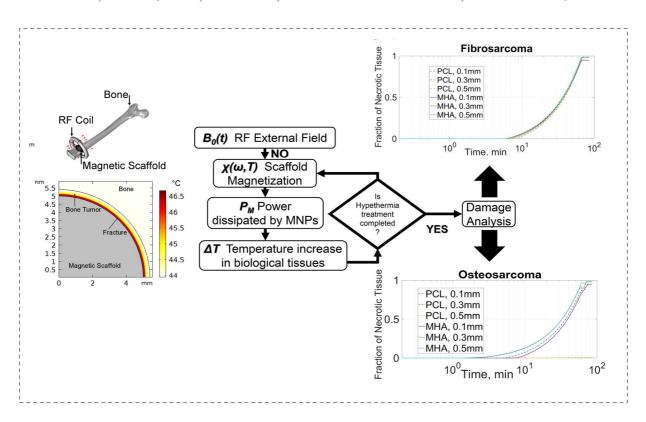
NUMERICAL INVESTIGATION OF BONE TUMOR HYPERTHERMIA TREATMENT USING MAGNETIC SCAFFOLDS

A. Fanti, Member, IEEE, M. B. Lodi, G. Vacca and G. Mazzarella, Senior Member, IEEE



Modelling and in silico experiments of magnetic scaffolds for in situ hyperthermia treatment against Fibrosarcoma and Osteosarcoma tumors.

Take-Home Messages

- Functionalization of bone scaffolds using magnetic nanoparticles allows hyperthermia of bone tumors in an effective way.
- The possibility of employing innovative magnetic scaffolds as therapeutic tool in orthopaedic oncology is analyzed via numerical simulations. Using a Cole-Cole model, non-linear material properties are evaluated to define external field parameter to perform an effective treatment for bone tumors such as Fibrosarcoma and Osteosarcomas.
- Accurate electromagnetic and thermal modeling of scaffolds and nanoparticles, in the whole range of involved temperature, is required to design effective and safe treatments.
- Different tumoral tissues and qualitative features such as the presence, size and type of surgical fracture, affect in a significant way the hyperthermia treatment.

Numerical Investigation of Bone Tumor Hyperthermia Treatment using Magnetic Scaffolds

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Abstract This works claims to define, via numerical simulations, magnetic field parameters to perform an effective in situ bone tumor hyperthermia treatment using magnetic scaffolds. A Cole-Cole model to describe the frequency response of the magnetic susceptibility of nanoparticles embedded in novel magnetic biomaterials is explored. The heating phenomena is investigated considering both the ischemic and inflamed state of the fracture gap at the bone/implants interface. Both Osteosarcoma and Fibrosarcoma tumors are analyzed. Magnetic hydroxyapatite and poly-ɛ-caprolactone scaffolds are investigated. From the thermal analysis, it is found that the fracture behaves as a resistance to heat conduction, therefore strength and frequency of external magnetic field has to be tuned to perform the treatment taking the fracture status into account. Moreover, numerical experiments indicate that low perfused Fibrosarcoma can be treated using moderate-strength field, whereas more intense external fields are required to treat strongly vascularized Osteosarcoma without damaging healthy bone tissue. Magnetic hydroxyapatite stands out to be the most performant and versatile material to treat both tumors. These simulations can be regarded as a starting point to analyze possible clinical use of magnetic scaffolds for in situ bone hyperthermia.

Keywords — Biomagnetics, Electromagnetic fields, Heat treatment, Hyperthermia, Magnetic particles, Tumors.

