET, and SPECT, have been second-choice imaging techniques when considering brain images.

Cancers 2022, 14, 4052 29 of 37



**Figure A3.** Comparison of standard MRI with (a) Gad-enhanced, (c) functional, (b) diffusion, and (d) perfusion MRI.

# Appendix B.1.4. Properties and Working Procedure of MRI

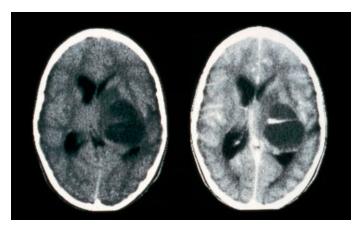
MRI is based on the magnetization properties of atomic nuclei, and three major functionality components of an MRI scanner are the magnet, gradient, and the radiofrequency (RF) system [109,168]. The human body comprises water ( $H_2O$ ), mainly made of hydrogen ions ( $H^+$ ), i.e., protons. The strong magnetic field produced by the external magnets present in the MRI scanner can act on these positively charged hydrogen ions of the tissue being examined and cause them to spin in an identical manner. By varying the strength and direction of this magnetic field, the direction of spin of the protons changes, enabling them to build a layer of details. When a radiofrequency (RF) current is then pulsed through the patient, the protons are stimulated and spin out of equilibrium, straining against the pull of the magnetic field. When the RF field is turned off, the MRI sensors are able to detect the energy released as the protons realign with the magnetic field. Physicians can tell the difference between various types of tissues based on different types of images created by this MRI mechanism. The complete working procedure diagram is shown in Figure A1b.

Appendix B.2. Other Imaging Modalities for Brain Image

Appendix B.2.1. Computed Tomography (CT)

CT [169] has been considered the most used radiography for brain tumor analysis after the MRI scans. *CT scanning* is painless, non-invasive, and an accurate imaging diagnosis technique adapted by the medical fraternity. Under this imaging technology, an X-ray beam acts as the primary source of capturing the image of the intended body part with a series of 2-D scans. These 2-D cross-sectional scans then combine to form the 3-D volume to better view the internal body organs. Due to the use of an X-ray beam of ionizing radiation, it is considered harmful to the living tissue of the human body and can cause cancer. However, this imaging technology is faster than other imaging technology to capture the internal tissue level, blood vessel, and skeletal structure image. Therefore, it is recommended by the health professional in an emergency for rapid assessment of the patient's condition to start the initial treatments to save their life. It is useful to detect brain cancer along with other cancer because of its tissue-level imaging capacity. Figure A4 shows the CT scans of brain cancer for glioma.

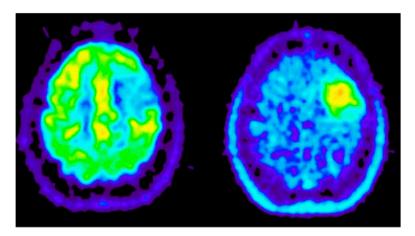
Cancers 2022, 14, 4052 30 of 37



**Figure A4.** CT scans of brain tumors (glioma). (Image source: www.wellcomecollection.org, accessed on 9 December 2021).

# Appendix B.2.2. Positron Emission Tomography (PET)

The PET [170] is a functional imaging technique that covers the image of tissue-level characterization to diagnose cancer precisely. PET is the nuclear medicine imaging that uses radioactive substances known as contrast agents or radiotracers or radiopharmaceuticals in the body to better highlight abnormal tissues. This is also non-invasive to help diagnose and assess medical conditions. A PET scan is also a variant of a CT scan and is sometimes performed along with a CT scan called a PET-CT scan to learn the tissue imaging of the cancerous regions. This perfusion PET imaging can detect the metabolic, cell proliferation, oxygenation, and apoptosis of the cancer region clearly to help the doctor for a minute-level inspection. Figure A5 shows the variants of PET scans such as <sup>18</sup>F-PET and <sup>18</sup>F-FDG PET scans.



**Figure A5.** <sup>18</sup>F-PET and <sup>18</sup>F-FDG PET scans. (Image source: www.semanticscholar.org, accessed on 9 December 2021).

# Appendix B.2.3. Single-Photon Emission Computed Tomography (SPECT)

The SPECT [171] is a nuclear tomographic imaging methodology using gamma rays that reveals the process of blood flows to tissues and organs as shown in Figure A6. The diagnostic imaging technique is in the form of 3D images of organs by multiple 2D cross-sectional images, as in the case of the CT scanning technique. The gamma-emitting radioisotope is infused into the patient's bloodstream before the SPECT scan. This imaging technique helps medical professionals to diagnose tumors, stroke, seizures, fractures, and infections. It is more sensitive to brain injury than either the MRI or CT imaging technique because it can detect reduced blood flow to injured sites.

Cancers 2022, 14, 4052 31 of 37

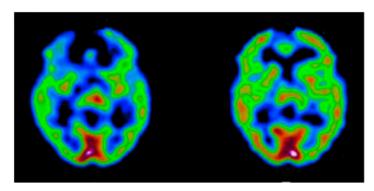


Figure A6. 99mTc-HMPAO SPECT scans [172].

#### References

1. Das, S.; Bose, S.; Nayak, G.K.; Satapathy, S.C.; Saxena, S. Brain tumor segmentation and overall survival period prediction in glioblastoma multiforme using radiomic features. *Concurr. Comput. Pr. Exp.* **2021**, *34*, e6501. [CrossRef]

