Periodontal microbiota of Sardinian children: comparing 200-year-old samples to present-day ones

Germano Orrù1,2, Maria Paola Contu1, Eleonora Casula1, Cristina Demontis1,2, Cornelio Blus2, Serge Szmukler-Moncler2, Gabriele Serrelli3, Carla Maserati4, Giorgio Carlo Steri4, Vassilios Fanos5, Ferdinando Coghe3, Gloria Denotti2

1Molecular Biology Service MBS, AOU Cagliari, Cagliari, Italy
2Dental Section, AOU Cagliari, Cagliari, Italy
3Laboratory Medicine Service, AOU Cagliari, Cagliari, Italy
4Hygiene & Public Health Section, Cagliari, Italy
5Neonatal Intensive Care Unit, Neonatal Pathology and Neonatal Section, AOU Cagliari, Cagliari, Italy

Abstract

Introduction: The microorganisms of the human oral cavity include more than 700 species or phenotypes of bacteria. Some “diseases of civilization” are strictly correlated to changes in the microbiome following the food revolution that occurred after WWII. For that reason, a precise recognition of the microbiome profile before and after this period should be useful to determine the health-compatible model of microbiome. The aim of this study was to compare the microbiome profiles (number of total cells, and pathogen types) of dental samples obtained from two distinct groups of children, a 200-year-old retrieved one and a present one.

Methods: Two different groups of samples have been studied. The first group was a set of 50 recent subgingival plaque samples obtained from children of age 2-8 years, 14 males and 36 females. They were enrolled by the Department of Dental Disease Prevention (University of Cagliari, in Sardinia, Italy) during standard dental care procedures. None reported periodontal disease and none had been under antibiotic therapy during the previous 6 months. The second group was an old retrieved group that included 24 teeth from 6 different 6- to 8-year-old crania fragments; they were obtained from a 200-year-old charnel-house located in Villaputzu, a city close to Cagliari. Representative periodontal bacteria have been identified by a previously published real-time PCR procedure (Sokransky et al., 1998) in which P. gingivalis and T. forsythia (red complex), A.
actinomycetemcomitans (green complex) and F. nucleatum (orange complex) were detected. In addition, the title of each pathogen was expressed as a percentage of the total bacteria (biofilm) in the sample.

Results and discussion: The profile of periodontal microbiomes, between recent/ancient samples showed a significant difference relative to Sokransky’s red complex bacteria (p < 0.05). In all analyzed periodontal strains, the pathogenic bacteria P. gingivalis and T. forsythia showed the highest title in the recent group.

Conclusions: Our hypothesis is that the transfer of “commensal-pathogen” as an absolute number on the oral biofilm might be linked to the distinct alimentary habits of the two populations. Some diet rich in reducing agents, such as processed meat-based foods, might be able to increase the average number of pathogen anaerobic bacteria in the oral microbiota. The outcome would be an increase of the oral systemic diseases reported with these pathogens. Our data suggest that the ancient Sardinian population was able to control the pathogen oral anaerobic biofilm by some diet rich in antioxidant compounds. Further investigations are required to focus on the genetic profile and the health status of this ancient population but it appears that molecular microbiology might be considered as the “time machine” in oral biology.

Keywords

Subgingival plaque, microbiota, children, ancient population.

Corresponding author

Germano Orrù, Molecular Biology Service MBS, AOU Cagliari, Cagliari, Italy; email: gerorru@gmail.com.

How to cite


Introduction

The microrganisms of the human oral cavity include more than 700 species or phenotypes of bacteria. Some “diseases of civilization” are strictly correlated to changes in the microbiome following the food revolution that occurred after WWII. For that reason, a precise recognition of the microbiome profile before and after this period should be useful to determine the health-compatible model of microbiome. Molecular biology can be applied to ancient human findings in order to detect relevant genetic data in different fields of medicine. The use of dental tissues as the most representative source of ancient DNA has recently been described [1-3]. The dental structure is characterized as a complex architecture; it is able to incorporate and preserve the nucleic acids from both the host and its ancient oral microbiota. Analysing these samples with molecular biology tools may provide a powerful method for studying ancient human habits, epidemiological diseases, human migrations and genetic drift [3]. The aim of this study was to investigate possible differences in the subgingival microbiota on the hard tissues of dental samples obtained from two distinct groups of children: a retrieved 200-year-old group and a present-day one.