I. INTRODUCTION

ONE cancers are a wide class of relatively rare B neoplasms, mostly afflicting patients between the ages of 10 and 20 years [1], with an incidence of about 3 cases out of 100'000 persons per year [1]-[3]. Bone neoplasms can be classified in benign, intermediate or malignant. Moreover, since tumors can originate from tissues different from bone, it is necessary to distinguish between boneforming cancers (Osteosarcomas, i.e. OS), cartilage-(Chondrosarcomas), bone-marrow sarcoma), and soft-tissue ones (Fibrosarcoma, i.e. FS) [2]. OS are aggressive malignant bone tumors whose cells produce bone matrix or osteoid [1]-[3]. Furthermore, OS can reach 5 cm and grow in an angiocentric fashion, implying a noticeable vascularization [1]-[3]. Prognostic factors depend on treatments, though survival rate ranges from 30% to 40% [1]. On the other hand, FS is a malignant neoplasm arranged in a fascicular architecture of interlaced trabecular bundles. It tends to extend to soft tissues [1]. A 10-year survival rate equal to 28% for this kind of neoplasm has been reported in [4]. Both OS and FS mostly affect long bones, namely distal femur, proximal or upper part tibia,

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and humerus [1], [2], [4]. Therefore, the distinct clinical features, together with the pathological and histological characteristics of the cited lesions, determine different staging and treatment. Currently, the main treatment strategies are chemotherapy, radiotherapy, osteotomies, ablative surgery or intralesional excision (i.e. curettage) [1], [3]. However, approximately 15% of surgical intervention results in tumor recurrence [3]. To overcome this clinical issue, several novel techniques aimed to kill residual cancer cells have been proposed, e.g. combined radiotherapy and chemotherapy protocols, hyperthermia, immunotherapy and smart drug nanocarriers [5]-[9].

Hyperthermia (HT) stands out to be a very promising approach [5], [9], [10]. Its thermo-biological rationale is to selectively rise tumor temperature above 42.5 °C for a time sufficient to impair transmembranal transport, inducing cytotoxicity determining cancer cells death and eliciting immune system response [9], [10]. Moreover, temperature rise increases cells permeability to antitumor drugs, allowing a synergic effect with chemotherapy [9], [10]. Hyperthermia can be induced by ultrasounds or Electromagnetic (EM) fields exposure [9]. Fan et al. developed a surgical method that exploited EM hyperthermia [11]. In their study, 62 patients with various bone tumors (among them 30 OS, 4 FS) have been treated, in situ, with an antenna array while monitoring tumor temperature to keep it above 65 °C. An 8% recurrence rate, infections and six deaths were reported. In [11] it is stressed the clinical need for a proper management of the

pathological fractures after surgery, such as grafts or the use of bone scaffolds to withstand tissue regeneration. This need has led to the development of novel magnetic biomaterials to be used both as scaffolds for tissue support after curettage, and as heat source for local magnetic induction HT treatment of bone tumor [12]-[14]. Following surgery, after magnetic scaffold placement, therapeutic heat is generated and conducted to surrounding tissues by remotely applying an alternate magnetic field (MF) through an external coil [13]-[15]. In this way, temperature can increase above 43°C, thus killing residual cancer cells and allowing healthy tissue to grow sustained by the implant [13], [14]. A very first attempt was the ferromagnetic bioglass intermedullary pin proposed by Ikenaga et al. [15]. The device was tested in vivo against VX2 tumor cells. A temperature raise of 13°C was obtained after 50 min, leading to a decrease in fracture rate [15]. After that, different kinds of ferro- [15], [16] and ferrimagnetic glass [16], [17] ceramics have been synthetized and characterized to ensure the use of these biomaterials for HT purposes [16]-[19].

Further interesting and innovative scaffolds biomaterials, able to perform in situ HT of bone tumor, are based on superparamagnetic iron oxide nanoparticles (SPIONs) [20]. Blocking temperature, susceptibility and power losses of SPIONs, which are determinant for hyperthermia applications [13], [20] can be easily tailored, [21]. Superparamagnetic scaffolds can be easily produced by embedding magnetic nanoparticles (MNPs) in porous traditional bone tissue engineering materials (e.g. ceramics, natural and synthetic polymers), as reported in [22]. Moreover, through a chemical doping of hydroxyapatite (Ha) with Fe ions, intrinsically magnetic and bioactive Fe/Ha nanoparticles and scaffolds (henceforth MHA) were synthetized [23]. As a matter of fact, different MNP can show completely different responses. For example, MHA demonstrated a temperature increase of 40 °C in 1 min, whereas the polycaprolactone (PCL) scaffold loaded with magnetite (Fe₃O₄) reached a 9 °C increase in 5 min, both using an external MF of 30 mT at 293 kHz [22], moreover using different exposure conditions.