- 2. Brain Tumor: Statistics. Available online: https://www.cancer.net/cancer-types/brain-tumor/statistics (accessed on 20 May 2022).
- 3. Khazaei, Z.; Goodarzi, E.; Borhaninejad, V.; Iranmanesh, F.; Mirshekarpour, H.; Mirzaei, B.; Naemi, H.; Bechashk, S.M.; Darvishi, I.; Sarabi, R.E.; et al. The association between incidence and mortality of brain cancer and human development index (HDI): An ecological study. *BMC Public Health* **2020**, *20*, 1–7. [CrossRef] [PubMed]
- 4. World Health Organization. The World Health Report 2001: Mental Health: New Understanding, New Hope. 2001. Available online: https://apps.who.int/iris/handle/10665/42390 (accessed on 9 December 2021).
- 5. Jena, B.; Nayak, G.K.; Saxena, S. An empirical study of different machine learning techniques for brain tumor classification and subsequent segmentation using hybrid texture feature. *Mach. Vis. Appl.* **2021**, *33*, 1–16. [CrossRef]
- 6. Morawski, M.; Kirilina, E.; Scherf, N.; Jäger, C.; Reimann, K.; Trampel, R.; Gavriilidis, F.; Geyer, S.; Biedermann, B.; Arendt, T.; et al. Developing 3D microscopy with CLARITY on human brain tissue: Towards a tool for informing and validating MRI-based histology. *NeuroImage* 2017, 182, 417–428. [CrossRef] [PubMed]
- 7. Broocks, G.; Flottmann, F.; Ernst, M.; Faizy, T.D.; Minnerup, J.; Siemonsen, S.; Fiehler, J.; Kemmling, A. Computed Tomography–Based Imaging of Voxel-Wise Lesion Water Uptake in Ischemic Brain. *Investig. Radiol.* **2018**, *53*, 207–213. [CrossRef]
- 8. Sanches, J.M.; Laine, A.F.; Suri, J.S. Ultrasound Imaging; Springer: Berlin/Heidelberg, Germany, 2012.
- 9. Saba, L.; Suri, J.S. *Neurological Disorders and Imaging Physics, Volume 1; Application of Multiple Sclerosis*; IOP Publishing Ltd.: Bristol, UK, 2019; Volume 1.
- 10. El-Baz, A.; Suri, J.S. Application to autism spectrum disorders and Alzheimer's. In *Neurological Disorders and Imaging Physics*; IOP Publishing Ltd.: Bristol, UK, 2019; Volume 3.
- 11. El-Baz, A.; Suri, J.S. Engineering and clinical perspectives of multiple sclerosis. In *Neurological Disorders and Imaging Physics*; IOP Publishing Ltd.: Bristol, UK, 2019; Volume 2.
- 12. Weiskopf, N.; Edwards, L.J.; Helms, G.; Mohammadi, S.; Kirilina, E. Quantitative magnetic resonance imaging of brain anatomy and in vivo histology. *Nat. Rev. Phys.* **2021**, *3*, 570–588. [CrossRef]
- 13. Shehata, M.; Khalifa, F.; Soliman, A.; Shaker, S.; Shalaby, A.; El-Baz, M.; Mahmoud, A.; Dwyer, A.C.; El-Ghar, M.A.; Ghazal, M.; et al. Early Classification of Renal Rejection Types: A Deep Learning Approach. In *Machine Learning in Medicine*; CRC Press: Boca Raton, FL, USA, 2021; pp. 257–280. [CrossRef]
- 14. Tandel, G.S.; Balestrieri, A.; Jujaray, T.; Khanna, N.N.; Saba, L.; Suri, J.S. Multiclass magnetic resonance imaging brain tumor classification using artificial intelligence paradigm. *Comput. Biol. Med.* **2020**, 122, 103804. [CrossRef]
- 15. Kuppili, V.; Biswas, M.; Sreekumar, A.; Suri, H.S.; Saba, L.; Edla, D.R.; Marinhoe, R.T.; Sanches, J.; Suri, J.S. Extreme Learning Machine Framework for Risk Stratification of Fatty Liver Disease Using Ultrasound Tissue Characterization. J. Med Syst. 2017, 41, 152. [CrossRef]
- 16. Acharya, U.R.; Faust, O.; Sree, S.V.; Molinari, F.; Suri, J.S. ThyroScreen system: High resolution ultrasound thyroid image characterization into benign and malignant classes using novel combination of texture and discrete wavelet transform. *Comput. Methods Programs Biomed.* **2011**, 107, 233–241. [CrossRef]
- 17. Acharya, U.R.; Sree, S.V.; Krishnan, M.M.R.; Molinari, F.; Garberoglio, R.; Suri, J.S. Non-invasive automated 3D thyroid lesion classification in ultrasound: A class of ThyroScan™ systems. *Ultrasonics* **2012**, *52*, 508–520. [CrossRef]
- 18. Acharya, U.R.; Sree, S.V.; Swapna, G.; Gupta, S.; Molinari, F.; Garberoglio, R.; Witkowska, A.; Suri, J.S. Effect of complex wavelet transform filter on thyroid tumor classification in three-dimensional ultrasound. *Proc. Inst. Mech. Eng. Part H J. Eng. Med.* 2013, 227, 284–292. [CrossRef] [PubMed]
- 19. Suri, J.S.; Rangayyan, R.M. Recent Advances in Breast Imaging, Mammography, and Computer-Aided Diagnosis of Breast Cancer. 2006. Available online: https://spie.org/Publications/Book/651880 (accessed on 9 December 2021).
- 20. Huang, S.-F.; Ruey-Feng, C.; Moon, W.K.; Lee, Y.-H.; dar-Ren, C.; Suri, J.S. Analysis of Tumor Vascularity Using Three-Dimensional Power Doppler Ultrasound Images. *IEEE Trans. Med Imaging* **2008**, *27*, 320–330. [CrossRef] [PubMed]

Cancers 2022, 14, 4052 32 of 37

21. Acharya, U.R.; Mookiah, M.R.K.; Sree, S.V.; Yanti, R.; Martis, R.J.; Saba, L.; Molinari, F.; Guerriero, S.; Suri, J.S. Evolutionary Algorithm-Based Classifier Parameter Tuning for Automatic Ovarian Cancer Tissue Characterization and Classification. *Ultraschall der Med. -Eur. J. Ultrasound* 2012, 35, 237–245. [CrossRef] [PubMed]