Methods

The present-day group included a set of 50 recent subgingival plaque samples obtained from 14 boys and 36 girls aged between 2 and 8 years. They were enrolled by the Department of Dental Disease Prevention (University of Cagliari, in Sardinia, Italy) during standard dental care procedures. All parents signed an informed consent form before the children took part in the microbiological analysis. None reported periodontal disease and none of the children had undergone antibiotic therapy during the previous 6 months. Their health status was recorded and the subgingival plaque was immediately stored at -20°C following a previously described method [4, 5]. The retrieved ancient group included 24 teeth from 6 different 6- to 8-year-old crania fragments obtained from the 200-year-old charnel-house located in Villaputzu, a small town close to Cagliari (Fig. 1). The location of these bones appeared well maintained and any sign of humidity was recorded during the sampling.

These samples were gently cleaned with a cotton swab to remove the superficial soil. For each cranium, bones and teeth status was recorded; signs of periodontal disease or caries were also recorded. The teeth were then extracted from the dental alveoli of the respective jaws
Sample pretreatment and DNA extraction were performed following the procedures described by Bolnick et al. and Orrù et al. [4, 5]. Representative periodontal bacteria described by Socransky et al. [6] were identified by a previously published real-time PCR procedure [5] in which \textit{P. gingivalis} and \textit{T. forsythia} (red complex) [7], \textit{A. actinomycetemcomitans} (green complex) [5] and \textit{F. nucleatum} (orange complex) [8] were detected. In addition, the titer of each pathogen was expressed as a percentage of the co-respective total bacteria (biofilm) in the sample detected by a set of universal primers designed on \textit{rrs} sequence of \textit{E. coli} (Fig. 3); this approach prevented any possible errors due to DNA degradation of the ancient samples during their time in the ossuary, i.e. errors in the accuracy of the real-time PCR procedure [9].

The real-time PCR reaction was performed with a LightCycler® instrument (Roche Diagnostics, Mannheim, Germany) and a SYBR® Premix Ex Taq™ Kit (TaKaRa-Clontech®), according to the manufacturer’s instructions [5].

**Results and discussion**

Observation showed that the 200-year-old group displayed different dental/bone diseases. These were extensive caries (21.4%), tooth wear (21%), dental calculus (7.1%), but no bone loss. These conditions are probably due to the lack of oral care and insufficient hygiene conditions. As described by Weyrich et al., the use of dental calculus (calcified tartar or plaque) was identified as a relevant recovery source of ancient DNA [10]. Indeed, subgingival and supragingival plaque is rich in calcium phosphates and silicates; it calcifies \textit{in situ} during the host life while forming layered fossilized concretions known as dental calculus. These concretions lock other kinds of material located in the oral cavity and preserve it from taphonomic and environmental alterations. It thus remains intact for long periods of time, indeed over millennia. The same formation is present in recent samples (dental plaque) [1, 11, 12]. According to this approach, the profile of periodontal microbiomes showed a significant difference between recent/ancient samples relative to Socransky’s red complex bacteria (p < 0.05), while no statistical significance was observed for the other examined periodontal pathogens (p > 0.05).

In all analyzed periodontal strains, the pathogenic bacteria \textit{P. gingivalis} and \textit{T. forsythia} showed the most important differences compared to other periodontal pathogens (Fig. 3). These strains are associated with severe infections in humans, such as periodontal disease, endocarditis, rheumatoid arthritis and pre-term birth [13, 14] and these human infections are often related to modern civilization [15]. In addition, a possible physiological activity for some periodontal bacteria, such as \textit{F. nucleatum}, in the flavor and taste perception of vegetables, has been suggested [16]. Our hypothesis is that the transfer of “pathogen/commensal” as percentage amount in the oral biofilm might be linked to the distinct alimentary habits of the two populations. A diet rich in reducing agents, such as processed meat-based foods, might be able to increase the average number of anaerobic bacteria in the oral microbiota. The outcome would be an increase in the oral systemic diseases reported with these pathogens in the last decades. In this context our data suggest that the ancient Sardinian young population was able to control the pathogens in the oral anaerobic biofilm by a diet rich in antioxidant compounds.
Conclusions

This preliminary study showed a difference in red complex bacteria titer in children that lived 200 years ago when compared to a present-day group. Further investigations are required to focus on the genetic profile and the health status of this ancient population but it appears that molecular microbiology might be considered as the “time machine” in oral biology.

Acknowledgements

The support of the Villaputzu administrative authorities during sample collection was highly appreciated. We also thank Hygiene & Public Health Department of the University of Cagliari for approving this study.
Declaration of interest

The Authors declare that there is no conflict of interest.

References