All the studies on magnetic scaffolds focus on the synthesis, characterization and proof-of-concept of such therapeutic biomaterials [24-29]. However, none of them has ever modeled completely the physical properties of the scaffolds to accurately define hyperthermia treatment, also including and simulating healthy and pathologic tissues. To start bridging the gap, this work deals with the numerical simulation of in vivo magnetic induction hyperthermia of Osteosarcoma and Fibrosarcoma tumor cells [1]-[5] in presence of the FeHa/PCl and MHA superparamagnetic scaffolds described in [13], [22], [23] when exposed to a time-varying MF. Aim of this work is to model scaffold and MNPs properties and to connect them with HT treatments of bone tumors. The final goal is to help choosing strength, frequency and waveform of external MF required to

perform an optimal treatment. In section II, SPIONs properties. In section III the bio-heat transfer model is presented, explained and discussed. In section IV details on geometry and parameters used in the simulations are TABLE I

PROPERTIES OF MAGNETIC SCAFFOLDS FOR BONE TUMOR HYPERTHERMIA

Magnetic Scaffold	FE-HA (MHA)	FE-HA/PCL
Magnetite Crystal radius (nm)	10	14
Magnetic Phase (vol %)	1.6	9.18
Saturation Magnetization (emu g ⁻¹)	0.95	6
Maximum Heating (°C)	40	10
Time (min)	1	5
Magnetic Flux Density Field Strength (mT)	30	30
Working Frequency (kHz)	293	293
Reference	[22],[23]	[22],[23]

summarized. The non-linear behavior of the heating phenomenon is taken into due account. The influence of physiological and pathological factors such as tumor stage and bone healing process are included in the presented analysis. In section V, optimized magnetic field parameter, temperature spatial distribution and temporal development in tumor region for the different materials analyzed therein are reported. Finally, in section VI some conclusions are drawn.

II. POWER DISSIPATION OF MAGNETIC BIOMATERIALS IN TISSUES

A. EM Model

SPIONs can be magnetized by an external magnetic field, and the strength of this magnetization is proportional to the (complex) magnetic susceptibility $\chi = \chi' - j\chi''$. For an alternating field at a frequency f, the magnetic volumetric power dissipation density is [30]:

$$P_M = \mu_0 \pi f H^2 \chi " \tag{1}$$

where $\mu_0 = 4\pi \cdot 10^{-7} \text{ Hm}^{-1}$ is the vacuum permeability. H in (1) is the magnetic field at the SPION location, which must be computed, given the external magnetic flux density. Maxwell's equations are therefore solved in the frequency domain to determine actual magnetic field, H, acting on the SPIONs. Moreover, since dielectric losses may contribute to heating phenomena [31], the electric field E, due to the Faraday Law, is determined to compute the associated dissipated power (P_E) and hence quantify the overall electromagnetic losses (Q_{EM}) to determine the temperature increase. The MNPs, with radius and saturation magnetizations given in Tab. I, are assumed to be uniformly dispersed in either a hydroxyapatite or a PCL scaffold described in [22], with radius r_s=0.5mm (Fig. 2). The commercial Finite Element Method (FEM) software COMSOL Multyphisics (COMSOL inc., Burlington, MA),

TABLE II
DIELECTRIC PROPERTIES OF SCAFFOLDS MATERIALS AND TISSUES

	ε(293 kHz)	σ(293 kHz), Sm ⁻¹	ε(409 kHz)	σ(409 kHz), Sm ⁻¹	References
На	12.5-j·0.5	2.1·10 ⁻³	11.98-j·0.47	2.5·10 ⁻³	[54]
PCL	2.2-j·0.125	10 ⁻⁵	2.15-j·0.120	$0.3 \cdot 10^{-4}$	[57]
Fracture (inflamed)	3580-j·33451	0.545	3360-j·24491	0.557	derived from [58]
Fracture (ischemic)	1321.5-j·12030	0.196	1241.5-j·8494	0.2	derived from [58]
Bone	192-j·1313.5	0.0214	182-j·959	2.18e-2	[60]
Bone tumors (FS and OS)	8000-j·17186	0.280	3600-j·11432	0.26	[59]

Magnetic Fields module is used to solve Maxwell equations and simulate superparamagnetic scaffold response.