- 22. Acharya, U.R.; Sree, S.V.; Kulshreshtha, S.; Molinari, F.; Koh, J.E.W.; Saba, L.; Suri, J.S. GyneScan: An Improved Online Paradigm for Screening of Ovarian Cancer via Tissue Characterization. *Technol. Cancer Res. Treat.* **2014**, *13*, 529–539. [CrossRef]
- 23. Shrivastava, V.; Londhe, N.D.; Sonawane, R.; Suri, J.S. Reliable and accurate psoriasis disease classification in dermatology images using comprehensive feature space in machine learning paradigm. *Expert Syst. Appl.* **2015**, 42, 6184–6195. [CrossRef]
- Shrivastava, V.; Londhe, N.D.; Sonawane, R.S.; Suri, J.S. Computer-aided diagnosis of psoriasis skin images with HOS, texture and color features: A first comparative study of its kind. Comput. Methods Programs Biomed. 2016, 126, 98–109. [CrossRef]
- 25. Maniruzzaman; Kumar, N.; Abedin, M.; Islam, S.; Suri, H.S.; El-Baz, A.S.; Suri, J.S. Comparative approaches for classification of diabetes mellitus data: Machine learning paradigm. *Comput. Methods Programs Biomed.* **2017**, *152*, 23–34. [CrossRef]
- 26. Maniruzzaman; Rahman, J.; Hasan, A.M.; Suri, H.S.; Abedin, M.; El-Baz, A.; Suri, J.S. Accurate Diabetes Risk Stratification Using Machine Learning: Role of Missing Value and Outliers. *J. Med Syst.* **2018**, *42*, 92. [CrossRef]
- 27. Sharma, P.; Choudhary, K.; Gupta, K.; Chawla, R.; Gupta, D.; Sharma, A. Artificial plant optimization algorithm to detect heart rate & presence of heart disease using machine learning. *Artif. Intell. Med.* **2019**, *102*, 101752. [CrossRef]
- 28. Acharya, U.R.; Faust, O.; S, V.S.; Alvin, A.; Krishnamurthi, G.; Seabra, J.; Sanches, J.; Suri, J.S. Understanding symptomatology of atherosclerotic plaque by image-based tissue characterization. *Comput. Methods Programs Biomed.* **2013**, 110, 66–75. [CrossRef]
- 29. Araki, T.; Ikeda, N.; Dey, N.; Chakraborty, S.; Saba, L.; Kumar, D.; Godia, E.C.; Jiang, X.; Gupta, A.; Radeva, P.; et al. A comparative approach of four different image registration techniques for quantitative assessment of coronary artery calcium lesions using intravascular ultrasound. *Comput. Methods Programs Biomed.* **2015**, *118*, 158–172. [CrossRef]
- Acharya, U.R.; Sree, S.V.; Krishnan, M.M.R.; Krishnananda, N.; Ranjan, S.; Umesh, P.; Suri, J.S. Automated classification of
  patients with coronary artery disease using grayscale features from left ventricle echocardiographic images. *Comput. Methods Programs Biomed.* 2013, 112, 624–632. [CrossRef] [PubMed]
- 31. Jena, B.; Nayak, G.K.; Saxena, S. Convolutional neural network and its pretrained models for image classification and object detection: A survey. *Concurr. Comput. Pr. Exp.* **2021**, *34*, e6767. [CrossRef]
- 32. Suri, J.S.; Biswas, M.; Kuppili, V.; Saba, L.; Edla, D.R.; Suri, H.S.; Cuadrado-Godia, E.; Laird, J.R.; Marinhoe, R.T.; Sanches, J.M.; et al. State-of-the-art review on deep learning in medical imaging. *Front. Biosci.* **2019**, 24, 392–426. [CrossRef]
- 33. Tandel, G.S.; Biswas, M.; Kakde, O.G.; Tiwari, A.; Suri, H.S.; Turk, M.; Laird, J.R.; Asare, C.K.; Ankrah, A.A.; Khanna, N.N.; et al. A Review on a Deep Learning Perspective in Brain Cancer Classification. *Cancers* **2019**, *11*, 111. [CrossRef] [PubMed]
- 34. Jain, P.K.; Sharma, N.; Saba, L.; Paraskevas, K.I.; Kalra, M.K.; Johri, A.; Laird, J.R.; Nicolaides, A.N.; Suri, J.S. Unseen Artificial Intelligence—Deep Learning Paradigm for Segmentation of Low Atherosclerotic Plaque in Carotid Ultrasound: A Multicenter Cardiovascular Study. *Diagnostics* **2021**, *11*, 2257. [CrossRef] [PubMed]
- 35. Jain, P.K.; Sharma, N.; Saba, L.; Paraskevas, K.I.; Kalra, M.K.; Johri, A.; Nicolaides, A.N.; Suri, J.S. Automated deep learning-based paradigm for high-risk plaque detection in B-mode common carotid ultrasound scans: An asymptomatic Japanese cohort study. *Int. Angiol.* **2022**, *41*, 9–23. [CrossRef]
- 36. Suri, J.S.; Agarwal, S.; Elavarthi, P.; Pathak, R.; Ketireddy, V.; Columbu, M.; Saba, L.; Gupta, S.K.; Faa, G.; Singh, I.M.; et al. Inter-Variability Study of COVLIAS 1.0: Hybrid Deep Learning Models for COVID-19 Lung Segmentation in Computed Tomography. *Diagnostics* 2021, 11, 2025. [CrossRef]
- 37. Suri, J.; Agarwal, S.; Pathak, R.; Ketireddy, V.; Columbu, M.; Saba, L.; Gupta, S.; Faa, G.; Singh, I.; Turk, M.; et al. COVLIAS 1.0: Lung Segmentation in COVID-19 Computed Tomography Scans Using Hybrid Deep Learning Artificial Intelligence Models. *Diagnostics* 2021, 11, 1405. [CrossRef]
- 38. Skandha, S.S.; Nicolaides, A.; Gupta, S.K.; Koppula, V.K.; Saba, L.; Johri, A.M.; Kalra, M.S.; Suri, J.S. A hybrid deep learning paradigm for carotid plaque tissue characterization and its validation in multicenter cohorts using a supercomputer framework. *Comput. Biol. Med.* **2021**, *141*, 105131. [CrossRef]
- 39. Saba, L.; Sanagala, S.S.; Gupta, S.K.; Koppula, V.K.; Johri, A.M.; Khanna, N.N.; Mavrogeni, S.; Laird, J.R.; Pareek, G.; Miner, M.; et al. Multimodality carotid plaque tissue characterization and classification in the artificial intelligence paradigm: A narrative review for stroke application. *Ann. Transl. Med.* **2021**, *9*, 1206. [CrossRef] [PubMed]
- 40. Pan, S.J.; Yang, Q. A survey on transfer learning. IEEE Trans. Knowl. Data Eng. 2009, 22, 1345–1359. [CrossRef]
- 41. Sanagala, S.S.; Nicolaides, A.; Gupta, S.K.; Koppula, V.K.; Saba, L.; Agarwal, S.; Johri, A.M.; Kalra, M.S.; Suri, J.S. Ten Fast Transfer Learning Models for Carotid Ultrasound Plaque Tissue Characterization in Augmentation Framework Embedded with Heatmaps for Stroke Risk Stratification. *Diagnostics* **2021**, *11*, 2109. [CrossRef] [PubMed]
- 42. Saba, L.; Sanagala, S.S.; Gupta, S.K.; Koppula, V.K.; Johri, A.M.; Sharma, A.M.; Kolluri, R.; Bhatt, D.L.; Nicolaides, A.; Suri, J.S. Ultrasound-based internal carotid artery plaque characterization using deep learning paradigm on a supercomputer: A cardiovascular disease/stroke risk assessment system. *Int. J. Cardiovasc. Imaging* **2021**, *37*, 1511–1528. [CrossRef]
- 43. Jena, B.; Saxena, S.; Nayak, G.K.; Saba, L.; Sharma, N.; Suri, J.S. Artificial intelligence-based hybrid deep learning models for image classification: The first narrative review. *Comput. Biol. Med.* **2021**, *137*, 104803. [CrossRef]
- 44. Saba, L.; Agarwal, M.; Patrick, A.; Puvvula, A.; Gupta, S.K.; Carriero, A.; Laird, J.R.; Kitas, G.D.; Johri, A.M.; Balestrieri, A.; et al. Six artificial intelligence paradigms for tissue characterisation and classification of non-COVID-19 pneumonia against COVID-19 pneumonia in computed tomography lungs. *Int. J. Comput. Assist. Radiol. Surg.* 2021, 16, 423–434. [CrossRef]

Cancers 2022, 14, 4052 33 of 37

45. Dong, M.; Lu, X.; Ma, Y.; Guo, Y.; Ma, Y.; Wang, K. An Efficient Approach for Automated Mass Segmentation and Classification in Mammograms. *J. Digit. Imaging* **2015**, *28*, 613–625. [CrossRef]