In a low viscosity regime, the frequency response of a dispersion of MNPs has been described in terms of a Debye model and the Rosensweig theory justifies the behavior of ferrofluids [30]. Whereas, in a significant viscosity regime, as suggested by Tampieri et al. [23] and measured by Hergt et al. [31], the sticky medium inhibits dissipation mechanisms, thus strongly influences the power loss of a MNPs ensemble. In a highly viscous or solid matrix the resonance vanishes, since Brownian relaxation cannot occur [31], [32]. Hence, only Néel mechanism should be considered to explain heating efficiency of scaffolds magnetic phase [31], [37]. In this configuration the interparticle strong dipolar interactions limit the MNPs relaxation dynamic [33]. Therefore, since the Debve response vanishes and long-range interactions between particles become important [31]-[34], magnetic scaffolds experimental heating curves from Bañobre-López et al. [20] and Tampieri et al. [23] should be described using a Cole-Cole model for magnetic susceptibility, $\chi(\omega)$, as [34], [35]:

$$\chi(\omega) = \chi' - j\chi'' = \frac{\chi_0}{1 + (j\omega\tau)^{1-\gamma}}$$
 (2)

where ω is the angular frequency. and γ is the so-called Cole-Cole exponent, which describes the broadness of the relaxation time distribution. Fitting the data from [31], γ was previously found to be 0.750 [34]. Therefore, power losses evaluated with Cole-Cole correction can justify magnetic scaffolds heating curves reported by Bañobre-López et al. [22] and Tampieri et al. [23]. The characteristic Néel relaxation time τ in Eq. (2) can be computed as [30]:

$$\tau = \frac{\sqrt{\pi}}{2} \beta \tau_0 \frac{e^{\beta}}{\sqrt{\beta^3}} \tag{3}$$

where $\beta = K_a V_m/k_B T$, being K_a the magnetic anisotropy energy in Jm⁻³, V_m the magnetic nanoparticle volume in m³, k_B the Boltzmann constant in JK⁻¹ and T the system temperature in K. The constant τ_0 can be evaluated according to [36] as:

$$\tau_0 = \frac{\pi}{4} \frac{\left(M_s(0)V_m\right)}{K_a \gamma_e} \left[\frac{1}{\eta_f} + \eta_f \left(\frac{M_s(T)}{M_s(0)}\right)^2 \right] \sqrt{\frac{k_B T}{K_a}} \left(1 + \frac{k_B T}{K_a}\right) \tag{4}$$

where γ_e is the electron gyromagnetic ratio in rad s⁻¹T⁻¹ and $\eta_f = \eta \gamma_e M_s(0)$, being η the damping factor, equal to 0.25

[36]. The values of $M_s(T)$ were derived from the curves reported in [23]. Finally, χ_0 is the equilibrium susceptibility and it is calculated as suggested by [37]:

$$\chi_0 = \frac{\mu_0 \varphi V_m M_s^2}{3k_B T} \tag{5}$$

where φ is the volume ratio of the magnetic phase in the scaffold. The power dissipated by magnetic scaffolds is a function of the frequency and intensity of the applied magnetic field, which can be tuned, tailored and remotely controlled [31], [34], [35], [38]. Moreover, power losses are strictly related to magnetic phase in bone scaffold, to particle radius and blocking temperature of the sample [19], [23], [35]. On the other hand, dissipation depends also on the magnetic susceptibility of the sample (scaffold [32], [34]). This susceptibility has a strong temperature dependence, as shown in Eq. (3) to (5), and therefore the heating process is a non-linear one. Therefore, the Maxwell and the heat equation must be solved using a coupled multiphysics approach [39]. The magnetic field frequency of interest is quite low, and therefore a coupled resolution scheme is directly implemented in COMSOL Multiphysics using Magnetic Fields and Heat Transfer Modules.

B. Numerical evaluation of magnetic material properties

As explicitly shown by eq. (5), the larger the saturation magnetization or the magnetic phase in scaffold volume, the stronger the initial or equilibrium susceptibility χ_0 . The imaginary part of $\chi(\omega)$ depends in turn from χ_0 and hence do power losses in magnetic implants. This argument agrees with experimental data for both MHA and FeHa/PCL [22] (see Tab. III). However, χ₀ inversely depends on temperature, since the energy barrier increases with T and the material become less magnetizable [36]-[38], [40]. In Fig. 1 the susceptibility dependence on temperature is shown for MHA. On the other hand, the characteristic Néel relaxation time strongly influences the power dissipated in magnetic scaffolds through Eqs. (3) and (4). Exploiting magnetization curve vs. temperature from [23], τ_0 and τ_N are depicted in Fig. 1. These results agree qualitatively with [41]. Merging these findings, it is possible to infer that P_M decreases with temperature since χ '' slightly diminishes, as shown in Fig. 1. Heating would therefore tend to saturate.

To assess the use of Eq. (2), the Debye model (imposing γ =0 in Eq. (2)) and the Cole-Cole model were employed in the case of bone alone, to compute the power dissipated and the final temperature increase in response to a constant magnetic flux density field (30 mT, 293 kHz [22], 60 min). Both $P_{\rm M}$ and temperature calculated with Debye are two

orders of magnitude lower than Cole-Cole ones and do not agree with experimental findings of [22]. Therefore, as hunched from [31] and suggested by [33], [34], the Cole-Cole model can be assumed as an effective model for the behavior of the materials in Tab. III.

III. BIO-HEAT TRANSFER MODEL

The temperature increase of tissues under the effect of an "exogenous" power source Q_{EM} can be determined by the Pennes bio-heat equation [42], [43]:

$$\rho C_{p} \frac{\partial T}{\partial t} = k \nabla^{2} T + \left[\rho_{b} C_{p,b} \omega_{b} \left(T_{a} - T \right) \right] + Q_{M} + Q_{EM}$$
(6)

where ρ is the density in g m⁻³, C_p is the specific heat capacity in Jkg⁻¹K⁻¹, k is the thermal conductivity in Wm⁻²K⁻¹. The term in brackets represents the capillary perfusion, i.e. blood thermal contribution which acts to equilibrate surrounding tissues with arterial temperature T_a=37 °C (310.15 K). The perfusion of normal and tumoral tissue was included in the model by a piecewise

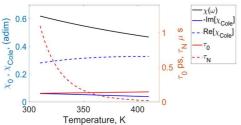


Fig. 1. Equilibrium magnetic susceptibility ($\chi 0$), real and imaginary part of susceptibility according to Cole-Cole model [38] as a function of temperature (adim). The external magnetic flux density field is set to 1 mT and 293 kHz. Temperature influence on constant $\tau 0$ according to [32] and on Néel relaxation time is drafted. Magnetic power losses as a function of temperature can be quantified with the presented model.

approximation to reproduce the temperature-dependence of chart.7 at page 4726s of [44] and Fig.6 at page 512 of [45]. The form of Eq. (6) ensures a 1.38% relative error with respect to in vivo studies [42], [43], [46]. There are two source terms in (6). One is the metabolic heat Q_M , while the term Q_{EM} is, in our case, the power dissipated due to dielectric and magnetic losses, and it is computed from electric and magnetic field assuming scaffolds and tissues as volumetric heat sources. Eq. (6) is subject to zero flux boundary condition at bone boundary, i.e. $-\hat{n} \cdot (k\nabla T) = 0$

. The initial temperature T_0 is assumed equal to 310.15 K for all tissues, except for the inflamed fracture (308.15 K) [47]. The fraction of damaged necrotic tissue has been evaluated considering the definition given by Sapareto and Dewey [48], [49], normalized to the damage time t_D . Both OS and FS tumors, and the fracture are considered damaged if temperature is kept above 43 °C (316.15 K) for 60 min [9]-[12], [17], [22], [25]. Of course, also healthy bone tissue is heated, but by conduction from the tumor. Therefore, it is necessary to design the hyperthermia treatment in an effective way while preventing the necrosis of healthy bone tissue, which occurs if temperature increases up to 47 °C

(320.15 K) for more than 1 min, as reported in [25], [50], [51].