- 46. Weiss, W.A. Genetics of brain tumors. Curr. Opin. Pediatrics 2000, 12, 543–548. [CrossRef]
- 47. Soto, A.M.; Sonnenschein, C. Environmental causes of cancer: Endocrine disruptors as carcinogens. *Nat. Rev. Endocrinol.* **2010**, *6*, 363–370. [CrossRef]
- 48. Kazerooni, A.F.; Bagley, S.J.; Akbari, H.; Saxena, S.; Bagheri, S.; Guo, J.; Chawla, S.; Nabavizadeh, A.; Mohan, S.; Bakas, S.; et al. Applications of Radiomics and Radiogenomics in High-Grade Gliomas in the Era of Precision Medicine. *Cancers* **2021**, *13*, 5921. [CrossRef]
- 49. Habib, A.; Jovanovich, N.; Hoppe, M.; Ak, M.; Mamindla, P.; Colen, R.R.; Zinn, P. MRI-Based Radiomics and Radiogenomics in the Management of Low-Grade Gliomas: Evaluating the Evidence for a Paradigm Shift. *J. Clin. Med.* **2021**, *10*, 1411. [CrossRef] [PubMed]
- 50. Kocak, B.; Durmaz, E.S.; Ates, E.; Ulusan, M.B. Radiogenomics in Clear Cell Renal Cell Carcinoma: Machine Learning–Based High-Dimensional Quantitative CT Texture Analysis in Predicting *PBRM1* Mutation Status. *Am. J. Roentgenol.* **2019**, 212, W55–W63. [CrossRef] [PubMed]
- 51. Trivizakis, E.; Papadakis, G.Z.; Souglakos, I.; Papanikolaou, N.; Koumakis, L.; Spandidos, D.A.; Tsatsakis, A.; Karantanas, A.H.; Marias, K. Artificial intelligence radiogenomics for advancing precision and effectiveness in oncologic care (Review). *Int. J. Oncol.* **2020**, *57*, 43–53. [CrossRef]
- 52. Louis, D.N.; Perry, A.; Wesseling, P.; Brat, D.J.; Cree, I.A.; Figarella-Branger, D.; Hawkins, C.; Ng, H.K.; Pfister, S.M.; Reifenberger, G.; et al. The 2021 WHO Classification of Tumors of the Central Nervous System: A summary. *Neuro-Oncology* **2021**, *23*, 1231–1251. [CrossRef] [PubMed]
- 53. Akkus, Z.; Ali, I.; Sedlar, J.; Kline, T.L.; Agrawal, J.P.; Parney, I.F.; Giannini, C.; Erickson, B.J. Predicting 1p19q chromosomal deletion of low-grade gliomas from MR images using deep learning. *arXiv* **2016**, arXiv:06939.
- 54. Banerjee, I.; Crawley, A.; Bhethanabotla, M.; Daldrup-Link, H.E.; Rubin, D.L. Transfer learning on fused multiparametric MR images for classifying histopathological subtypes of rhabdomyosarcoma. *Comput. Med. Imaging Graph.* **2017**, *65*, 167–175. [CrossRef]
- 55. Iwatate, Y.; Hoshino, I.; Yokota, H.; Ishige, F.; Itami, M.; Mori, Y.; Chiba, S.; Arimitsu, H.; Yanagibashi, H.; Nagase, H.; et al. Radiogenomics for predicting p53 status, PD-L1 expression, and prognosis with machine learning in pancreatic cancer. *Br. J. Cancer* 2020, 123, 1253–1261. [CrossRef]
- 56. Park, E.K.; Lee, K.-S.; Seo, B.K.; Cho, K.R.; Woo, O.H.; Son, G.S.; Lee, H.Y.; Chang, Y.W. Machine Learning Approaches to Radiogenomics of Breast Cancer using Low-Dose Perfusion Computed Tomography: Predicting Prognostic Biomarkers and Molecular Subtypes. *Sci. Rep.* **2019**, *9*, 1–11. [CrossRef]
- 57. Saha, A.; Harowicz, M.R.; Grimm, L.; Kim, C.E.; Ghate, S.V.; Walsh, R.; Mazurowski, M.A. A machine learning approach to radiogenomics of breast cancer: A study of 922 subjects and 529 DCE-MRI features. *Br. J. Cancer* **2018**, *119*, 508–516. [CrossRef]
- 58. Yu, D.; Zhou, M.; Yang, F.; Dong, D.; Gevaert, O.; Liu, Z.; Shi, J.; Tian, J. Convolutional neural networks for predicting molecular profiles of non-small cell lung cancer. In Proceedings of the 2017 IEEE 14th International Symposium on Biomedical Imaging (ISBI 2017), Melbourne, VIC, Australia, 18–21 April 2017; pp. 569–572. [CrossRef]
- 59. Zhou, Z.; Chen, L.; Sher, D.; Zhang, Q.; Shah, J.; Pham, N.-L.; Jiang, S.; Wang, J. Predicting Lymph Node Metastasis in Head and Neck Cancer by Combining Many-objective Radiomics and 3-dimensioal Convolutional Neural Network through Evidential Reasoning. In Proceedings of the 2018 40th Annual International Conference of the IEEE Engineering in Medicine and Biology Society (EMBC), Honolulu, HI, USA, 18–21 July 2018. [CrossRef]
- 60. Chang, K.; Bai, H.X.; Zhou, H.; Su, C.; Bi, W.L.; Agbodza, E.; Kavouridis, V.K.; Senders, J.T.; Boaro, A.; Beers, A.; et al. Residual Convolutional Neural Network for the Determination of IDHStatus in Low- and High-Grade Gliomas from MR Imaging. *Clin. Cancer Res.* 2018, 24, 1073–1081. [CrossRef]
- 61. Chang, P.; Grinband, J.; Weinberg, B.D.; Bardis, M.; Khy, M.; Cadena, G.; Su, M.-Y.; Cha, S.; Filippi, C.G.; Bota, D.; et al. Deep-Learning Convolutional Neural Networks Accurately Classify Genetic Mutations in Gliomas. *Am. J. Neuroradiol.* **2018**, *39*, 1201–1207. [CrossRef] [PubMed]
- 62. Ha, R.; Mutasa, S.; Karcich, J.; Gupta, N.; Van Sant, E.P.; Nemer, J.; Sun, M.; Chang, P.; Liu, M.Z.; Jambawalikar, S. Predicting Breast Cancer Molecular Subtype with MRI Dataset Utilizing Convolutional Neural Network Algorithm. *J. Digit. Imaging* **2019**, 32, 276–282. [CrossRef] [PubMed]
- 63. Kawaguchi, R.; Takahashi, M.; Miyake, M.; Kinoshita, M.; Takahashi, S.; Ichimura, K.; Hamamoto, R.; Narita, Y.; Sese, J. Assessing Versatile Machine Learning Models for Glioma Radiogenomic Studies across Hospitals. *Cancers* **2021**, *13*, 3611. [CrossRef] [PubMed]
- 64. Kickingereder, P.; Bonekamp, D.; Nowosielski, M.; Kratz, A.; Sill, M.; Burth, S.; Wick, A.; Eidel, O.; Schlemmer, H.-P.; Radbruch, A.; et al. Radiogenomics of Glioblastoma: Machine Learning–based Classification of Molecular Characteristics by Using Multiparametric and Multiregional MR Imaging Features. *Radiology* 2016, 281, 907–918. [CrossRef]
- 65. Korfiatis, P.; Kline, T.L.; Lachance, D.H.; Parney, I.; Buckner, J.C.; Erickson, B.J. Residual Deep Convolutional Neural Network Predicts MGMT Methylation Status. *J. Digit. Imaging* **2017**, *30*, 622–628. [CrossRef]
- 66. Li, Z.; Wang, Y.; Yu, J.; Guo, Y.; Cao, W. Deep Learning based Radiomics (DLR) and its usage in noninvasive IDH1 prediction for low grade glioma. *Sci. Rep.* **2017**, *7*, 1–11. [CrossRef]

Cancers 2022, 14, 4052 34 of 37

67. Liang, S.; Zhang, R.; Liang, D.; Song, T.; Ai, T.; Xia, C.; Xia, L.; Wang, Y. Multimodal 3D DenseNet for IDH Genotype Prediction in Gliomas. *Genes* 2018, *9*, 382. [CrossRef]