IV. SIMULATION

A. Geometry

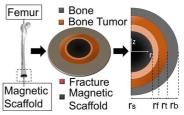


Fig. 2. The case of a scaffold implanted after long bone tumor surgical treatment is analyzed. A transverse section of bone, residual cancer tissue, fracture gap and scaffolds are considered, with radius r_b , r_t , r_l , r_s respectively. Given a spherical scaffold, system geometry is 2D axial-symmetric with r- and z-axes origin set in scaffold center.

OS and FS affect mainly long bones extremities [1]-[4]. Therefore, in this paper, the case of a magnetic scaffold implanted in distal femur after surgical intervention is analyzed, as depicted in Fig. 2. Since the aim of this work is to evaluate the main determinants for in situ hyperthermia treatment of bone neoplasms through magnetic scaffolds, the problem is simplified considering a circular 2D axial-symmetric geometry (Fig. 2).

Therefore, a very narrow volume of tissue around the implant is considered [15], [16], [20], [34], [52]. A homogenous uniform external magnetic flux density field B₀ is assumed applied along the z-axis. The origin of the system is assumed centered on the scaffold. The orthopaedic implant has a radius of 5 mm [13], [15], [34], [35], [52]. Near the scaffold, there is a region where new bone forms to heal surgical fracture [53]-[54]. Healing process starts with hematoma and local ischemia (first 7 days), and ends with an inflammatory phase (up to 14 days), which leads to callus formation [53]. For this reason, a small fracture with radius varying in the range $0.1 \div 0.5$ mm [54] has been considered, as in Fig. 2. The tumor region is modeled as a uniform tissue spherical area of radius r_t. The bone tumor volume is assumed to be reduced after surgical intervention [1]-[4], [11], [12]. The tumor radius has been varied from 0.1 mm to 0.5 mm.

B. Simulation Parameters

The relative magnetic permeability of the fracture, the tumor and the bone is assumed equal to one. As stated in Section II.A, also the dielectric losses are needed to compute the proper temperature distribution under the external MF. In order to model frequency response of both dielectric permittivity and conductivity of hydroxyapatite, data and relations from [55]-[56] were employed. For PCL it is possible to refer to [57]-[58] for its dielectric properties. The dielectric permittivity of the fracture gap was estimated employing the mixing model used in [59]. In fact, it is possible to discriminate between the ischemic and inflamed phase by weighing bone permittivity with blood

one. Whereas, tumor dielectric properties are derived from data of a 15 days tumor staging by the work of Swarup et al. [60]. Dielectric permittivity and electrical conductivity of all materials are reported in Tab. II for the frequencies of interest. Dielectric properties are assumed to vary linearly with temperature (with a variation of 3% °C⁻¹) as suggested by Rossman et al. [61].

Some considerations on thermal proprieties of the involved materials are also in order. The Cp (except for blood) and all the k have been assumed linear with temperature, with a 1 °C increase of 0.5% and 0.33% respectively [61], [62]. Cp for blood has been assumed linear with a 1 °C reduction of 1% [61]. Bone properties are taken from the IT'IS database [63]. For the two stages, fracture properties are derived from [64]. Regarded histological section presented in [1]-[3], bone tumor cells can be assumed uniformly dispersed in bone matrix [62], hence the uniformity assumption is justified. The cases of OS and FS are both considered. Highly vascularized OS is represented with a perfusion rate about 250 times higher than fibrosarcoma, as derived from [45]. On the other hand, to the best of the authors' knowledge, no data about thermal parameters are available for any of the studied bone tumor. Tumor thermal conductivity and heat capacity has been therefore assumed [46] to be equal to those of bone [63], since from histological sections found in [2]-[5] both OS and FS cells are regarded as a uniform dispersion in bone matrix. The metabolic heat source term, Q_M , is set to 57240 Wm⁻³ to account for the higher metabolic activity of cancer cells [46]. All thermal properties employed in the simulations are reported in Table III at the initial temperature. Employing data from Tab. I-III, the temperature influence on magnetic power losses is first investigated. Then, in absence of tumor (r_t=0 mm), heating for different radius of the fracture was assessed for both MHA and FeHa/PCL in presence of an uniform external magnetic field of 1 mT and 293 kHz. Finally, considering a small fracture ($r_f = 0.1$ mm), Fibrosarcoma and Osteosarcoma tumors volume was varied, and magnetic field strength, frequency and waveform were tailored to obtain the maximum damage of cancer cells (see summary table in the supplementary material). This will be available at http://ieeexplore.ieee.org.

V. RESULTS

The working frequencies and the order of magnitude of the external magnetic field have been chosen equal to those of [22] and [25]. The evaluated spatial temperature patterns are qualitatively similar in each case analyzed, therefore

only the ΔT distribution for OS after 85 min is presented in Fig. 3. A time-lapse animation of the heating in the case of MHA and both FS and OS with 0.5 mm radius is however given in the supplementary material. This will be available at http://ieeexplore.ieee.org. Being the external field homogeneous, and since the scaffold is spherical, the resulting heating is uniform and homogeneous. Henceforth relevant attention is paid only to temporal variation of temperature in tissues [22], [50] and to damage analysis

The first test has been performed for $r_t=0$ (i.e. without a tumor), for different gap width (for $r_f = 0.1, 0.3, 0.5 \text{ mm}$) and status (inflamed and ischemic stages). As the actual heating does not take place as direct conduction from magnetic implant to the bone, the presence of the surgical fracture gap has been included to assess its influence. In Fig. 4 is reported the average temperature in the gap during the heating, for B₀=1 mT @ 293 kHz, in presence of the MHA scaffold. Using PCL scaffold produce analogous results and, hence, they are not reported herein. Fig.3 shows clearly that the external field required by the HT treatment must be evaluated taking into account the fracture, and its state. Otherwise, the required external field will be largely underestimated. In case of an ischemic fracture, the strength of external field should be increased with respect to the inflamed state to achieve the same desired temperature increase. Therefore, in situ hyperthermia treatment with magnetic scaffolds should be performed immediately after surgery. FS is a poorly vascularized type of bone tumor; therefore, it holds that $P_M \gg \rho_B C_{p,B} \omega_B (T_a - T)$, as observed during numerical experimentation. Hence, moderate magnetic field strength allow to keep the temperature above 43°C for 60 min in tumor [10] (Fig. 5a). To prevent healthy bone to overcome the safety limit (47°C for 1 min or more [25]) the field envelope cannot be constant, and must be properly designed. The envelope of Fig. 5a has been optimized to get an effective treatment. For PCL, a 17 mT field at 293 kHz is sufficient to completely disrupt bone tumor of any dimension, as presented in Fig. 5b. On the other hand, MHA, which has stronger saturation magnetization (Tab. I), requires 10 mT. The OS is a very active and perfused tumor [1], therefore stronger fields are required to achieve the therapeutic performance. A constant magnetic flux density (see Fig. 5c) applied for 85 min, can efficiently treat OS with MHA scaffolds. The required external field must, however, be increased for larger rt. For MHA the magnetic field strengths ranges from 20 mT up to 50 mT at 293 kHz. For the FeHa/PCL scaffold, the frequency was augmented to 409 kHz, as in [25]. Without

TABLE III THERMAL PROPERTIES AT 37°C OF SCAFFOLDS MATERIAL AND TISSUES

	$k \left(Wm^{\text{-}1}K^{\text{-}1}\right)$	$C_p (J kg^{\text{-}l} K^{\text{-}l})$	$Q_{\mathrm{M}}\left(Wm^{\text{-}3}\right)$	$\omega_{\mathrm{B}}\left(s^{\text{-l}}\right)$	References
На	1.3	700	/	/	[34]
PCL	1.75	1300	/	/	[34]
Fracture (inflamed)	0.558	2450	5262.5	6.95e-3	[46]
Fracture (ischemic)	0.1	2450	342.1	0.262e-3	[46]
Bone	0.32	1313	286.2÷57240	0.175e-3	[60]
FS	0.32	1313	286.2÷57240	2.4e-3	[60],[62]
OS	0.32	1313	286.2÷57240	0.595	[60],[62]