- 68. Smedley, N.F.; Hsu, W. Using deep neural networks for radiogenomic analysis. In Proceedings of the 2018 IEEE 15th International Symposium on Biomedical Imaging (ISBI 2018), Washington, DC, USA, 4–7 April 2018; pp. 1529–1533.
- 69. Wang, S.; Shi, J.; Ye, Z.; Dong, D.; Yu, D.; Zhou, M.; Liu, Y.; Gevaert, O.; Wang, K.; Zhu, Y.; et al. Predicting EGFR mutation status in lung adenocarcinoma on computed tomography image using deep learning. *Eur. Respir. J.* **2019**, *53*, 1800986. [CrossRef]
- 70. Yoon, H.-J.; Ramanathan, A.; Alamudun, F.; Tourassi, G. Deep radiogenomics for predicting clinical phenotypes in invasive breast cancer. In Proceedings of the 14th International Workshop on Breast Imaging (IWBI 2018), Atlanta, GA, USA, 8–11 July 2018; Volume 10718. [CrossRef]
- 71. Zhu, Z.; Albadawy, E.; Saha, A.; Zhang, J.; Harowicz, M.R.; Mazurowski, M.A. Deep learning for identifying radiogenomic associations in breast cancer. *Comput. Biol. Med.* **2019**, *109*, 85–90. [CrossRef]
- 72. Andreassen, C.N.; Alsner, J.; Overgaard, J. Does variability in normal tissue reactions after radiotherapy have a genetic basis—where and how to look for it? *Radiother. Oncol.* **2002**, *64*, 131–140. [CrossRef]
- 73. The Cancer Genome Atlas Research Network. Comprehensive genomic characterization defines human glioblastoma genes and core pathways. *Nature* **2008**, 455, 1061–1068. [CrossRef] [PubMed]
- 74. Kalpathy-Cramer, J.; Freymann, J.B.; Kirby, J.; Kinahan, P.E.; Prior, F.W. Quantitative Imaging Network: Data Sharing and Competitive AlgorithmValidation Leveraging The Cancer Imaging Archive. *Transl. Oncol.* **2014**, *7*, 147–152. [CrossRef] [PubMed]
- 75. Bhattacharjee, K.; Pant, M. Hybrid particle swarm optimization-genetic algorithm trained multi-layer perceptron for classification of human glioma from molecular brain neoplasia data. *Cogn. Syst. Res.* **2019**, *58*, 173–194. [CrossRef]
- 76. Madhavan, S.; Zenklusen, J.-C.; Kotliarov, Y.; Sahni, H.; Fine, H.A.; Buetow, K. Rembrandt: Helping Personalized Medicine Become a Reality through Integrative Translational Research. *Mol. Cancer Res.* **2009**, *7*, 157–167. [CrossRef]
- 77. Gusev, Y.; Bhuvaneshwar, K.; Song, L.; Zenklusen, J.-C.; Fine, H.; Madhavan, S. The REMBRANDT study, a large collection of genomic data from brain cancer patients. *Sci. Data* **2018**, *5*, 180158. [CrossRef]
- 78. Gevaert, O.; Mitchell, L.A.; Achrol, A.S.; Xu, J.; Echegaray, S.; Steinberg, G.K.; Cheshier, S.H.; Napel, S.; Zaharchuk, G.; Plevritis, S.K. Glioblastoma Multiforme: Exploratory Radiogenomic Analysis by Using Quantitative Image Features. *Radiology* **2014**, 273, 168–174. [CrossRef]
- 79. Wiest, R.; Reyes, M.; Meier, R.; Gutman, D.A.; Bauer, S.; Alexander, B.; Velazquez, E.R.; Dunn, W.D.; Aerts, H.J.W.L. Fully automatic GBM segmentation in the TCGA-GBM dataset: Prognosis and correlation with VASARI features. *Sci. Rep.* **2015**, 5, 16822. [CrossRef]
- 80. Clark, K.; Vendt, B.; Smith, K.; Freymann, J.; Kirby, J.; Koppel, P.; Moore, S.; Phillips, S.; Maffitt, D.; Pringle, M.; et al. The Cancer Imaging Archive (TCIA): Maintaining and Operating a Public Information Repository. *J. Digit. Imaging* **2013**, 26, 1045–1057. [CrossRef]
- 81. Goldman, M.J.; Craft, B.; Hastie, M.; Repečka, K.; McDade, F.; Kamath, A.; Banerjee, A.; Luo, Y.; Rogers, D.; Brooks, A.N.; et al. Visualizing and interpreting cancer genomics data via the Xena platform. *Nat. Biotechnol.* **2020**, *38*, 675–678. [CrossRef]
- 82. Saxena, S.; Kazerooni, A.F.; Toorens, E.; Bakas, S.; Akbari, H.; Sako, C.; Mamourian, E.; Koumenis, C.; Mohan, S.; Bagley, S.J.; et al. NIMG-73. CAPTURING GLIOBLASTOMA HETEROGENEITY USING IMAGING AND DEEP LEARNING: APPLICATION TO MGMT PROMOTER METHYLATION. *Neuro-Oncology* **2021**, 23, vi146. [CrossRef]
- 83. Becnel, L.B.; Pereira, S.; Drummond, J.A.; Gingras, M.-C.; Covington, K.R.; Kovar, C.L.; Doddapaneni, H.V.; Hu, J.; Muzny, N.; McGuire, A.L.; et al. An open access pilot freely sharing cancer genomic data from participants in Texas. *Sci. Data* **2016**, *3*, 160010. [CrossRef] [PubMed]
- 84. Milius, D.; Dove, E.S.; Chalmers, D.; Dyke, S.O.M.; Kato, K.; Nicolás, P.; Ouellette, B.F.; Ozenberger, B.; Rodriguez, L.L.; Zeps, N.; et al. The International Cancer Genome Consortium's evolving data-protection policies. *Nat. Biotechnol.* **2014**, 32, 519–523. [CrossRef] [PubMed]
- 85. Jäkel, S.; Dimou, L. Glial Cells and Their Function in the Adult Brain: A Journey through the History of Their Ablation. *Front. Cell. Neurosci.* **2017**, *11*, 24. [CrossRef]
- 86. Jessen, K.R. Glial cells. Int. J. Biochem. Cell Biol. 2004, 36, 1861–1867. [CrossRef]
- Hanani, M.; Spray, D.C. Emerging importance of satellite glia in nervous system function and dysfunction. Nat. Rev. Neurosci. 2020, 21, 485–498. [CrossRef]
- 88. Glia Cell Pictotial Representation. Available online: https://courses.lumenlearning.com/wm-biology2/chapter/glial-cells/(accessed on 14 July 2022).
- 89. Yan, H.; Parsons, D.W.; Jin, G.; McLendon, R.; Rasheed, B.A.; Yuan, W.; Kos, I.; Batinic-Haberle, I.; Jones, S.; Riggins, G.J.; et al. *IDH1* and *IDH2* Mutations in Gliomas. *N. Engl. J. Med.* **2009**, *360*, 765–773. [CrossRef] [PubMed]
- 90. Mabray, M.C.; Barajas, R.F.; Cha, S. Modern Brain Tumor Imaging. Brain Tumor Res. Treat. 2015, 3, 8–23. [CrossRef]
- 91. Suh, C.H.; Kim, H.S.; Jung, S.C.; Choi, C.G.; Kim, S.J. Imaging prediction of isocitrate dehydrogenase (IDH) mutation in patients with glioma: A systemic review and meta-analysis. *Eur. Radiol.* **2018**, 29, 745–758. [CrossRef]
- 92. Mack, S.C.; Hubert, C.G.; Miller, T.E.; Taylor, M.; Rich, J.N. An epigenetic gateway to brain tumor cell identity. *Nat. Neurosci.* **2015**, 19, 10–19. [CrossRef]
- 93. Tommasini-Ghelfi, S.; Murnan, K.; Kouri, F.M.; Mahajan, A.S.; May, J.L.; Stegh, A.H. Cancer-associated mutation and beyond: The emerging biology of isocitrate dehydrogenases in human disease. *Sci. Adv.* **2019**, *5*, eaaw4543. [CrossRef]

Cancers 2022, 14, 4052 35 of 37

94. Hu, N.; Richards, R.; Jensen, R. Role of chromosomal 1p/19q co-deletion on the prognosis of oligodendrogliomas: A systematic review and meta-analysis. *Interdiscip. Neurosurg.* **2016**, *5*, 58–63. [CrossRef]

- 95. Han, Y.; Xie, Z.; Zang, Y.; Zhang, S.; Gu, D.; Zhou, M.; Gevaert, O.; Wei, J.; Li, C.; Chen, H.; et al. Non-invasive genotype prediction of chromosome 1p/19q co-deletion by development and validation of an MRI-based radiomics signature in lower-grade gliomas. *J. Neuro-Oncology* **2018**, *140*, 297–306. [CrossRef] [PubMed]
- 96. Gupta, K.; Salunke, P. Molecular markers of glioma: An update on recent progress and perspectives. *J. Cancer Res. Clin. Oncol.* **2012**, 138, 1971–1981. [CrossRef]
- 97. Cabrini, G.; Fabbri, E.; Lo Nigro, C.; Dechecchi, M.C.; Gambari, R. Regulation of expression of O6-methylguanine-DNA methyltransferase and the treatment of glioblastoma (Review). *Int. J. Oncol.* **2015**, 47, 417–428. [CrossRef] [PubMed]
- 98. Silber, J.R.; Bobola, M.S.; Blank, A.; Chamberlain, M.C. O6-Methylguanine-DNA methyltransferase in glioma therapy: Promise and problems. *Biochim. Biophys. Acta* **2012**, *1826*, 71–82. [CrossRef] [PubMed]
- 99. González-González, R.; López-Verdín, S.; Lavalle-Carrasco, J.; Molina-Frechero, N.; Isiordia-Espinoza, M.; Carreón-Burciaga, R.G.; Bologna-Molina, R. Current concepts in ameloblastoma-targeted therapies in B-raf proto-oncogene serine/threonine kinase V600E mutation: Systematic review. *World J. Clin. Oncol.* 2020, 11, 31–42. [CrossRef] [PubMed]
- 100. Maraka, S.; Janku, F. BRAF alterations in primary brain tumors. Discov. Med. 2018, 26, 51-60.
- 101. Werner, J.-M.; Brunn, A.; Fink, G.R.; Galldiks, N. Magnetic Resonance Imaging Reveals a Pronounced Treatment Response of a Isocitrate Dehydrogenase— and B-Raf Proto-Oncogene—Wildtype Epithelioid Glioblastoma. *World Neurosurg.* 2019, 127, 213–215. [CrossRef]
- 102. Zhang, Y.; Dube, C.; Gibert, M.; Cruickshanks, N.; Wang, B.; Coughlan, M.; Yang, Y.; Setiady, I.; Deveau, C.; Saoud, K.; et al. The p53 Pathway in Glioblastoma. *Cancers* **2018**, *10*, 297. [CrossRef]
- 103. Goldhoff, P.; Clarke, J.; Smirnov, I.; Berger, M.S.; Prados, M.D.; James, C.D.; Perry, A.; Phillips, J.J. Clinical Stratification of Glioblastoma Based on Alterations in Retinoblastoma Tumor Suppressor Protein (RB1) and Association With the Proneural Subtype. J. Neuropathol. Exp. Neurol. 2012, 71, 83–89. [CrossRef]
- 104. Xu, H.; Zong, H.; Ma, C.; Ming, X.; Shang, M.; Li, K.; He, X.; Du, H.; Cao, L. Epidermal growth factor receptor in glioblastoma. *Oncol. Lett.* **2017**, *14*, 512–516. [CrossRef] [PubMed]
- 105. Karsy, M.; Neil, J.A.; Guan, J.; Mahan, M.A.; Colman, H.; Jensen, R.L. A practical review of prognostic correlations of molecular biomarkers in glioblastoma. *Neurosurg. Focus* **2015**, *38*, E4. [CrossRef] [PubMed]
- 106. Hill, J.R.; Kuriyama, N.; Kuriyama, H.; Israel, M.A. Molecular genetics of brain tumors. Arch. Neurol. 1999, 56, 439-441. [CrossRef]
- 107. Wechsler-Reya, R.; Scott, M.P. The Developmental Biology of Brain Tumors. Annu. Rev. Neurosci. 2001, 24, 385–428. [CrossRef]
- 108. Yao, Y.; Mack, S.C.; Taylor, M.D. Molecular genetics of ependymoma. Chin. J. Cancer 2011, 30, 669–681. [CrossRef] [PubMed]
- 109. Vlaardingerbroek, M.T.; Boer, J.A. Magnetic Resonance Imaging: Theory and Practice; Springer Science & Business Media: Berlin/Heidelberg, Germany, 2013.
- 110. MRI Basics. Available online: https://case.edu/med/neurology/NR/MRI%20Basics.htm (accessed on 14 July 2022).
- 111. Edelman, R.R. The History of MR Imaging as Seen through the Pages of *Radiology*. *Radiology* **2014**, 273, S181–S200. [CrossRef] [PubMed]
- 112. Corrias, G.; Micheletti, G.; Barberini, L.; Suri, J.S.; Saba, L. Texture analysis imaging "what a clinical radiologist needs to know". *Eur. J. Radiol.* **2021**, *146*, 110055. [CrossRef]
- 113. Wu, G.; Jochems, A.; Refaee, T.; Ibrahim, A.; Yan, C.; Sanduleanu, S.; Woodruff, H.C.; Lambin, P. Structural and functional radiomics for lung cancer. *Eur. J. Pediatr.* **2021**, *48*, 3961–3974. [CrossRef]
- 114. Rizzo, S.; Botta, F.; Raimondi, S.; Origgi, D.; Fanciullo, C.; Morganti, A.G.; Bellomi, M. Radiomics: The facts and the challenges of image analysis. *Eur. Radiol. Exp.* **2018**, *2*, 36. [CrossRef]
- 115. Murgia, A.; Balestrieri, A.; Crivelli, P.; Suri, J.S.; Conti, M.; Cademartiri, F.; Saba, L. Cardiac computed tomography radiomics: An emerging tool for the non-invasive assessment of coronary atherosclerosis. *Cardiovasc. Diagn. Ther.* **2020**, *10*, 2005–2017. [CrossRef]
- 116. Srivastava, S.K.; Singh, S.K.; Suri, J.S. Effect of incremental feature enrichment on healthcare text classification system: A machine learning paradigm. *Comput. Methods Programs Biomed.* **2019**, 172, 35–51. [CrossRef] [PubMed]
- 117. Torigian, E.A.; Huang, S.S.; Houseni, M.; Alavi, A. Functional imaging of cancer with emphasis on molecular techniques. *CA Cancer J. Clin.* **2007**, 57, 206–224. [CrossRef] [PubMed]
- 118. Cau, R.; Bassareo, P.; Suri, J.S.; Saba, L. Reply to "Structural And/Or Functional Underpinnings of Magnetic Resonance Imaging Bi-Atrial Strain Impairment in Patients With Takotsubo Syndrome". Can. Assoc. Radiol. J. 2022, 73, 599. [CrossRef] [PubMed]
- 119. Porcu, M.; Cocco, L.; Saloner, D.; Suri, J.S.; Montisci, R.; Carriero, A.; Saba, L. Extracranial Carotid Artery Stenosis: The Effects on Brain and Cognition with a Focus on Resting-State Functional Connectivity. *J. Neuroimaging* **2020**, *30*, 736–745. [CrossRef]
- 120. Wu, D.H.; Chen, Z.; North, J.C.; Biswas, M.; Vo, J.; Suri, J.S. Machine learning paradigm for dynamic contrast-enhanced MRI evaluation of expanding bladder. *Front. Biosci.* **2020**, *25*, 1746–1764. [CrossRef]
- 121. Banchhor, S.K.; Londhe, N.D.; Araki, T.; Saba, L.; Radeva, P.; Laird, J.R.; Suri, J.S. Wall-based measurement features provides an improved IVUS coronary artery risk assessment when fused with plaque texture-based features during machine learning paradigm. *Comput. Biol. Med.* 2017, 91, 198–212. [CrossRef]

Cancers 2022, 14, 4052 36 of 37

122. Than, J.C.; Saba, L.; Noor, N.M.; Rijal, O.M.; Kassim, R.M.; Yunus, A.; Suri, H.S.; Porcu, M.; Suri, J.S. Lung disease stratification using amalgamation of Riesz and Gabor transforms in machine learning framework. *Comput. Biol. Med.* 2017, 89, 197–211. [CrossRef]