this frequency change, large OS could not be successfully treated, as shown in Figures 5c and 5d. It should also be noted that FeHa/PCL fails in treating OS with radius higher than 0.1 mm even using a field at 409 kHz, unless the field intensity is significantly larger than 50 mT [22], [25]. This finding implies that different MNPs, with a higher volume ratio, should be embedded in the scaffold, as well as; field strength and frequency ought to be increased to achieve the same performances of MHA. However, the product $f \cdot H$ has to be controlled, as dictated by Pankhurst et al. [33]. The biocompatibility of the implant is a major requirement for this application [20]. Despite these reasonable limitations, the presented simulation model can give a first-estimate of hyperthermia potential of any type of magnetic scaffold.

VI. CONCLUSION

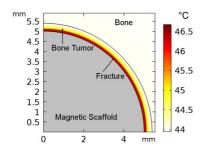


Fig. 3. Zoom of the 2D distribution of temperature in tissues. The case of a 0.3 mm OS is presented. A 28 mT and 293 kHz field is employed.

A coupled thermal-electromagnetic model for hyperthermia treatment of bone tumor using MNPs and magnetic scaffolds has been devised. This model takes into account

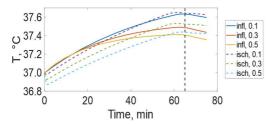


Fig. 4. Average temperature in fracture volume vs. time and for both the inflamed and ischemic case. The dashed black line indicates the time point at which the external RF magnetic field is turned off.

in an integrated way several important non-linear properties of magnetic materials, but also physiological and physical features of the in vivo hyperthermia treatment of bone tumors, which were only partly considered before. Particular care has been given to take into account the strong temperature dependence of nanoparticles magnetic susceptibility, which leads to non-linear effects during the treatment. An important finding is the relevance of the status of the surgical fracture gap on the tumor heating, which supports the use of hypertermia several days after surgical intervention. As a result, the temperature in tumour tissues can be evaluated with more accuracy, allowing a

more realistic design of the treatment. This has been found particularly important in FS treatment. As a matter of fact, the FS has a poor vascular structure, without an optimized design of an uniform external magnetic field, and therefore, can be easily overheated during the treatment, increasing the temperature of healthy bone tissues into the unsafe region. It is worth nothing that the presented model can be quite easily adapted to lively problems, such as the use of magnetic scaffolds in thermally-activated drug delivery.

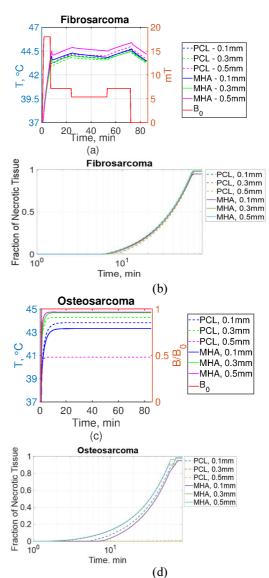


Fig. 5a. Average temperature increase in Fibrosarcoma region. 5b. Fraction of necrotic tissue for Fibrosarcoma (FS) in presence of an inflamed fracture. B₀=17 mT for FeHa/PCL, whereas 20 mT for MHA (at 293kHz, as in [22]). Exposure time is 85min. The optimal applied field can be modeled as a descending ramp since FS is a low perfused tumor. Both magnetic biomaterials are able to kill 100% residual cancer cells, even for r₁=0.5 mm. 5c. Average temperature increase in Osteosarcoma region. MHA scaffolds are able to keep temperature above the therapeutic threshold. FeHa/PCL succeed for small tumor region (r₁=0.1 mm) but is inadequate for tumors with radius higher than 0.3 mm. 5d. Fraction of necrotic tissue for Osteosarcoma (OS) in presence of an inflamed fracture. Exposure time is 85min.

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