- 123. Araki, T.; Jain, P.K.; Suri, H.S.; Londhe, N.D.; Ikeda, N.; El-Baz, A.; Shrivastava, V.K.; Saba, L.; Nicolaides, A.; Shafique, S.; et al. Stroke Risk Stratification and its Validation using Ultrasonic Echolucent Carotid Wall Plaque Morphology: A Machine Learning Paradigm. *Comput. Biol. Med.* 2017, 80, 77–96. [CrossRef] [PubMed]
- 124. Negreros-Osuna, A.A.; Parakh, A.; Corcoran, R.B.; Pourvaziri, A.; Kambadakone, A.; Ryan, D.P.; Sahani, D.V. Radiomics Texture Features in Advanced Colorectal Cancer: Correlation with *BRAF* Mutation and 5-year Overall Survival. *Radiol. Imaging Cancer* **2020**, *2*, e190084. [CrossRef] [PubMed]
- 125. Alzubaidi, L.; Duan, Y.; Al-Dujaili, A.; Ibraheem, I.K.; Alkenani, A.H.; Santamaría, J.; Fadhel, M.A.; Al-Shamma, O.; Zhang, J. Deepening into the suitability of using pre-trained models of ImageNet against a lightweight convolutional neural network in medical imaging: An experimental study. *PeerJ Comput. Sci.* 2021, 7, e715. [CrossRef]
- 126. Sudeep, P.; Palanisamy, P.; Rajan, J.; Baradaran, H.; Saba, L.; Gupta, A.; Suri, J.S. Speckle reduction in medical ultrasound images using an unbiased non-local means method. *Biomed. Signal Process. Control* **2016**, *28*, 1–8. [CrossRef]
- 127. Fei, B.; Zhang, S.; Savado, O.; Suri, J.; Lewin, J.S.; Wilson, D.L. Three-dimensional automatic volume registration of carotid MR images. In Proceedings of the 25th Annual International Conference of the IEEE Engineering in Medicine and Biology Society (IEEE Cat. No.03CH37439), Cancun, Mexico, 17–21 September 2003. [CrossRef]
- 128. Guo, Y.; Sivaramakrishna, R.; Lu, C.-C.; Suri, J.S.; Laxminarayan, S. Breast image registration techniques: A survey. *Med Biol. Eng. Comput.* **2006**, *44*, 15–26. [CrossRef] [PubMed]
- 129. Saxena, S.; Jena, B.; Gupta, N.; Das, S.; Sarmah, D.; Bhattacharya, P.; Nath, T.; Paul, S.; Fouda, M.M.; Kalra, M.; et al. Role of Artificial Intelligence in Radiogenomics for Cancers in the Era of Precision Medicine. *Cancers* 2022, 14, 2860. [CrossRef] [PubMed]
- 130. Kazerooni, A.F.; Saxena, S.; Toorens, E.; Tu, D.; Bashyam, V.; Akbari, H.; Mamourian, E.; Sako, C.; Koumenis, C.; Verginadis, I.; et al. Clinical measures, radiomics, and genomics offer synergistic value in AI-based prediction of overall survival in patients with glioblastoma. *Sci. Rep.* 2022, *12*, 1–13. [CrossRef] [PubMed]
- 131. Calabrese, E.; Villanueva-Meyer, J.E.; Cha, S. A fully automated artificial intelligence method for non-invasive, imaging-based identification of genetic alterations in glioblastomas. *Sci. Rep.* **2020**, *10*, 1–11. [CrossRef]
- 132. Kumar, P.K.; Darshan, P.; Kumar, S.; Ravindra, R.; Rajan, J.; Saba, L.; Suri, J.S. Magnetic resonance image denoising using nonlocal maximum likelihood paradigm in DCT-framework. *Int. J. Imaging Syst. Technol.* **2015**, 25, 256–264. [CrossRef]
- 133. Skandha, S.S.; Gupta, S.K.; Saba, L.; Koppula, V.K.; Johri, A.M.; Khanna, N.N.; Mavrogeni, S.; Laird, J.R.; Pareek, G.; Miner, M.; et al. 3-D optimized classification and characterization artificial intelligence paradigm for cardiovascular/stroke risk stratification using carotid ultrasound-based delineated plaque: Atheromatic<sup>TM</sup> 2.0. *Comput. Biol. Med.* **2020**, *125*, 103958. [CrossRef]
- 134. Agarwal, M.; Saba, L.; Gupta, S.K.; Johri, A.M.; Khanna, N.N.; Mavrogeni, S.; Laird, J.R.; Pareek, G.; Miner, M.; Sfikakis, P.P.; et al. Wilson disease tissue classification and characterization using seven artificial intelligence models embedded with 3D optimization paradigm on a weak training brain magnetic resonance imaging datasets: A supercomputer application. *Med Biol. Eng. Comput.* 2021, 59, 511–533. [CrossRef]
- 135. Kazerooni, A.F.; Bakas, S.; Rad, H.S.; Davatzikos, C. Imaging signatures of glioblastoma molecular characteristics: A radiogenomics review. *J. Magn. Reson. Imaging* **2019**, *52*, 54–69. [CrossRef]
- 136. Alberch, P. From genes to phenotype: Dynamical systems and evolvability. Genetica 1991, 84, 5–11. [CrossRef]
- 137. Ahnert, S.E. Structural properties of genotype–phenotype maps. J. R. Soc. Interface 2017, 14, 20170275. [CrossRef] [PubMed]
- 138. Aguilar-Rodríguez, J.; Peel, L.; Stella, M.; Wagner, A.; Payne, J.L. The architecture of an empirical genotype-phenotype map. *Evolution* **2018**, 72, 1242–1260. [CrossRef] [PubMed]
- 139. Gullo, R.L.; Daimiel, I.; Morris, E.A.; Pinker, K. Combining molecular and imaging metrics in cancer: Radiogenomics. *Insights Into Imaging* **2020**, *11*, 1–17. [CrossRef] [PubMed]
- 140. Jena, B.; Nayak, G.K.; Paul, S.; Saxena, S. An Exhaustive Analytical Study of U-Net Architecture on Two Diverse Biomedical Imaging Datasets of Electron Microscopy Drosophila ssTEM and Brain MRI BraTS-2021 for Segmentation. SN Comput. Sci. 2022, 3, 1–13. [CrossRef]
- 141. Milletari, F.; Navab, N.; Ahmadi, S.-A. V-net: Fully convolutional neural networks for volumetric medical image segmentation. In Proceedings of the 2016 Fourth International Conference on 3D Vision (3DV), Stanford, CA, USA, 25–28 October 2016; pp. 565–571.
- 142. Jena, B.; Thakar, P.; Nayak, V.; Nayak, G.K.; Saxena, S. Malaria Parasites Detection Using Deep Neural Network. In *Deep Learning Applications in Medical Imaging*; IGI Global: Hershey, PA, USA, 2021; pp. 209–222. [CrossRef]
- 143. Jena, B.; Nayak, G.K.; Saxena, S. Comprehensive Review of Abdominal Image Segmentation using Soft and Hard Computing Approaches. In Proceedings of the 2020 International Conference on Computer Science, Engineering and Applications (ICCSEA), Gunupur, India, 13–14 March 2020; pp. 1–5. [CrossRef]
- 144. Jena, B.; Nayak, G.K.; Saxena, S. High-Performance Computing and Its Requirements in Deep Learning. In *High-Performance Medical Image Processing*; Apple Academic Press: Palm Bay, FL, USA, 2022; pp. 255–288. [CrossRef]
- 145. Jena, B.; Nayak, G.K.; Saxena, S. Maximum Payload for Digital Image Steganography Obtained by Mixed Edge Detection Mechanism. In Proceedings of the 2019 International Conference on Information Technology (ICIT), Bhubaneswar, India, 19–21 December 2019; pp. 206–210. [CrossRef]

Cancers 2022, 14, 4052 37 of 37

146. Bodalal, Z.; Trebeschi, S.; Nguyen-Kim, T.D.L.; Schats, W.; Beets-Tan, R. Radiogenomics: Bridging imaging and genomics. *Abdom. Radiol.* **2019**, *44*, 1960–1984. [CrossRef] [PubMed]

- 147. Shui, L.; Ren, H.; Yang, X.; Li, J.; Chen, Z.; Yi, C.; Zhu, H.; Shui, P. Era of radiogenomics in precision medicine: An emerging approach for prediction of the diagnosis, treatment and prognosis of tumors. *Front. Oncol.* **2020**, *10*, 3195.
- 148. Singh, G.; Manjila, S.; Sakla, N.; True, A.; Wardeh, A.H.; Beig, N.; Vaysberg, A.; Matthews, J.; Prasanna, P.; Spektor, V. Radiomics and radiogenomics in gliomas: A contemporary update. *Br. J. Cancer* 2021, 125, 641–657. [CrossRef]
- 149. Agarwal, M.; Agarwal, S.; Saba, L.; Chabert, G.L.; Gupta, S.; Carriero, A.; Pasche, A.; Danna, P.; Mehmedovic, A.; Faa, G.; et al. Eight pruning deep learning models for low storage and high-speed COVID-19 computed tomography lung segmentation and heatmap-based lesion localization: A multicenter study using COVLIAS 2.0. *Comput. Biol. Med.* 2022, 146, 105571. [CrossRef]
- 150. Suri, J.S.; Agarwal, S.; Chabert, G.L.; Carriero, A.; Paschè, A.; Danna, P.S.C.; Saba, L.; Mehmedović, A.; Faa, G.; Singh, I.M.; et al. COVLIAS 2.0-cXAI: Cloud-Based Explainable Deep Learning System for COVID-19 Lesion Localization in Computed Tomography Scans. *Diagnostics* 2022, *12*, 1482. [CrossRef]
- 151. Hoffman, R.R.; Mueller, S.T.; Klein, G.; Litman, J. Metrics for explainable AI: Challenges and prospects. arXiv 2018, arXiv:04608.
- 152. Suri, J.S.; Agarwal, S.; Carriero, A.; Paschè, A.; Danna, P.S.C.; Columbu, M.; Saba, L.; Viskovic, K.; Mehmedović, A.; Agarwal, S.; et al. COVLIAS 1.0 vs. MedSeg: Artificial Intelligence-Based Comparative Study for Automated COVID-19 Computed Tomography Lung Segmentation in Italian and Croatian Cohorts. *Diagnostics* 2021, 11, 2367. [CrossRef] [PubMed]
- 153. Suri, J.S.; Agarwal, S.; Gupta, S.K.; Puvvula, A.; Viskovic, K.; Suri, N.; Alizad, A.; El-Baz, A.; Saba, L.; Fatemi, M.; et al. Systematic Review of Artificial Intelligence in Acute Respiratory Distress Syndrome for COVID-19 Lung Patients: A Biomedical Imaging Perspective. *IEEE J. Biomed. Heal. Inform.* 2021, 25, 4128–4139. [CrossRef] [PubMed]
- 154. Banchhor, S.K.; Londhe, N.D.; Araki, T.; Saba, L.; Radeva, P.; Khanna, N.N.; Suri, J.S. Calcium detection, its quantification, and grayscale morphology-based risk stratification using machine learning in multimodality big data coronary and carotid scans: A review. *Comput. Biol. Med.* **2018**, *101*, 184–198. [CrossRef] [PubMed]
- 155. Holzinger, A.; Langs, G.; Denk, H.; Zatloukal, K.; Müller, H. Causability and explainability of artificial intelligence in medicine. *WIREs Data Min. Knowl. Discov.* **2019**, *9*, e1312. [CrossRef] [PubMed]
- 156. Castro, D.C.; Walker, I.; Glocker, B. Causality matters in medical imaging. Nat. Commun. 2020, 11, 1–10. [CrossRef] [PubMed]
- 157. Chou, Y.L.; Moreira, C.; Bruza, P.; Ouyang, C.; Jorge, J. Counterfactuals and causability in explainable artificial intelligence: Theory, algorithms, and applications. *Inf. Fusion* **2022**, *81*, 59–83. [CrossRef]
- 158. Saba, L.; Banchhor, S.K.; Londhe, N.D.; Araki, T.; Laird, J.R.; Gupta, A.; Nicolaides, A.; Suri, J.S. Web-based accurate measurements of carotid lumen diameter and stenosis severity: An ultrasound-based clinical tool for stroke risk assessment during multicenter clinical trials. *Comput. Biol. Med.* 2017, 91, 306–317. [CrossRef]
- 159. El-Baz, A.; Suri, J.S. Big Data in Multimodal Medical Imaging; CRC Press: Boca Raton, FL, USA, 2019.
- 160. Mendes, A.C.; Ribeiro, A.S.; Oros-Peusquens, A.M.; Langen, K.J.; Shah, J.; Ferreira, H.A. Brain connectivity study of brain tumor patients using MR-PET data: Preliminary results. *EJNMMI Phys.* **2015**, 2, 1–2. [CrossRef]
- 161. Porcu, M.; Operamolla, A.; Scapin, E.; Garofalo, P.; Destro, F.; Caneglias, A.; Suri, J.S.; Falini, A.; DeFazio, G.; Marrosu, F.; et al. Effects of White Matter Hyperintensities on Brain Connectivity and Hippocampal Volume in Healthy Subjects According to Their Localization. *Brain Connect.* 2020, 10, 436–447. [CrossRef]
- 162. Saba, L.; Sanfilippo, R.; Sannia, S.; Anzidei, M.; Montisci, R.; Mallarini, G.; Suri, J.S. Association Between Carotid Artery Plaque Volume, Composition, and Ulceration: A Retrospective Assessment With MDCT. *Am. J. Roentgenol.* **2012**, *199*, 151–156. [CrossRef]
- 163. Suri, J.S.; Puvvula, A.; Biswas, M.; Majhail, M.; Saba, L.; Faa, G.; Singh, I.M.; Oberleitner, R.; Turk, M.; Chadha, P.S.; et al. COVID-19 pathways for brain and heart injury in comorbidity patients: A role of medical imaging and artificial intelligence-based COVID severity classification: A review. *Comput. Biol. Med.* 2020, 124, 103960. [CrossRef] [PubMed]
- 164. Holdsworth, S.J.; Bammer, R. Magnetic resonance imaging techniques: fMRI, DWI, and PWI. In *Seminars in Neurology*; Thieme Medical Publishers: New York, NY, USA, 2008; Volume 28, pp. 395–406.
- 165. Öz, G.; Alger, J.R.; Barker, P.B.; Bartha, R.; Bizzi, A.; Boesch, C.; Bolan, P.J.; Brindle, K.M.; Cudalbu, C.; Dinçer, A.; et al. Clinical Proton MR Spectroscopy in Central Nervous System Disorders. *Radiology* **2014**, 270, 658–679. [CrossRef] [PubMed]
- 166. Sourbron, S. Technical aspects of MR perfusion. Eur. J. Radiol. 2010, 76, 304–313. [CrossRef]
- 167. Jeurissen, B.; Descoteaux, M.; Mori, S.; Leemans, A. Diffusion MRI fiber tractography of the brain. *NMR Biomed.* **2017**, 32, e3785. [CrossRef]
- 168. Singleton, M.J. Functional Magnetic Resonance Imaging. Yale J. Biol. Med. 2009, 82, 233.
- 169. Buzug, T.M. Computed tomography. In *Springer Handbook of Medical Technology*; Springer: Berlin/Heidelberg, Germany, 2011; pp. 311–342.
- 170. Bailey, D.L.; Maisey, M.N.; Townsend, D.W.; Valk, P.E. Positron Emission Tomography; Springer: Berlin/Heidelberg, Germany, 2005.
- 171. Holly, T.A. Single Photon-Emission Computed Tomography; Springer: Berlin/Heidelberg, Germany, 2010.
- 172. Catafau, A.M. Brain SPECT in clinical practice. Part I: Perfusion. J. Nucl. Med. 2001, 42, 259–271. [PubMed]